

## REVIEW

[View Article Online](#)  
[View Journal](#) | [View Issue](#)

Cite this: *Polym. Chem.*, 2021, **12**, 6198

Received 30th August 2021,  
Accepted 12th October 2021

DOI: 10.1039/d1py01172c

[rsc.li/polymers](https://rsc.li/polymers)

## Nanoengineering with RAFT polymers: from nanocomposite design to applications

Wentao Peng,<sup>id</sup> Yingying Cai,<sup>id</sup> Luise Fanslau and Philipp Vana<sup>id</sup> \*

Reversible addition–fragmentation chain-transfer (RAFT) polymerization is a powerful tool for the precise formation of macromolecular building blocks that can be used for the construction of well-defined nanocomposites. Especially when combining RAFT polymers with uniform inorganic/metallic nanoparticles, a vast variety of complex nanocomposites become available that may find applications especially in the field of life sciences, e.g., bio-imaging, drug delivery or cancer-therapy, but also in areas such as energy conversion or catalysis. RAFT polymerization not only provides the possibility to control the size and the macromolecular architecture of the polymer building blocks, but inherently delivers highly functional end-groups that can often directly be employed as linker-sites for installing the polymeric components into the final nanocomposites. This review describes recent advances in this vivid field and concentrates on innovations in the fabrication method and design strategies for polymer/inorganic nanohybrids. The methodology of synthesizing RAFT polymer with the aim of surface functionalization and the design options for anchoring RAFT polymer on surfaces are covered. A series of core–shell nanostructures with the focus on novel functionalities brought by RAFT polymer brushes will be reviewed with focus on the detailed macromolecular design for each specific application's scenario. Examples of ordered nano-assemblies using RAFT polymer linkers will be reviewed in order to demonstrate the advantages of RAFT polymer for introducing different morphologies, interactions and chirality to the final functional nanostructures.

## 1. Introduction

Since its debut in 1998,<sup>1</sup> reversible addition–fragmentation chain-transfer (RAFT) polymerization has quickly become a fantastic tool for designing and fabricating polymers with

Institut für Physikalische Chemie, Georg-August-Universität Göttingen,  
Tammannstrasse 6, 37077 Göttingen, Germany. E-mail: [pvana@uni-goettingen.de](mailto:pvana@uni-goettingen.de)



Wentao Peng

Wentao Peng received his B. Sc. in Chemistry from the University of Göttingen in Germany in 2013, and obtained his Ph. D. in Chemistry under the supervision of Prof. Philipp Vana in 2020. He is currently a Postdoctoral Associate in Prof. Vana's group with a focus on the self-assembly of nanomaterials (metal and silica-based nanoparticles) using functional macromolecular linkers.



Yingying Cai

Yingying Cai received her B. Sc. in Chemistry in 2015 before completing her M. Sc. in 2017 from the University of Göttingen. Currently, she is a doctoral student in the group of Prof. Philipp Vana at the University of Göttingen. Her research interest focuses on developing new polymer-inorganic nanodevices for photon luminary application using silica coating and surface-initiated RAFT polymerization. She is also developing new approaches for colloidal self-assembly of silica and noble metal nanoparticles using functional polymer.



special functions. The precision engineering using RAFT polymerization includes well-defined polymer size,<sup>2</sup> versatile end-group design,<sup>3,4</sup> convenient access for block copolymer (BCP) synthesis,<sup>5–7</sup> and flexible topological design.<sup>8–12</sup> These options can be combined simultaneously to give RAFT polymers the ability for a wide range of applications including e.g., stimuli-responsiveness,<sup>9,13–15</sup> therapeutic-delivery,<sup>16–20</sup> bio-imaging and diagnosis,<sup>17,21,22</sup> optoelectronics,<sup>23</sup> polymerization induced self-assembly,<sup>24–27</sup> interface engineering,<sup>28</sup> smart nanoreactors,<sup>29</sup> structured templating,<sup>30</sup> bioseparation,<sup>31</sup> and photo-curing 3D printing.<sup>32,33</sup>

The rise of RAFT polymers has revolutionarily changed the game of surface engineering of nanoparticles (NPs). A great variety of anchoring strategies was developed to immobilize RAFT polymer brushes onto a plethora of nanosurfaces.<sup>28,34</sup> The polymer-functionalized inorganic nanomaterials carrying a combined talent from both functional polymers and inorganic nanomaterials have received extensive product proliferation to encompass an extraordinary array of applications especially in the biomedical field.<sup>35–37</sup>

The colloidal self-assembly of nanomaterials has stimulated enormous research interest in the last decade.<sup>38–43</sup> In many cases, the construction of these well-defined nanostructures all share the same goal: creating synergic interaction between nanocomponents. On this topic, RAFT polymer has the auspicious talent as a linker for guiding the colloidal self-assembly of nanomaterials using its topological and anchoring design. We will see how RAFT polymer engineering can create the synergy between nanocomponents and dynamically changes the colloidal arrangement of each nanocomponent for modern applications.

The current review focus on the recent advances in both fabrication methods and design strategies for polymer/inorganic nanohybrids for multifunctional applications (illustrated in Fig. 1). We will start with the methodology of synthesizing RAFT polymers with the aim of surface functionalization. We will go through the important design options for RAFT polymers including a detailed discussion on anchoring strategies for diverse nanomaterials. Then we will review a series of classic core-shell nanostructures with the focus on



**Luise Fanslau**

*Luise Fanslau received her B. Sc. degree in Molecular Biotechnology from Heidelberg University, Germany, where she was working on photoswitchable DNA in the group of Prof. Dr Andres Jäschke. After working one year at GlaxoSmithKline Biologicals in Dresden, Germany, she pursued an MPhil Biotechnology degree at the University of Cambridge, UK, researching on novel DNA vaccine delivery nanoparticles in*

*the group of Dr Ljiljana Fruk and graduated with distinction. Currently, she is a doctoral student in the group of Prof. Philipp Vana at the University of Göttingen, working on RAFT polymer nanocomposites. She receives a Kekulé scholarship of the Fonds der Chemischen Industrie.*

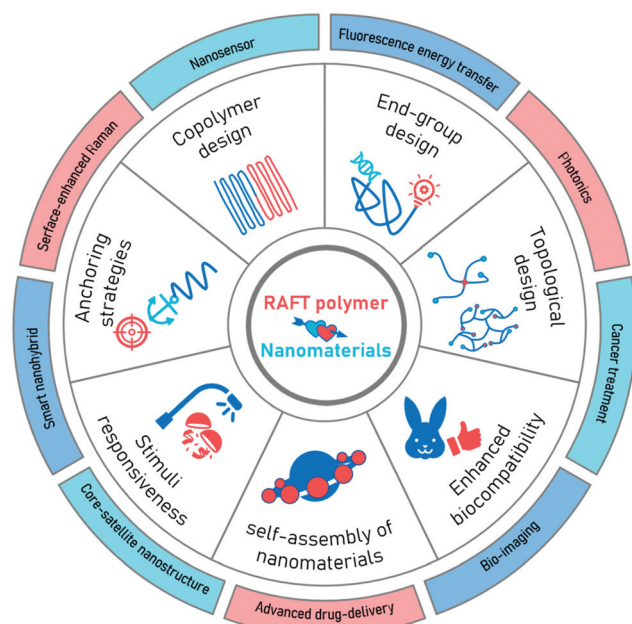


**Philipp Vana**

*Philipp Vana studied Chemistry at the University of Vienna, Austria, where he obtained a Master of Natural Sciences in 1996 and a Doctor of Natural Sciences in 1999 after investigating chain-length dependent termination kinetics in radical polymerisation in Professor Oskar-Friedrich Olaj's group. Parallel to his PhD, he studied economics and law and obtained a diploma from the Technical University Vienna and a Master*

*of Business Administration from the Danube-University Krems, Austria. Between 2001 and 2003 he was a Schrödinger-Fellow of the Austrian Science Fund at the Centre for Advanced Macromolecular Design (CAMD) at the University of New South Wales, Australia, where he work in the group of Professor Tom Davis in the field of RAFT polymerisation kinetics. In 2003, he established an independent research group at the University of Göttingen, Germany, specifically focusing on macromolecular design and functional polymer materials. In 2005 he became a Fellow of the Japan Society for the Promotion of Science at Kyoto University in Professor Takeshi Fukudas group, where he moved into the field of polymer brushes and surface modifications. In 2008 he was granted the prestigious Heisenberg-Professorship of the German Research Foundation (DFG) at the University of Göttingen, where he finally became a Full Professor for Macromolecular Chemistry in 2010 after declining offers from Leipzig University and Duisburg-Essen University. Philipp has published more than 140 original research papers in addition to several book chapters, reviews and patents.*





**Fig. 1** Schematic illustration of design options for the surface modification of inorganic NPs with RAFT polymer and the fields of applications for the hybrid nanomaterial.

the novel functionalities brought by the polymer brush. These functional nanostructures cover a wide range of applications including bioimaging, drug-delivery, cancer treatment, fluorescence energy transfer based nanosensors and hybrid responsiveness. We will focus on the detailed macromolecular design for each specific application scenario. Thereafter, examples of ordered nanoassemblies using the RAFT polymer linker will be reviewed to showcase the advantages brought by the design flexibility of RAFT polymers. This part also highlights the possibility to introduce novel morphologies, interactions and chirality to the nanostructure using the sophisticated design of the RAFT polymer shells. The review will be concluded by discussing the future challenges and opportunities for surface-bound RAFT polymers.

It is worth mentioning here that the offering of RAFT polymerization in nanoengineering cannot be fully covered within the scope of this review. Especially polymerization-induced self-assembly (PISA) has been a surge of interest in the fabrication of block copolymer nano-objects with controlled morphology, *e.g.*, nano-sphere, -worm, and -vesicles. In this research area, a great number of the literatures are based on RAFT technique. The readers interested in this topic can refer to more elaborate publications<sup>24,44–50</sup> on PISA technique.

## 2. Construction options for RAFT polymer towards nanosurface

RAFT polymerization is a type of reversible deactivation radical polymerization (RDRP) developed by the Australian

Commonwealth Scientific and Industrial Research Organization (CSIRO).<sup>1</sup> This polymerization technique confers the favored living characteristics to radical polymerization. It combines many useful features of radical polymerization, such as ease and cost-effectiveness of synthesis, compatibility with many different monomers, and broad reaction conditions.<sup>23,51</sup> The control of the polymerization kinetic enables precise design and engineering of functional polymers which induced an explosion of innovation in polymer science over the last decades. Compared with other RDRP techniques, the major competitor for RAFT is the atom transfer radical polymerization (ATRP). Both of them offer a very high degree of design and control on polymer architecture and flexible end-group modification. The major advantage of RAFT polymerization is the straight-forward and insensitive reaction condition: in ATRP, a copper-based complex must be usually added for the polymerization and removed from the product. However, compared with other RDRP techniques, the color of RAFT moieties in the polymer product could be an issue for many applications unless they are removed by one extra reaction step. Notably, for the long-chain polymer ( $n > 500$ ), ATRP often offers better control than the RAFT counterpart. A selection of recent work and reviews is included here on the comparison of RDRP polymerization techniques.<sup>52–54</sup>

A series of grafting strategies have been developed to immobilize RAFT polymers onto the surface of nanomaterials. Understanding the surface chemistry between RAFT polymer and the specific surface of NPs is crucial for the effective polymer functionalization.

### 2.1. RAFT polymerization

The mechanism of RAFT polymerization is illustrated in Fig. 2A. After a radical initiation event, the propagating radical chains react with the RAFT agent, which usually consists of a thiocarbonylthio group. This leads to a reversible degenerate chain transfer, rendering the former radical chain inactive (“dormant”) and releasing the previously dormant chain as radical species able to propagate. The RAFT equilibrium offers a rapid exchange between the active propagating radical and the dormant radicals, while the majority of the chains are kept in dormant form. As result, all polymer chains have equal time and possibility to propagate. This controlled kinetic gives the polymer product a narrow molecular weight distribution, *i.e.*, a similar degree of polymerization. Practically, the amount of radical initiator must be carefully considered to maintain the favored RAFT kinetic, since the concentration of active species in the reaction mixture depends on the concentration of the initiator.<sup>2</sup>

Owing to the clear mechanism of polymer growth in the controlled radical polymerization, RAFT technique allows for a high degree of control over resulting molecular weight distribution, copolymer composition, and macromolecular architecture.<sup>23</sup> Due to the living character of the polymerization, BCPs can be easily synthesized. Also, the end-group functionality is preserved, providing a reactive handle for further modification





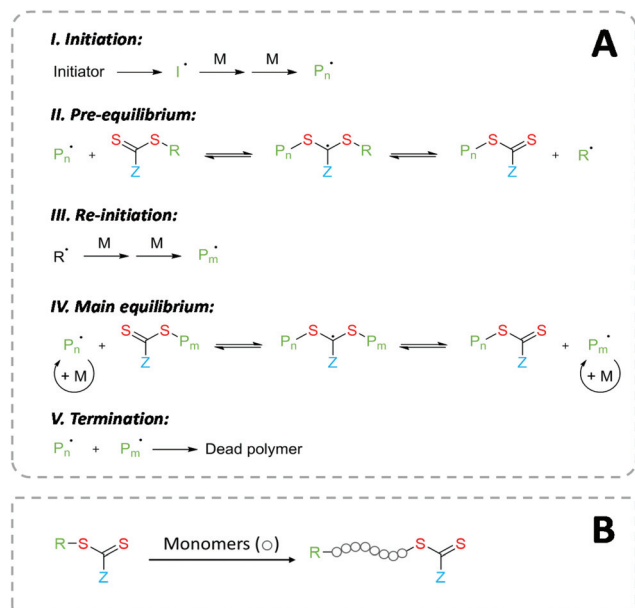


Fig. 2 (A) Proposed Mechanisms of RAFT polymerization. (B) Overall RAFT polymerization process.

or serving directly as anchoring moieties for surface functionalization.<sup>23</sup>

Since the invention of RAFT polymerization, extensive research efforts were initiated towards developing versatile RAFT agents which are suitable for a great number of monomer and reaction conditions, including aerobic, biological, or continuous-flow conditions.<sup>23,55,56</sup> Boosted by growing industry demands, the range of reported RAFT agents as well as their commercial availability is rapidly expanding.<sup>23,57,58</sup> Development of the initiation process for RAFT polymerization also has been developed rapidly, amongst others *via* light, enzymes, electric fields, or ultra-sonication.<sup>56</sup>

Over the years, the precision, versatility, and simplicity in both design and synthesis provided by RAFT polymerization quickly found application in various fields.<sup>23</sup> The offering of RAFT polymers has quickly gone beyond the conventional branches for polymers. For further information on RAFT polymerization, *e.g.*, the match of RAFT polymer with its suitable monomer classes, the recent development of new RAFT agents, the process of polymerization, and more studies on the mechanism and kinetics, the reader is referred to more elaborate publications on those specific topics.<sup>2,23,51,56,57,59</sup>

## 2.2. End group modification

Introducing functional groups at specific positions in the polymer chain is the most basic and practical approach for extending the functionality of polymers. RAFT polymerization offers flexible options for positioning the RAFT group after polymerization: both in-chain and at the chain ends. The chain-end design has the advantage of further chemical modification without considering the degeneration of the polymer chain. Fig. 2B shows the position of the RAFT agent after

polymerization: the RAFT moiety part (Z group) retains at their  $\omega$ -end, while the R-group functionality stays at the  $\alpha$ -end.<sup>60</sup> It is worth mentioning that besides the favored RAFT polymer with end-group functionalities, polymer species not possessing both of these end groups always exist in the product. This “dead” polymer originates from irreversible termination of propagating polymer radicals, which also border the molecular weight distribution since the terminating product has twice the molecular weight of the propagating radical and this process occurs during the entire polymerization period. The formation of the termination product can be controlled by reducing the amount of initiator.<sup>59</sup> A low initiator concentration keeps the ratio of active/dormant species healthy, *i.e.*, maintains the living character of the reaction.<sup>51</sup>

We have seen that using RAFT polymerization allows a simultaneous end-group design for both chain ends of the polymer. However, one needs to consider that both Z- and R-group of the chain transfer agent (CTA) must be designed not only in terms of required end group functionality, but also the reactivity of the CTA during polymerization. R-Groups need to be a good leaving group and effective at reinitiating polymerization as free radicals, whereas Z-groups modulate the rate of addition and fragmentation and need to provide stability to the intermediate radical species.<sup>3,59</sup> On this topic, a great library of RAFT agents with distinct end group functionalities, such as carboxyls, hydroxyls, protected amines, clickable groups, and fluorophores, their syntheses, and compatibility with monomers has been reported in the literature.<sup>51,61–64</sup>

Another widely used approach bypassing the limitation of the design option for RAFT agents is the modification of the  $\omega$ -group after polymerization: the thiocarbonylthio moiety provides a suitable and chemically versatile handle for introducing desired functional moieties, as shown in Fig. 3. On this topic, the chemical modification options for the RAFT group are reviewed in detail in the literature.<sup>3,4,60</sup>

Employing a reaction with nucleophiles or ionic reducing agents, the thiocarbonylthio moiety can be converted into a thiol group. Prominent examples include the reaction with borohydrides,<sup>65,66</sup> aminolysis,<sup>67,68</sup> or hydroxyls.<sup>60</sup> Owing to the specific color of RAFT moieties, this process can be optically monitored. The resulting thiol end group can be directly used for binding to various nanosurfaces (see section 2.4) or further

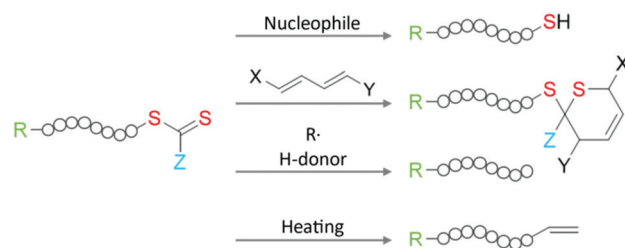


Fig. 3 Schematic illustration of commonly used chemical pathways for RAFT group modification.



reactions including nucleophilic substitution at carbons or sulfurs, nucleophilic addition to activated double bonds, radical addition with alkenes and alkynes, or oxidation to disulfides.<sup>69,70</sup> It is worth mentioning here that the oxidation of thiols will yield a disulfide bond that links two thiolated polymers together and doubles the molecular weight. For multi-thiolated polymers, the oxidation will cause a covalently cross-linked polymer network. Special attention must be given to the synthesis and storage of thiolated polymer to avoid oxidation (inert atmosphere or low pH condition).<sup>67</sup> Alternatively, the thiocarbonylthio moiety can be used as a dienophile in hetero Diels–Alder reactions with dienes.<sup>60</sup> The reaction of the thiocarbonylthio group with oxidizing agents such as hydrogen peroxide, air, or ozone results in *e.g.*, hydroxyl, hydroperoxide, or thiocarbonate end groups.<sup>4</sup>

Furthermore, the RAFT moieties can be easily and completely removed from the polymer, if necessary, especially when it comes to the color, chemical reactivity, and biosafety of the product.<sup>4</sup> To completely remove the thiocarbonylthio moiety, the RAFT polymer can be reduced with radicals and hydrogen atom donors without a significant side reaction.<sup>60,71</sup> Interestingly, by choosing an appropriate radical species, it is possible to even retrieve the RAFT agent.<sup>72</sup> Lastly, thermolysis can be used to completely remove the thiocarbonylthio moiety and involves heating of the polymer to 120–200 °C, resulting in an unsaturated end group functionality.<sup>60,73</sup>

Bioconjugation with RAFT polymers has always been a highlight for diverse hybrid applications in the biomedical field. Especially when it comes to the surface-bound RAFT polymer brush, converting RAFT end groups to specific bio-binding moieties quickly provides the direct conjugation for biomolecules. For example, well-established site-specific bioconjugation methods targeting cysteines (in native proteins)<sup>74</sup>

can be used on the thiol group. Maleimide,<sup>75–77</sup> vinyl sulfone,<sup>78</sup> and activated disulfide<sup>79,80</sup> moieties can be built in the RAFT polymer or converted from RAFT moieties for bioconjugation. On this topic, large efforts were made to optimize the detailed bioconjugation from RAFT polymers. The recent advances in this area are extensively reviewed elsewhere.<sup>54,81–88</sup>

### 2.3. Grafting strategies

The power of functional RAFT polymers is rarely applied in their bulk form. Since their discovery, RAFT polymers were used to enhance the performance of diverse materials and quickly found applications in various fields of research and industry.<sup>2</sup> RAFT polymers are particularly useful for surface modification on nanoscale materials, such as NPs or membranes. The enormous surface area and comparable size of nanomaterials and single polymer chains maximize the effect brought by the polymer functionalization.

The binding of RAFT polymers onto the surface of nanomaterials requires specific chemistry towards each type of surface. A number of different anchoring strategies are available to link RAFT polymers to a surface, which often come with their own merits and shortcomings. Researchers should carefully consider suitable anchoring options depending on the properties of their system, including the type of polymer, the surface character of the NP, the properties of the protecting ligand and the required capping density of the polymer on the surface.

In general, there are two most commonly employed grafting strategies: grafting-to and grafting-from approaches. In the grafting-to strategy, the pre-synthesized polymer is introduced to the surface, whereas in grafting-from, the polymer synthesis is initiated from the NP surface (Fig. 4). Grafting-to is considered a more straightforward method,<sup>89</sup> since the grafting

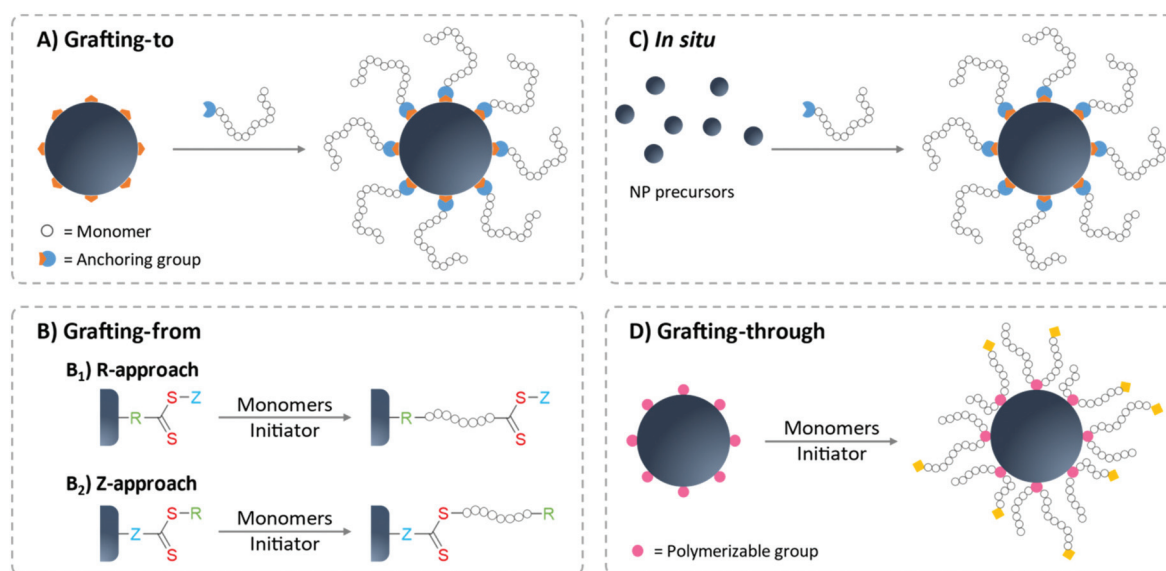


Fig. 4 Illustration of the most commonly used strategies to immobilize polymers onto the surface of NPs. (A) Grafting-to, (B) grafting-from with B<sub>1</sub> for R-approach and B<sub>2</sub> for Z-approach, (C) *in situ* synthesis and (D) grafting-through approach.



step can often be facilitated by a simple ligand exchange reaction: polymers carrying a functional group with a stronger affinity towards the NP surface tends to replace the weaker binding protecting ligand. Also, polymer and NPs are individually prepared to allow for separate syntheses conditions and hence tight control over polymer characteristics.<sup>88,90,91</sup> In contrast, grafting from surfaces demands the attachment of the RAFT agent before RAFT polymerization. In this stage, the surface-bound RAFT agent might alter the colloidal stability of the system since the protecting effect of these small molecules is very limited. Special care on the storage and purification steps must be given to avoid irreversible aggregation.<sup>65,92</sup> Moreover, the polymerization conditions must be optimized for the NPs as well to ensure the complete dispersion of NPs in the polymerization mixture. For the modification of biosurfaces or bio-containing surfaces such as proteins, the grafting-from approach can struggle to retain the biological functionality after polymerization.<sup>91</sup> Furthermore, the characterization of the surface-bound polymer is not as straightforward as for grafting-to approaches.<sup>65,93</sup> Oftentimes, the free polymers formed in solution are characterized as a substitute for the surface-anchored ones, as they are quite similar with respect to molecular weight and dispersity.<sup>94,95</sup>

However, in many cases, grafting-from outperform grafting-to approaches in terms of obtained grafting density.<sup>35,90,91,96,97</sup> This difference can be explained by the steric hindrance effect during the grafting-to process: the approaching polymer chain is sterically blocked by the grafted polymer chains as well as by binding site masking *via* random coil formation of the approaching polymers.<sup>90,91</sup> The grafting-to approach often shows chain length dependence and performs better with shorter polymer chains.<sup>89,98</sup> Other challenges for grafting-to approaches are the ligand exchange process and the efficiency of the anchoring chemistry. Those issues are highly specific and will be discussed in section 2.4. The reader should keep in mind that the surface properties of NPs differ strongly from their bulk counterpart, mainly due to the protection ligand from the synthesis. Oftentimes, choosing NPs with suitable ligand and solvent combination is the key to a successful polymer-ligand exchange.

Notably, the molecular design of the attached RAFT agent plays a critical role when it comes to polymerization characteristics of the grafting-from approach. If the RAFT agent is coupled to the surface *via* its *R*-group (*R*-group approach), the polymer chains grow while being attached to the surface. The propagating radical is located at the solvent-facing polymer terminus, thereby able to readily engage with monomers and RAFT agents in the solution during the whole polymerization process, even at high molecular weights (Fig. 4B). Consequently, high monomer conversion and narrow polymer dispersity of the grafted polymer can be achieved.<sup>88,95,99</sup> Concerns could arise due to the termination between surface-bound radicals (both inter- and intra-particle termination). However, since an excess of free RAFT agent is often added to the polymerization mixture, the termination reaction is then dominated between the solution radicals.<sup>100</sup> Similar to the

solution polymerization without NPs, the amount of initiator must be controlled to avoid the formation of termination product on the surface-grafted polymer shell,<sup>101</sup> which precludes further polymerization to longer polymers or to BCPs. An important advantage brought by *R*-approach is that the RAFT moieties will stay on the outward-facing chain termini of the surface-grafted polymer brushes which can be utilized for further modifications.

In contrast, if the RAFT agent is anchored *via* the *Z*-group (also called transfer-to), the thiocarbonylthio groups are permanently attached to the surface during and after polymerization. This means that radical chain propagation and termination only occur in solution. This approach only allows active radicals to recombine back onto the surface-grafted *Z*-groups, giving the advantage of avoiding the anchoring of any terminated product (Fig. 4B).<sup>100</sup> However, since the recombination of polymer radicals with the surface RAFT *Z*-groups has a similar steric issue as the grafting-to approach, in practical, *R*-group approaches are often favored over *Z*-group approaches, especially for longer polymers.<sup>88,90,101,102</sup> Furthermore, for the first step of anchoring RAFT agents onto the surface, the *R*-group offers easier access to introducing anchoring moieties over the *Z*-group.<sup>100</sup> Also, the *Z*-group approach locates the thiocarbonylthio group directly on the surface of the NPs, which comes with a potential risk of chain stability loss due to hydrolysis or aminolysis of the RAFT agent.<sup>103</sup>

Another approach for introducing polymers onto nano-surfaces is the grafting-through method, in which the monomer is anchored on the surface instead of the RAFT agent.<sup>104,105</sup> During polymerization, this group is incorporated into the growing polymer chain (Fig. 4D). This often leads to intermolecular cross-coupling as one polymer chain can incorporate polymerizable groups from more than one NP surface.<sup>106</sup> This characteristic can be beneficial for the fabrication of gels or matrices, but is rather undesired for colloidal systems.

In all aforementioned grafting approaches, the particles are synthesized prior to polymer functionalization. Another important strategy to fabricate polymer-capped NPs is using the polymer micelle as a template during NP synthesis. This approach can often be designed in a one-pot and *in situ* experiment, performing synthesis and polymer capping simultaneously (Fig. 4C). This direct synthesis approach brings several advantages, for one the polymer usually acts as a stabilizer or surfactant during the synthesis, circumventing the need for additional chemicals, which might require further clean-up/ligand exchange.<sup>35,107</sup> On this topic, the design of the BCP offers a very intriguing architecture of the polymer micelle morphology which can further be used to create nanomaterials according to the shape of the polymer template. Readers can refer to a recent review article on this topic.<sup>108</sup>

In summary, understanding the mechanism and character of each approach is necessary for the successful design of the experiment. The approach of introducing polymer shells on the surface of NPs must be chosen according to the specific type of NP including its surface ligands and solvent con-



ditions. More specific strategies to introduce RAFT polymers on distinct types of NPs are discussed in section 2.4.

## 2.4. Surface chemistry for anchoring polymers

One of the most crucial chemical engineering tasks for the fabrication of polymer-NP structures is the junction point between the polymer and NPs. The linkage between the synthetic polymer and surface of NPs requires strong interaction, or even covalent bonds, to prevent the structure from disassembling. The anchoring itself is a dynamic process and must be approached using an effective chemical reaction or strong adsorption. For this task, customized to the property of the surface, functional groups bearing such anchoring competence are introduced to the polymer chain. The position for these anchoring moieties in the polymer architecture is highly flexible, they can be present as part of the RAFT agent (end group),<sup>109,110</sup> integrated as polymer side chains,<sup>111,112</sup> or introduced after polymerization.<sup>113,114</sup> Here, we demonstrate strategies for polymer functionalization of a selection of the most popular NPs. Table 1 and Fig. 5 give an overview of the content in this section.

**2.4.1. Noble metal nanosurfaces.** Noble metal nanocomposites, especially gold, silver, and palladium NPs have a substantial impact in diverse fields including electronics,<sup>115,116</sup>

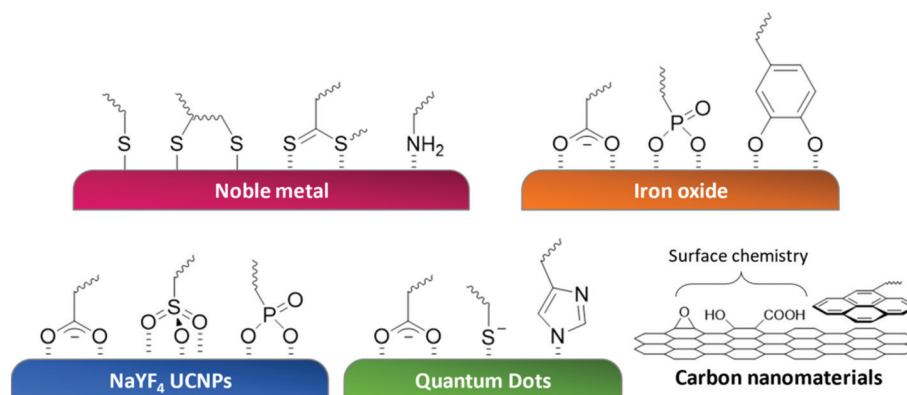
optics,<sup>117,118</sup> sensors,<sup>119,120</sup> catalysis,<sup>121–123</sup> energy conversion,<sup>124</sup> photochemistry,<sup>125</sup> nano-patterning,<sup>126</sup> bio-imaging,<sup>127,128</sup> drug delivery<sup>129,130</sup> and medicines.<sup>131–133</sup> These particles are often very sensitive to impurities and solvent change, thus for most applications, surface modification is inevitable for enhanced colloidal stability, but it may also be used for introduction of facile functions, *e.g.*, stimuli-responsiveness.

In general, sulfur-containing groups, *i.e.*, thiols,<sup>134</sup> disulfides<sup>135</sup> and RAFT groups<sup>136,137</sup> have strong interaction with noble metal NPs. The inherently thiocarbonylthio-terminated RAFT polymer offers a well-known and straightforward approach to form dense polymer shells on the surface of Au and Ag NPs.<sup>138–140</sup>

Owing to the strong interaction between the RAFT group and the surface of noble metal NPs, the grafting-to approach works well here: the RAFT polymer can be synthesized separately without the presence of NPs avoiding the risk of causing any aggregation of NPs during the polymerization step. The self-assembly of RAFT polymers with noble metal NPs often has simple protocols by typically mixing them in a good solvent for both polymer and particles. However, a sufficient NP surface activity is required: strongly bound surfactants on NPs often jeopardize the ligand exchange process, especially

**Table 1** Examples of commonly used anchoring groups to immobilize polymers on different nanosurfaces

	Gold	Silver	Iron oxide	NaYF <sub>4</sub> UCNPs	QDs	Carbon nanomaterials
Carboxylic groups			141–143	144 and 145	146 and 147	148–153
Thiol groups	65, 68, 89 and 154–162	163			164	
Dithiol groups	161	138 and 165				
Dithiobenzoate groups	136 and 166	138				
Trithiocarbonate groups	109, 136, 139, 156 and 167–174	140 and 175				
Thiocarbamate groups	157					
Sulfo group				144		
Amine groups	155 and 176–179					105
Phosphate groups			111 and 180–182	144 and 183		
Catechol groups		184	185–187			
Imidazole groups					112, 188 and 189	
Pyrene moieties						110, 190 and 191



**Fig. 5** Schematic illustration of commonly used anchoring strategies to immobilize polymers on different nanosurfaces.





for macromolecules. The detailed ligand exchange of each type of nanocomponent must be studied specifically. In some cases, the partial removal of original protecting ligand is necessary.<sup>167,175,192</sup>

To discuss the performance of RAFT polymer shells on noble metal NPs, the interaction between polymer and particle as well as the grafting density are the key factors. Here, thiol and RAFT groups can be both applied as anchoring moiety. The thiol group is covalently bound to the surface of gold NPs through a redox reaction. The strong RS–Au bonds<sup>134,193</sup> provide rapid and robust binding with a binding strength of approximately 170 kJ mol<sup>−1</sup> (ref. 194) and total net stabilization energy of roughly −20 kJ mol<sup>−1</sup> considering the break of the S–H bond and formation of the H–H bond.<sup>195</sup> The RAFT group, *e.g.*, dithiocarbonate, has a bonding energy of −16 kJ mol<sup>−1</sup> (ref. 136) and free energy of adsorption of −36 kJ mol<sup>−1</sup> (ref. 137) towards the gold surface. The thiol group seems to be a better choice due to its stronger and irreversible bonding despite extra reaction and purification steps from the RAFT polymer. Under this consideration, RAFT groups are often reduced to thiol groups prior to particle functionalization.<sup>196–199</sup> Slavin *et al.* demonstrated that the capping density of the polymer brush on the gold surface with diverse sulfur-containing groups has the following trend: disulfide > dithiocarbonate ~ trithiocarbonate > free thiol for diverse polymer chain lengths.<sup>200</sup> Wang *et al.* showed that the end group fidelity of thiol-functionalized polymer has a significant impact on the capping density of polymers on AuNPs: the unfunctionalized polymer (from radical termination processes) has a jeopardizing effect on the grafting-to process.<sup>201</sup>

For RAFT groups, Blakey *et al.* also confirmed that the RAFT group does not dissociate upon adsorption to the AuNPs.<sup>137</sup> In fact, a number of high-density RAFT polymer-capped noble metal NP hybrids have been fabricated by successful ligand exchange with *e.g.*, citrate,<sup>139,202,203</sup> oleylamine,<sup>140</sup> cetyltrimethylammonium bromide (CTAB)<sup>167,192,204</sup> and tetraoctylammonium bromide (TOAB).<sup>140</sup> Our group also confirmed the high long-term thermal stability of the RAFT–Au interaction up to 90 °C on both AuNPs and AgNPs covered with RAFT-terminated polyethylene shells.<sup>140</sup> Thiol and RAFT moieties both have numerous successful results in fabricating well-defined polymer brush shells on noble metal NPs. The binding efficiency of RAFT and thiol groups on palladium NPs was compared by Rui *et al.*<sup>205</sup> As result, no significant difference was found. In the author's opinion, both RAFT moieties and thiols work great as anchoring functionality for noble metal NPs.

The grafting-to approach serves as the dominating synthesis strategy for modification of noble metal NPs with polymers. As the sulfur-containing groups of the RAFT agent act as the reactive center during chain transfer reactions, surface anchoring *via* RAFT groups is less suitable for grafting-from syntheses. Other functional groups like amines,<sup>176</sup> alkynes,<sup>206,207</sup> catechols<sup>208</sup> or phosphonates<sup>209</sup> can be used for attachment to gold surfaces. However, the stronger aurophilicity and inherent presence of the RAFT group make the sulfur-containing anchoring approaches much more popular.

Similar to gold, silver NPs possess an affinity towards sulfur-containing moieties (thiols,<sup>163</sup> disulfides,<sup>165</sup> and RAFT moieties.<sup>138,175</sup> The Ag–S bound has a binding energy of 217 kJ mol<sup>−1</sup> (ref. 210) and a colloid formation free energy of −25 kJ mol<sup>−1</sup> (ref. 211) (ethanethiol) for NP surface. The thermodynamic values of thiol-binding on AgNPs and AuNPs are very similar. The strong anchoring performance of thiocarbonate group on AgNPs is also confirmed.<sup>175</sup> For further noble metal NPs, thiol also provide a well-known solid anchoring performance, *e.g.*, on Pt and Rh NPs.<sup>197,212</sup> Notably, the situation of Pd–S interaction is much complicated since the thiol can react with Pd forming PdS<sub>x</sub> layer. Readers should refer to the detailed works<sup>213,214</sup> on this specific topic if the surface properties of the NPs are crucial for the applications.

Besides AuNPs and PtNPs, metal NPs are notorious for their oxidation tendency. Depending on the synthesis route, oxygen-free conditions could be required for maintaining the surface of metal NPs active for surface modification. In fact, anchoring strategies targeted on the oxidized surface also have been reported *e.g.* for ZnNPs,<sup>215</sup> AgNPs,<sup>184</sup> and gallium–indium (liquid metals) based NPs<sup>216</sup> typically using catechol or carboxylic anchoring groups.

**2.4.2. Superparamagnetic iron oxide NPs.** Superparamagnetic iron oxide NPs (IONPs), *e.g.*, magnetite (Fe<sub>3</sub>O<sub>4</sub>) and maghemite (γ-Fe<sub>2</sub>O<sub>3</sub>) have found extensive application especially in the medical field, *e.g.*, for magnetic resonance imaging,<sup>217</sup> drug delivery<sup>218,219</sup> and tumor treatment (hyperthermia)<sup>220–222</sup> due to its highly favored magnetic properties. Catechols form very stable complexes with Fe<sub>2</sub>O<sub>3</sub> and Fe<sub>3</sub>O<sub>4</sub> surfaces<sup>223,224</sup> and are therefore widely used as an anchoring group for these magnetic NPs.<sup>34</sup> Notably, the binding affinity of catechols to iron oxide nanosurfaces has been shown to increase by the introduction of electronegative groups such as −NO<sub>2</sub> to the aromatic ring.<sup>225,226</sup> Using a grafting-to approach, catechol-terminated polymers can be introduced onto the surface of unfunctionalized iron oxide<sup>226</sup> or even successfully exchanged with oleate/oleylamine<sup>227</sup> and oleic acid<sup>228</sup> protecting ligands. In these publications, the close interparticle distance indicates the low capping density of the polymer, which is probably caused by the combination of the grafting-to approach and limited anchoring performance. To introduce a dense polymer shell on the surface of iron oxide, Ohno and coworkers have demonstrated a grafting-from approach by using triethoxysilane as an anchoring group for surface-initiated atom transfer radical polymerization (ATRP) on both Fe<sub>3</sub>O<sub>4</sub> NPs<sup>229</sup> and β-FeOOH nanorods (NRs).<sup>230</sup>

Two other surface affinity groups found in the literature for attachment of polymers to iron oxide NPs are phosphonates and carboxyls. Out of the two, phosphonates create more stable linkages with higher grafting densities;<sup>223,231</sup> however, the use of phosphonate moieties often requires protection prior to polymerization.<sup>180,232</sup> Carboxyls can also be used for surface modification,<sup>142</sup> but it has been reported that this anchoring is temperature-sensitive and replaceable with other ligands.<sup>223</sup>

**2.4.3. Quantum dots (QDs).** QDs are strong fluorescent nanocrystals made from a range of semiconductor core





materials including CdSe, CdS, CdTe, and PbSe, which are often equipped with a shell made from *e.g.*, ZnS for oxidative protection as well as quantum yield enhancement.<sup>233</sup> As fluorescent nanomaterials with tunable luminance wavelength up to the near-infrared (NIR) region, QDs have a very high potential to be applied as nano-fluorescent marker for biomedical imaging tasks.<sup>234</sup> Introducing polymer shells on the surface of QD has been a common route towards enhancement of biocompatibility and introduction of further functionalities for QDs.<sup>112,188</sup>

The surface modification of QD often brings an impact on its fluorescence performance since the photon luminance properties of QD are strongly dependent on the surface properties and surface–ligand interaction. First, a strong ligand–surface interaction and dense ligand shell are required to prevent QD surface oxidation since the oxidation layer destroys the perfection of the QD surface causing the change in the energy level and luminance.<sup>235,236</sup> Also, the type of surface ligand can influence the electronic properties of the QD.<sup>237</sup>

Although small molecules, *e.g.*, thiol derivatives are often-times used for QD surface modification.<sup>236,238</sup> The nature of the thiol–QD interaction is very distinct from the simple Au–S interaction: on CdTe QD, a thin layer of CdS is formed and the anchoring originates from the interaction between CdS (or ZnS shell) and thiolate rather than with thiol moieties.<sup>239,240</sup> The colloidal stability provided by small molecules is generally limited. One way of circumventing this issue is to use macromolecular ligands, which can be easily prepared *via* RAFT polymerization. Here, a multiply-binding copolymer design has been proven to be very effective: one type of monomer contains an anchoring group for QD binding, such as thiol,<sup>164</sup> carboxyl,<sup>146,147</sup> imidazole<sup>112,188,189</sup> and 8-hydroxyquinoline<sup>241</sup> moieties. Another part of the polymer adjusts dispersibility and provides additional properties.

An alternative option for surface modification and QD stabilization is the use of amphiphilic BCPs. QD synthesis often employs hydrophobic surfactant molecules, rendering them water-insoluble.<sup>233</sup> The hydrophobic part of the amphiphilic BCP can interact with this outer QD layer, whilst the hydrophilic block confers hydrophilicity and added functionalities such as pH-responsiveness as demonstrated by Liu *et al.*<sup>242</sup>

**2.4.4. Upconverting NPs (UCNPs).** UCNPs are another important material class designed for photoluminescence, and are especially suitable for biomedical applications.<sup>243,244</sup> The as-synthesized UCNPs often come with hydrophobic protecting ligands (*e.g.*, oleic acid, oleylamine), making surface ligand exchange inevitable for any application in aqueous media. Since the surface of UCNPs does not have access to any known covalent anchoring option, interactions based on charge and hydrogen bonds are often applied. The anchoring performance is considered to increase in the order –SH, –NH<sub>2</sub>, –COOH, –PO<sub>3</sub>H<sub>2</sub>,<sup>245</sup> but no detailed studies are available to the best of the authors' knowledge. The single linkage provided by these interactions is relatively weak compared with the covalent binding and sensitive to environmental change (pH,

temperature, *etc.*). For this reason, multiply-binding ligand design is also preferred here to achieve strong binding performance.<sup>246,247</sup> Duong *et al.* performed a detailed study on different NaYF<sub>4</sub>:Yb/Er UCNP surface binding groups. They synthesized RAFT BCPs comprised of a block of either phosphate, sulphonic, or carboxylic acid-containing monomers and a block of hydrophilic polyethylene glycol (PEG)-like monomers. After the ligand exchange with oleic acid molecules on the surface of UCNPs, they investigated the colloidal stability of the particles in different solvent conditions. They demonstrated that phosphate groups were the most effective anchoring moieties for providing colloidal stability.<sup>144</sup> Alternatively, small molecules containing multiple phosphate groups, *e.g.*, alendronate, can be used for UCNP surface modification followed by RAFT agent anchoring for a grafting-from approach.<sup>183</sup> Notably, also amines can be used for surface binding, although they provide even less stability than carboxyl groups.<sup>247</sup>

One of the most applied approach to effectively introduce polymers or biomolecules to UCNPs is using a silica coating. With a nano-thin silica-shell around UCNPs, surface modification with well-established silane chemistry can be applied without any special modification. RAFT polymerization can also proceed from the silica surface (see section 2.4.6 for more information).<sup>248–250</sup>

**2.4.5. Carbon nanomaterials.** Carbon-based nanomaterials such as carbon nanotubes and graphene have also been used in combination with RAFT polymerization. The most common strategy for surface modification of these materials starts with oxidation, which can introduce functional moieties such as hydroxyls, carboxyls, and epoxides to the surface. There are many protocols to perform the oxidation such as using H<sub>2</sub>O<sub>2</sub>, H<sub>2</sub>SO<sub>4</sub>, HNO<sub>3</sub>, and KMnO<sub>4</sub>.<sup>105,148–151,251</sup> In this step, the choice of oxidizing agent has a significant influence on the type and abundance of functional groups, with KMnO<sub>4</sub> being the most efficient and introducing the highest density of carboxyl moieties, whereas H<sub>2</sub>O<sub>2</sub> is the least reactive but leads to the highest density of surface hydroxyl moieties.<sup>251</sup>

The choice of oxidation depends on the desired anchoring chemistry. An esterification reaction between the hydroxyl group on carbon nanomaterials and carboxyl groups on the RAFT moieties is often used to introduce RAFT agents to the surface for grafting-from polymerizations.<sup>150–153</sup> Alternatively, one can perform a nucleophilic ring-opening reaction of epoxide groups using amines, which has been demonstrated by Mardani *et al.* to introduce a polymerizable methacrylate on graphene oxide (GO) for a subsequent grafting-through RAFT polymerization.<sup>105</sup> Moreover, it would be possible to couple the surface carboxyl groups with primary amine moieties using *N*-hydroxysuccinimide (NHS) click chemistry.<sup>252</sup> A detailed comparison between grafting strategies for RAFT polymers and click chemistry is reported by Hwang and coworkers.<sup>253</sup>

Another useful approach to immobilize polymers to the surface of GO is using pyrene moieties as anchoring groups due to a strong  $\pi$ – $\pi$  stacking interaction.<sup>110,190,191</sup> Owing to this



strong binding energy (comparable with covalent attachment), the straightforward grafting-to protocols have been proven to be very effective.<sup>110,190</sup>

**2.4.6. Nanomaterials with silica coating.** Colloidal silica coating is a fantastic tool to modify the surface of NPs and simplify the surface modification process. Nanomaterial coated with a thin layer of silica possesses the surface properties of silica (*e.g.*, high colloidal stability in water and biocompatibility)<sup>254</sup> and has access to all well-established silica chemistry.<sup>254</sup> The protocol for silica coating is often easy to perform and always comes with good control over the silica layer thickness. As silica coating can be performed with a very wide range of core materials,<sup>255</sup> this can be seen as the “all-round strategy” for the introduction of surface modifications, especially when the bare NP core surface does not provide effective surface chemistry for functionalization.

The classic Stöber process for silica coating requires the dispersion of NPs in ethanol (or mixture with water). With the addition of ammonia, tetraethylorthosilicate (TEOS) is hydrolyzed into silica precursors and subsequently condensates, forming silica on the surface of NPs layer by layer.<sup>256,257</sup> The silica shell thickness can be tuned by varying *e.g.*, the amount of ammonia and TEOS or the reaction time.<sup>258–261</sup> For a successful silica coating, perfect dispersion of the nanomaterial during the reaction is crucial to avoid aggregation. The perfect dispersion of originally hydrophilic nanomaterials in ethanol requires surface modification. For this task, polyvinylpyrrolidone (PVP) provides generally good adsorption on diverse nanomaterials, making them suitable for the silica coating condition.<sup>255</sup>

Another approach based on the water-in-oil (reverse) microemulsion technique has the special ability to perform one-on-one silica coating if the dosing of particles roughly matches the number of micelles. In these approaches, NPs must be dispersed in nonpolar media, *e.g.*, cyclohexane.<sup>258</sup> Silica coating takes place on the NP surface isolated in water micelles. The resulting particles are highly spherical and possess a narrow size distribution. Generally, microemulsion systems are able to work at higher NP concentrations compared to the Stöber coating with much lower risk of aggregation.<sup>262</sup> However, microemulsion-based approaches possess an upper limit for the size of the used NP cores as well as the resulting silica-shell thickness, which is amongst other influenced by the size of the microemulsion droplets and the type of used surfactant.<sup>263</sup> For instance, Han *et al.* reported an upper size limit for the resulting silica-coated particles of approximately 90 nm.<sup>262</sup>

Grafting strategies for polymer attachment onto silica surfaces has been extensively studied. By far the most efficient and widely used strategy to introduce RAFT polymers on silica is the surface-initiated grafting-from approach. The RAFT moieties are often first introduced to the surface of silica *via* a two-step reaction. First, the surface is functionalized with amine groups using aminosilanes followed by RAFT agent anchoring *via* activated carboxyl groups.<sup>99,168,248–250,264,265</sup> The choice of silane group has strong impact on the result since the number

of alkoxyisilyl groups on each silane has very different performance: the tri- and di-alkoxyisilanes have the risk of formation of interparticle crosslinking causing the formation of aggregated material.<sup>266</sup> Although the monoalkoxyisilane is sensitive to moisture during the reaction, the crosslinking can be completely suppressed.<sup>99</sup> In a similar way, carboxyl-containing polymers can be linked to the aminosilane-functionalized particles in a grafting-to approach using the same coupling reaction.<sup>267,268</sup> The large selection of silanes also allows for other approaches to anchor RAFT groups on the surface of silica, such as *via* chlorosilanes<sup>269</sup> or alkyne-containing silanes.<sup>154</sup> Furthermore, the silica core can be removed by etching with HF or NaOH.<sup>270</sup> This allows the formation of hollow polymer shell by cross-linking the grafted polymer before the etching process.<sup>271–274</sup>

## 2.5. Topological design

RAFT polymerization also provides a vast selection of topological design options. To this end, star,<sup>8</sup> branched, hyperbranched, network,<sup>10,11,275</sup> comb-like, bottlebrush,<sup>12</sup> and in-chain polyfunctional<sup>139,172,173</sup> RAFT polymers are extensively studied. Readers interested in the design, synthesis, and selection of the RAFT agents for those polymer structures are referred to the abovementioned literature. With the knowledge of RAFT mechanisms and suitable *R*- and *Z*-approaches, the transfer of topology from RAFT agent to the RAFT polymer can be executed in a very precise manner with the control of the positioning of the RAFT moieties in the polymer.

Using linear polymer with a single anchoring end group can only form a simple core-shell nanostructure. However, RAFT polymers with a higher order of topological design can provide intriguing competence for further functional extension, *e.g.*, linking further NPs onto the existing core-shell structure. As an example, our group reported the formation of spherical AuNPs superstructures crosslinked with multiblock RAFT polymers containing several in-chain RAFT groups along their backbone. The interparticle distance inside these superstructures can be controlled with the chain length of the linking polymer.<sup>172,173</sup>

The topological design of the polymers has a key impact on the self-assembly of nanomaterials. In section 3.2.3, more examples of hierarchical nanoassemblies fabricated by multi-arm and hyperbranched polymer linkers are reviewed in detail. It is worth mentioning that a change in the topology could affect the stimuli-responsive parameter of the polymer, the polymer content should be adjusted according to the application scenario by *e.g.*, adding another monomer to the polymer.<sup>9</sup>

## 3. Design with RAFT polymers in nanoengineering

The innovation in molecular engineering for a RAFT polymer can be applied to every point of the polymer chain. The development of new functional monomers combined with sophisti-



cated structural design makes RAFT polymers the perfect key building block for fabricating a wide range of nanostructures. In this chapter, we will demonstrate the recent advances in the development of surface-bound RAFT polymers in the realm of nano application.

The “smartness” of RAFT polymers has been rapidly growing during the last decade. The possibility to build multiple intriguing abilities into one (block) copolymer makes RAFT polymers a versatile tool for the surface modification of nanomaterials. With a wide selection of well-established anchoring strategies, the RAFT polymers can be applied as smart linkers for connecting distinct nanomaterials. We will demonstrate how the structural coding from the polymer design can be used to introduce and dynamically control the self-assembly, interparticle-interaction, energy transfer and stimuli-responsiveness of the hybrid nanostructures.

### 3.1. Creating functions with RAFT polymer shells for NPs

The most straightforward application of polymers for NP modification is using them as capping ligands, forming core-shell nanostructures. Once the NP is covered with the polymer shell, these nanostructures often carry the solubility (dispersibility) and functionality of the polymer shell. In recent years, with the rapid innovation of both NP synthesis and functional polymer design, new types of polymer-capped NPs with diverse hybrid functions are emerging. Especially the combination of orthogonal functions in RAFT polymer brushes using copolymer design strategies opens up towards the next level of smart NPs.

To introduce multiple functions to one nanostructure, the (block) copolymer brush design provides the most straightforward and reliable approach. The synergy between different polymer units gives rise to many advantages. This section will review the recent development of RAFT polymer-functionalized core-shell nanomaterials and showcase the novel design options and collaborative properties between the polymer and nanocore. We will focus diverse research topics covering enhancement of the dispersion behavior, hybrid stimuli responsiveness and smart linkers for fluorescence energy transfer-based nanosensors. Table 2 summarizes our selection of publications on the novel functions brought by modern RAFT polymer shells.

**3.1.1. Tuning the interparticle interaction and dispersibility.** For almost any application scenario, the dispersibility of NPs must be sufficient to maintain its favored properties. The interaction between the surface of NPs and the solvent/matrix environment is critical for the dispersibility of NPs. However, the long-term stability of the (colloidal) dispersion also strongly depends on the interparticle interaction. The flexible choice of monomer and the steric protection provided by functional polymer shells enables a wide adaptability and a significant improvement on the stability in various solvents<sup>294,295</sup> and matrices.<sup>140,290</sup>

The requirement of surface modification for nanomaterials regarding this topic is especially challenging when it comes to biomedical applications. Since numerous types of NPs must be

synthesized using hydrophobic ligands, *e.g.*, QDs, iron oxide NPs and UCNPs, these NPs are rendered non-dispersible in aqueous medium without modification.

Besides good water-dispersibility and long-term colloidal stability, the biomedical application further requires the nanomaterials to have high biocompatibility and minimal non-specific interaction in biological environment (antifouling). A very wide choice of capping polymers have been established for this purpose, including PEG,<sup>296,297</sup> poly(2-hydroxyethyl methacrylate) (PHEMA),<sup>298</sup> PVP,<sup>299,300</sup> poly(2-oxazoline)<sup>180</sup> and zwitterionic polymers.<sup>301,302</sup> For general biological applications, PEG and its derivatives are the most widely used ones for their well-known character and commercial availability. Notably, the capping density is crucially important for the performance of PEG capped nanoparticles in biological environment.<sup>303,304</sup> To create high capping density of grafted polymer, efficient and reliable anchoring strategies are required. Again, RAFT polymerization provides a very straightforward synthesis of functional polymers containing PEG derivatives with suitable anchoring groups for diverse nanomaterials.<sup>111,146,181,188,189,238</sup>

Zwitterionic polymers possess equal cationic and anionic moieties on the chain: the polymer maintains charge neutrality while carrying high ion density. This unique property makes them highly resistant to nonspecific protein adsorption due to their strong hydration ability resulting from the high charge density.<sup>302</sup> RAFT polymerization can also polymerize carboxy betaine, sulfobetaine, and phosphorylcholine-based zwitterionic monomers and anchor these polymers onto various NPs.<sup>68,112,161,305</sup>

In many cases, the limiting factor for stability is the anchoring point between the polymer and surface. To avoid any detachment of the functional polymer, in addition, multiple anchoring groups can be introduced as anchoring blocks in the polymer design. Combining both hydrophilic block and anchoring blocks provides long-term colloidal stability across a wide pH-range and against contaminations.<sup>112,161,188,189</sup>

Lequeux and co-workers fabricated zwitterionic vinylimidazole BCPs using RAFT polymerization to decorate the surface of QDs (Fig. 6A). To immobilize the polymer onto the surface of QDs, they used a multidentate anchoring approach *via* the imidazole block. The sulfobetaine-zwitterion polymer provides robust and long-term stability. The inserted primary amines in the zwitterionic block allow for oriented bioconjugation with IgG antibodies and carry remarkable targeting performance towards specific proteins with a very low level of unspecific binding to live cells. Combining high stability provided by the polymer shell and the robust luminance performance of QDs, the observation time can exceed the 48 h window which is rarely feasible with conventional approaches.<sup>112</sup>

Emrick and co-workers coated AuNRs with zwitterionic polymers with multi-thiol anchoring monomer units (Fig. 6B). A random copolymer was prepared by lipoic acid-substituted hydroxyethyl methacrylate (LA-HEMA) and methacrylamide phosphorylcholine (MPC) monomers. The polymer is then reduced with NaBH<sub>4</sub> to convert RAFT and disulfide moieties of





**Table 2** Examples of nanomaterials functionalized with polymer shell synthesized from RAFT polymerization and their applications

Nanomaterial	Grafted polymer	Applications	Ref.
AgNP	Branched PEG- <i>star</i> -poly((2-hydroxypropyl methacrylate)- <i>co</i> -(dithio-di-2,1-ethanediy bismethacrylate)- <i>co</i> -(lipoic acid methacrylate)- <i>co</i> -(2-(4-nitro-3-benzyl carbonate camptothecin)phenoxyethyl methacrylate))	Water dispersibility; biocompatibility; photo-responsive drug-release tracked by fluorescence (NSET)	165
AgNP	Fluorescent dye functionalized PAA	pH responsive metal-enhanced fluorescence behavior	276
AgNP	Poly(2-(2-hydroxyethoxy)ethyl methacrylate- <i>co</i> -methacryloyloxy-3-thiahexanoyl-camptothecin)	pH-sensitive drug-delivery with tracking of drug-release progress <i>via</i> NSET controlled fluorescence	138
AgNP	Poly(methyl methacrylate- <i>co</i> -(2-hydroxypropyl-9-anthroate methacrylate))	Surface plasmon resonance enhanced upconversion of fluorescence	163
Au nanocage	PNIPAM- <i>co</i> -poly(acrylamide)	High-intensity focused ultrasound induced drug release	277
AuNP	Poly(PEG methacrylate)	Controlled aggregation upon addition of 2-phenyl-2-(phenylcarbonothioylthio) acetate, yielding nanoaggregation with tunable optical properties <i>via</i> surface plasmonic coupling	278
AuNP	$\omega$ -Carboxyl-terminated PNIPAM	Thermo- and pH-controlled reversible phase transfer between water and chloroform	279
AuNP	Azobenzene-containing methacrylic polymer	UV-light triggered aggregation of AuNPs	166
AuNP	PNIPAM with pillararene end-group	Introduces hexafluorophosphate-pillararene host-guest interaction alone with thermoresponsiveness from PNIPAM	280
AuNP	Poly(di-(ethylene glycol) methyl ether methacrylate)- <i>b</i> -(poly(6- <i>O</i> -vinylazelaioyl-D-glucose))	Water dispersibility; biocompatibility; lectin recognition	66
AuNP	Polystyrene- <i>b</i> -PEG with trithiocarbonate group as the junction between two blocks	Amphiphilic colloid properties	281
AuNP	Poly(hydroxyethyl acrylamide) with amino-benzylguanine or tris-NTA amine end-group	Bioconjugation with anti-freeze proteins	198
AuNP, AgNP, CdSe QD	Thiol-terminated <i>meta</i> -triphenylamine polymer	Electron conductivity	282
AuNR	Poly(MPC- <i>co</i> -HEMA-DHLA)	Water dispersibility; high resistance against cyanide ion etching; antifouling; cytocompatibility	161
AuNR	PNIPAM	NIR light triggered photothermal drug-release	199
AuNR	PEG- <i>b</i> -poly( <i>N</i> -vinylcaprolactam)	NIR triggered drug-release	283
AuNR	3-Arm PNIPAM	NIR-light triggered photothermal aggregation of AuNRs in simulated blood fluids	167
Carbon NP	Copolymer of NIPAM and spiropyran containing units	Reversible thermo- and light-responsive fluorescent behaviors	284
CdSe/CdS/ZnS QD	Random copolymer or BCP of PEG methacrylate and <i>N</i> -methacryloyl succinimide	Water dispersibility; stable interaction with QDs against L-glutathione	189
CdSe/CdS/ZnS QD	Copolymer consist of three monomers: PEG side chain monomer; imidazole groups containing unit; primary amines or biotin groups	Water dispersibility; antifouling; high stability; conjugation with dye or protein	188
CdSe/CdS/ZnS QD	Poly((methacrylamidosulfobetaine- <i>co</i> - <i>N</i> -(3-aminopropyl) methacrylamide hydrochloride)- <i>b</i> -(4-vinylimidazole))	Extracellular stability; antifouling; bioconjugation	112
CdSe/ZnS and CdTe/ZnS QDs	Polymers consist of styrene, NIPAM, vinyl carbazole and 8-hydroxyquinoline derivative monomers	Multi-emission hybrid micelles	241
CdSe/ZnS QD	Poly(2-( <i>N,N</i> -diethylamino) ethyl methacrylate- <i>b</i> -poly(2-methacryloyloxyethyl phosphorylcholine- <i>co</i> -( <i>n</i> -Butyl methacrylate)- <i>co</i> -( <i>p</i> -Nitrophenyloxycarbonyl oligo(ethylene glycol) methacrylate)) conjugated with fluorescent dye	Water dispersibility; cytocompatibility; pH-sensitive FRET between QD and dye	242
CdSe/ZnS QD, Au	Poly(St) on AuNPs, poly(4-vinylpyridine) on QDs	By forming doubly alternating arrays of Au NPs and QDs within onion-like poly(St- <i>b</i> -(4-vinylpyridine)) BCP particle, the hybrid microphers has a solvent-responsive fluorescence switching ability	285
CdSe/ZnS QD	PNIPM with end-functionalized fluorescent dye	Thermoresponsive dual photoemission nanosensor using LCST of PNIPAM and FRET between QD and dye	286
CdTe QD	Dye-labeled poly( <i>N</i> -(2-thioethyl methacrylamide)	Water dispersibility; pH-controlled release of the DOX; FRET between donor QDs and acceptor dye on the polymer	164
CdTe QD	Poly(PEG methacrylate) with adenosine attached to the end of the PEG branches	Water dispersibility; antifouling; bioconjugation	238
GO	Poly(NIPAM- <i>co</i> -butyl methacrylate), poly(NIPAM- <i>co</i> -dimethylaminopropylacrylamide) and PNIPAM, all with addition fluorescent block	Precise detection of temperature between 25–45 °C within micronized domains using fluorescence color	110
GO and CdSe/ZnS QD	PAA and poly(2-vinylpyridine)	Efficient colorimetric pH sensor based on FRET between GO and QDs	287
GQD	Poly(7-(4-(acryloyloxy)butoxy)coumarin)- <i>b</i> -PNIPAM	pH, temperature and metal ion responsive multicolor fluorescence behavior based on FRET	288



Table 2 (Contd.)

Nanomaterial	Grafted polymer	Applications	Ref.
IONP	Homopolymer or BCP of POEGA and PDMAEA	IONP siRNA nano-carriers with antifouling-shell; cytocompatibility	181
IONP	Thiabendazole imprinted PNIPAM	Reusable and specific recognition of benzimidazole	269
IONP	PNIPAM	Hyperthermia and heat-triggered drug-delivery	187
IONP	Poly(oligo(2-ethyl-2-oxazine) methacrylate)	Water dispersibility; negligible cytotoxicity; thermoresponsive behavior with tunable LCST	180
IONP	DBM-terminated poly(OEGA)- <i>b</i> -(poly(phosphonate acrylate))	Water dispersibility; antifouling; bioconjugation; cell tracking <i>via</i> fluorescence labelling	111
IONP	PNIPAM- <i>b</i> -PAA	Thermal and pH dual responsive drug-delivery	268
Mesoporous silica NP	Poly(PEG acrylate)	Thermoresponsive drug-delivery with large cargo capacity	289
Montmorillonite	Poly(methyl acrylate)	Enhanced mechanical properties of poly(methyl acrylate)	104
MoS <sub>2</sub>	P7AC- <i>b</i> -PNIPAM	Real-time photothermal heating and imaging using LCST of polymer and FRET between MoS <sub>2</sub> and fluorescent polymer block	114
NaYF <sub>4</sub> based UCNF	PAA- <i>b</i> -PEG with terminated RGD peptide	Cell imaging and labeling with drug-delivery functions	183
NaYF <sub>4</sub> :Yb/Er UCNF	POEGA- <i>b</i> -PAA, POEGA- <i>b</i> -poly(monoacryloxyethyl phosphate) and POEGA- <i>b</i> -poly(2-acrylamido-2-methyl-1-propanesulphonic acid)	Colloidal stability in water	144
PbS/CdS QD	Copolymer bearing carboxylic acid anchoring group and PEG chains	Long-term colloidal stability in isotonic saline; <i>in vivo</i> NIR imaging	146
Silica coated Fe <sub>3</sub> O <sub>4</sub> NP	PNIPAM	Inhibition of magnetic interaction between Fe <sub>3</sub> O <sub>4</sub> NPs cores	264
Silica coated Fe <sub>3</sub> O <sub>4</sub> NP	Poly(NIPAM- <i>co</i> -GMA) with hydrazine moieties in GMA units	Temperature and pH-controlled drug release	265
Silica NP	Polyisoprene	Incorporation into a polyisoprene matrix	290
Silica NP	PAA- <i>b</i> -poly(St)	Proton conductive membrane	291 and 292
Silica NP	Poly(NIPAM- <i>b</i> -spirooxazine acryloyl- <i>b</i> -DMAEA)	Multi-responsiveness towards UV-vis light, pH, temperature	293

LA into thiols. They used a grafting-to approach to immobilize poly(MPC-*co*-HEMA-DHLA) onto AuNRs. Notably, the ligand exchange of the polymer with CTAB on AuNRs is challenging due to the dense protection provided by the original ligand: the CTAB excess must be removed carefully. Here, the multiply-anchoring moieties could have promoted the ligand exchange process. The poly(MPC-*co*-HEMA-DHLA)-capped AuNRs demonstrated exceptionally high stability against challenging conditions. The polymer shell even offers the resistance against cyanide ion etching for AuNRs. This structure was shown to be non-cytotoxic and demonstrated antifouling properties.<sup>161</sup>

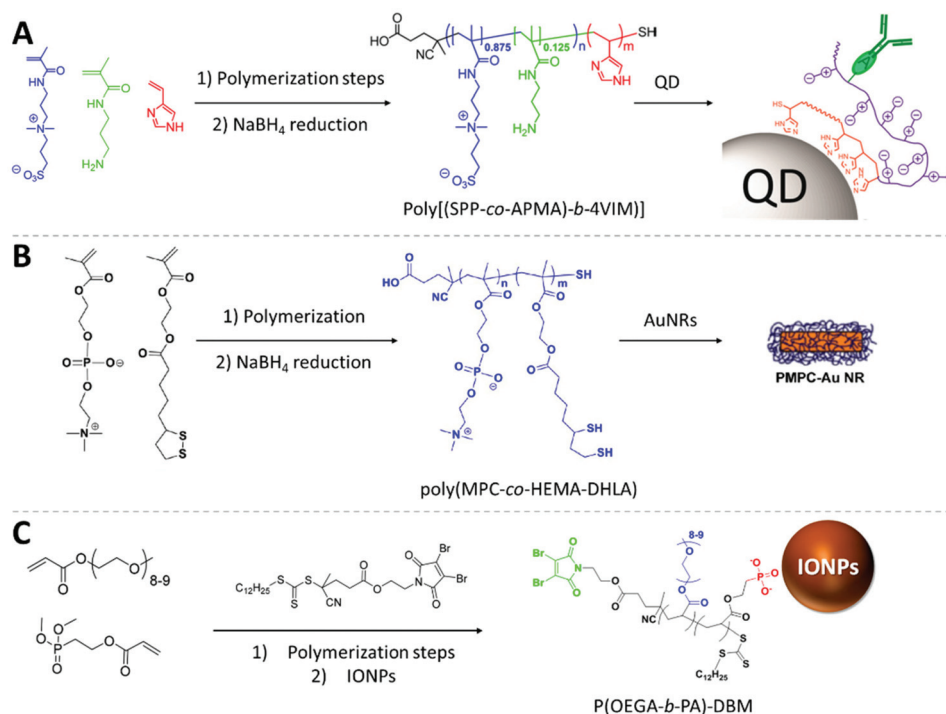
Davis and co-workers have reported the synthesis of superparamagnetic iron oxide NPs coated with dibromomaleimide (DBM)-terminated BCPs consisting of PEG acrylate and phosphonate acrylate (PA) monomers (Fig. 6C). The PA block serves as a multiply-binding anchor for the iron oxide surface providing a stable grafting performance. The poly(oligoethylene glycol) methyl ether acrylate (POEGA) block offers biocompatibility. The DBM group was integrated in the *R*-group of the RAFT agent and remains at the brush termini after polymerization which allows for covalent conjugation with biomolecules *via* chemoselective reactions with both thiol and amino-terminal molecules. Using DBM moieties, they demonstrated the conjugation with an NIR dye and peptides. This work impressively demonstrated the possibility to introduce the highly

favorable bio-targeting and fluorescence tracking function to iron oxide NPs including high colloidal stability and antifouling character with only one type of RAFT polymer. The combination of all these features created an all-around nanostructure for cancer treatment.<sup>111</sup>

We have seen that both random and BCP design with hydrophilic monomers are used for enhancement of colloidal stability. An interesting study was published recently by Dunlap *et al.*, who compared the relative stability between block and random copolymers consisting of PEG and imidazole-containing (QD-anchoring) monomers. They found that BCPs have a more stable interaction with QDs than random copolymers against L-glutathione. Also, higher molecular weights contributed to higher colloidal stability of QDs for both types of polymers.<sup>189</sup>

**3.1.2. Thermoresponsive polymer shells.** RAFT polymerization is a very universal tool to introduce stimuli-responsive properties to nanomaterials. Among all the stimuli factors, thermal responsiveness is one of the most useful and widely studied abilities. In general, the solubility of such a stimuli-responsive polymer will transitionally change with temperature and induce a solution-cloud transition. This thermoresponsive transition is a combination of enthalpic and entropic effects. The mechanism and recent development on this type of polymer are reviewed in detail elsewhere.<sup>15,306,307</sup> The solubility transition often comes with a release of solvent molecules,





**Fig. 6** Fabrication strategies of RAFT copolymers and the illustrated core-shell polymer-capped NPs. (A) The 3-[3-methacrylamidopropyl-(dimethyl)-ammonio]propane-1-sulfonate-co-(N-(3-aminopropyl)methacrylamide hydrochloride) block serves as hydrophilic and bio-binding block. The 4-vinylimidazole block is used as a multidentate anchoring block for the QD surface. Adapted with permission from ref. 112. Copyright 2015 American Chemical Society. (B) The anchoring of the copolymers on AuNRs is enabled by the thiol groups on the HEMA-DHLA unit, and MPC zwitterionic units provide the antifouling property. Adapted with permission from ref. 161. Copyright 2013 American Chemical Society. (C) The phosphonate groups from the PA block anchor on the surface of IONPs, the POEGA block provides biocompatibility, the DBM end groups serve as binding site for further functionalization with fluorophores. Adapted with permission from ref. 111. Copyright 2018 American Chemical Society.

*e.g.*, water, and shrinkage of the polymer chain. This property makes thermoresponsive polymers useful candidates for drug-delivery applications since the release of a drug can be accomplished simultaneously.<sup>308</sup>

By anchoring the thermoresponsive polymer onto the surface of NPs, the cloud transition or aggregation can be effectively transferred to the nanostructure.<sup>309</sup> This approach was employed successfully for a wide range of applications especially in the biomedical field including drug delivery, tissue engineering, bioimaging and hyperthermia.<sup>289,310–313</sup> The convenient temperature-dependent reversible dispersibility also finds its applications in catalysis<sup>314,315</sup> and sensing.<sup>316–318</sup> Furthermore, combining the thermoresponsiveness with other types of stimuli-responsive polymers can create multi-responsive properties.<sup>319,320</sup>

Although the temperature-triggered aggregation of thermoresponsive polymer-capped NPs is a very useful function, the origin of these transitions is the release of solvent molecules attached to the polymer chain. As an example, the widely used poly(*N*-isopropylacrylamide) (PNIPAM) collapses above its lower critical solution temperature (LCST) due to a loss of most of its attached water molecules in the fully hydrated chain.<sup>321</sup> Jones *et al.* have recently demonstrated the importance of residual free PNIPAM in the LCST-triggered aggrega-

tion of PNIPAM capped AuNPs. They have used five centrifugation cycles to completely remove unbound PNIPAM. Interestingly, only with the absence of free polymer, the polymer-capped AuNPs do not aggregate above the LCST.<sup>322</sup> If the LCST-induced aggregation can be avoided, the LCST transition of the polymer can be applied as colloidal actuator to switch the length of the polymer in the hierarchical core-satellite nanostructures.<sup>154,168,171,174</sup> Furthermore, the switch of the surface property can also trigger colloidal self-assembly of NPs into ordered structures.<sup>323,324</sup> Polymer possessing upper critical solution temperature (UCST) behavior<sup>306,325</sup> is also a topic of interest for nano-applications.<sup>326</sup> Especially polymer simultaneously possesses both LCST and UCST (*e.g.*, using block-copolymer approach) offers very interesting dual-responsiveness: (I) LCST < UCST (insoluble temperature window).<sup>327</sup> (II) LCST > UCST (soluble window).<sup>113,328</sup> Section 3.2.3 will give some detailed examples for the stimuli-responsive hybrid nanocomposites.

The synergies of thermoresponsive polymer brushes with its nanocore opened up possibilities to induce temperature change not only by simple heating, but also through other stimuli. Recent works have used nanocores to produce heat from *e.g.*, NIR light,<sup>114,160,167,283</sup> magnetic fields,<sup>187</sup> and ultrasound.<sup>277</sup> The highlight in these works includes the simul-





taneous drug release from the polymer shell: upon application of these external stimuli, the thermal therapy and drug therapy can be induced simultaneously.

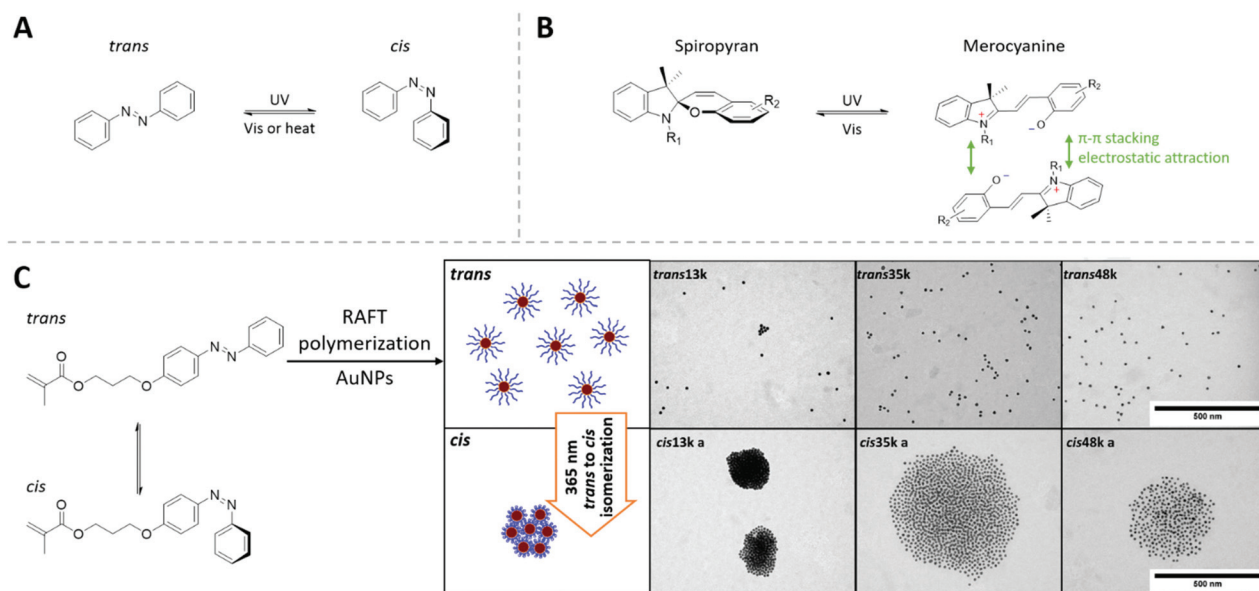
**3.1.3. pH-Responsive polymer shells.** In drug delivery applications, pH-responsiveness has been frequently introduced to the drug carrier due to the change in pH in diverse environments. The bloodstream has a pH of 7.4, while the extracellular regions of a tumor have a value of 6.4–6.8. The pH value even drops below 5 in lysosomal organelles.<sup>329</sup> This change of pH can be used as biological stimulus to achieve targeted drug release. There are two popular approaches to introduce pH-responsiveness in the polymer chain. One is introducing ionizable groups, typically acid or base groups. Among them, the tertiary amine is particularly popular for its simple preparation and tunable  $pK_a$  value for flexible application scenarios.<sup>330</sup> The other option is adding pH-responsive (*e.g.*, acid-labile) linkages between polymer and the drugs.<sup>138,265</sup> The fabrication principle and modern development of pH-responsiveness including its special detailed design for a wide range of biomedical applications have been reviewed recently in detail elsewhere.<sup>16,331,332</sup>

RAFT polymerization technique also offers effective designs for creating pH-responsive polymer-functionalized core-shell nanostructures. Pourjavadi *et al.* demonstrated a very good example of the combination of pH and thermal dual responsive magnetic drug-delivery nanohybrids. They fabricated a copolymer of NIPAM and glycidyl methacrylate (GMA) monomers on the surface of silica coated  $Fe_3O_4$  NPs *via* surface-initiated RAFT polymerization and introduced acid-labile hydrazine moieties on the epoxide ring of the GMA monomer. Doxorubicin (DOX) is then bound onto the hydrazine moieties

which will be released at lower pH values (<5.4). The advantage of the double responsiveness is that the LCST transition of PNIPAM at higher temperatures helps to further “squeeze out” the remaining DOX on the GMA units and accomplishes a high drug release in a short time.<sup>265</sup>

Sahoo *et al.* also introduced pH and thermal dual responsive polymer shells onto  $Fe_3O_4$  NPs. They first synthesized a BCP of PNIPAM and polyacrylic acid (PAA) block. The polymer is immobilized on the surface of amine-functionalized  $Fe_3O_4$  NPs using carboxyl groups from the PAA block. DOX is then introduced onto the polymer shell using the charge interaction between positively charged DOX and negatively charged PAA. At a lower pH value (<5.0), the PAA block gets protonated causing the release of DOX and the LCST transition of the PNIPAM block at higher temperatures further induces a more complete drug release performance.<sup>268</sup>

**3.1.4. Light-switchable polymer shells.** Photo-switchable molecules allow for reversible isomerization between different structural isomers upon irradiation with light of different wavelengths. This is often accompanied by a change in properties such as dipole moment or color. The most commonly studied photoswitches are azobenzenes and spiropyrans. Their switching mechanisms are illustrated in Fig. 7. The UV light-induced isomerization of azobenzene has a low polar (1.2 D) *trans* state and a high polar (4.9 D) *cis* state and the back-switch can be reversibly achieved by irradiation with visible light and heating.<sup>333</sup> This significant change in the dipole moment leads to a strong shift in the hydrophobicity (Fig. 7A). Spiropyran derivatives can be photoswitched between a spirocyclic form (colorless) and a merocyanine form (colored) with a break of the C–O bond. The merocyanine form has a conju-



**Fig. 7** (A) The light-switchable *cis*-to-*trans* transition of azobenzene molecules. (B) The photon-switching mechanism of spiropyran derivatives between its spiropyran form and merocyanine form, as well as the attractive interaction between two merocyanine units. (C) *Cis* and *trans* azobenzene monomer and the light-switchable aggregation of AuNPs mediated by azobenzene-containing RAFT polymer. Adapted with permission from ref. 166. Copyright 2016 Elsevier Ltd.



gated  $\pi$  system and possesses a zwitterionic mesomere: this structure offers a strong attractive interaction between two merocyanine units by a combination of  $\pi$ - $\pi$  stacking and electrostatic attraction (Fig. 7B). In this way, the attractive interaction between spiropyrans can be switched by light irradiation.<sup>334</sup>

The light-switchable solubility and interaction can be introduced onto the surface of the NPs *via* small molecules containing these light-switchable units. Upon irradiation, these NPs tend to self-assemble into aggregates with controlled size ("supraspheres") in suitable solvent conditions. The disassembly of these aggregates can be achieved *via* the switch-back process with the NPs returning to their dispersed state.<sup>335,336</sup>

The photoswitching property can be introduced to polymers, *e.g.*, our group has demonstrated the fabrication of AuNPs covered with photoresponsive polymer shells made by azobenzene-containing methacrylic monomers using RAFT polymerization. In toluene, the light-induced *cis*-to-*trans* transition varied the interaction between polymer-capped AuNPs and induces the controlled aggregation of AuNPs. The tunable chain length of grafted polymer brush brought the advantage to control the interparticle-distance in a very straightforward manner (Fig. 7C).<sup>166</sup>

Zhang *et al.* synthesized AuNPs covered with spiropyran-containing polymers by surface-initiated ATRP polymerization. These nanostructures also self-assemble into AuNP oligomers upon light irradiation. Since these AuNPs are brought in very close range, their surface plasmonic coupling provides a strong surface-enhanced Raman (SER) signal in the oligomer state.<sup>337</sup>

**3.1.5. Fluorescence energy transfer controlled by surface-bound polymer brushes.** The interaction of nanomaterials with fluorescent events can result in a very sensitive change of fluorescent performance and thus be used for sensor applications. Both significant enhancement and complete quenching can be achieved by controlling the energy transfer between fluorophore and nanomaterial. There are varieties of energy transfer processes, such as Förster resonance energy transfer (FRET),<sup>338–340</sup> nanometal surface energy transfer (NSET),<sup>341–343</sup> and plasmon resonance energy transfer (PRET),<sup>343–345</sup> which are widely used for tuning the fluorescence performance and building sensor applications.<sup>343,346</sup> Although the detailed physical process behind these energy transfer processes are different, the principle for building the donor-acceptor pair for energy transfer is very similar.

The first requirement here is the overlap between donor emission and acceptor absorption. In general, materials with tunable and broad absorption and emission spectra are good candidates for energy transfer-based applications. Nobel metal nanocrystals, *e.g.*, gold and silver nanocrystals, have broad plasmonic bands which are tunable by changing their size and aspect ratio, and thus can be used as super quenching unit.<sup>347</sup> The unique electronic properties of graphene and MoS<sub>2</sub> also make them perfect quenching materials.<sup>338,348–350</sup> The fluorescent NPs including semiconductor QDs, graphene QDs (GQDs) and UCNPs have strong, broad and tunable absor-

bance and luminance, which make them good candidates as both donors<sup>338,351</sup> and acceptors.<sup>352,353</sup>

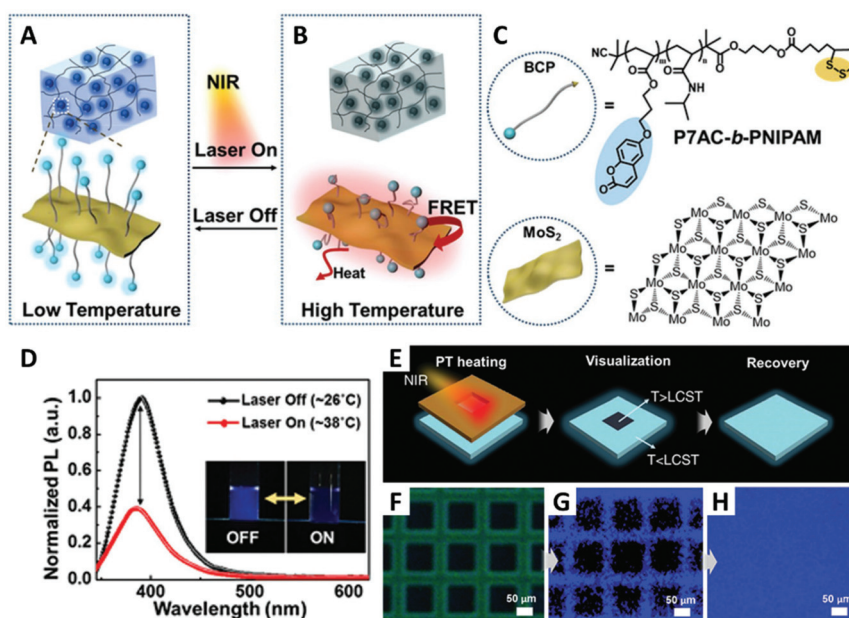
The energy transfers process between the donor and acceptor is strongly dependent on their distance. In case of FRET, an efficient energy transfer only takes place when the distance between donor and acceptor is smaller than 10 nm.<sup>342</sup> This makes nanomaterial-induced energy transfer perfect candidates as sensors to indicate near-field changes.

Smart polymers have the ability to change their size towards certain stimuli, for example, PNIPAM has a significantly shrunk size above its LCST. These stimuli-responsive polymers can be built between the donor and acceptor molecules, using their transition to switch on and off the fluorescence. Kim and coworkers have demonstrated a series of smart fluorescent nanosensors using diverse nanomaterials based on fluorescence energy transfer. These nanosensors are capable to detect thermal,<sup>110,288,354</sup> photothermal,<sup>114</sup> pH,<sup>287,288</sup> and metal ion<sup>288</sup> changes due to a stimuli-responsive RAFT polymer linker.

In their recent publication, they used MoS<sub>2</sub> as acceptor (strong FRET quencher) and a RAFT BCP containing a thermo-responsive PNIPAM block and a fluorescent poly(7-(4-(acryloyloxy)butoxy)coumarin) block (P7AC, emits blue light at ~392 nm) as donor (Fig. 8). After RAFT polymerization, dithiol-sulfide moieties were introduced to the BCPs on their chain ends (*R*-group), which provide strong anchoring on the surface of ultrathin MoS<sub>2</sub> nanosheets. The BCP brush-capped MoS<sub>2</sub> was then fabricated by a simple grafting-to experiment with the PNIPAM block positioned between the MoS<sub>2</sub> nanosheet and the fluorescent block. MoS<sub>2</sub> has very high absorption coefficient for both the visible and NIR region. Upon NIR irradiation, the photothermal ability of MoS<sub>2</sub> induces the LCST transition of the PNIPAM block. After LCST transition, the PNIPAM block contracts and pulls the P7AC block close to MoS<sub>2</sub> causing FRET quenching (reduced fluorescence intensity). They then dispersed BCP-MoS<sub>2</sub> nanohybrids in a hydrogel and demonstrated the high-resolution (sub-micrometer) fluorescence thermal mapping during photothermal treatment.<sup>114</sup>

In another example, they have introduced three types of BCPs with blue, green, or red fluorescent polymer blocks (donor) onto the surface of graphene (acceptor/quencher) *via* pyrene anchoring groups (by a strong  $\pi$ - $\pi$  stacking interaction) (Fig. 9). The three different fluorophore blocks are copolymerized with three thermal responsive polymer blocks with different LCSTs, respectively. The LCST of the thermo-responsive block was tuned to be 27, 32 and 39 °C for blue, green, red, respectively. The precise adjustment of LCST for PNIPAM is achieved by copolymerizing PNIPAM with different monomers. After the BCP is anchored to the surface of graphene, the PNIPAM block acts as thermoresponsive spacer between fluorophore donor and acceptor: by increasing temperature, the LCST is surpassed, resulting in PNIPAM block shrinkage, bringing the fluorescent block close to graphene and causing fluorescence quenching. In this way, the intensity of the three fluorescent colors represents three different LCSTs





**Fig. 8** (A and B) Schematic illustration of the BCP-functionalized MoS<sub>2</sub> nanosheets in hydrogel and the on/off transition controlled by NIR laser irradiation. Upon NIR irradiation, the PNIPAM block collapses due to the rise of local temperature, and the fluorescence of the P7AC block is quenched by MoS<sub>2</sub>. (C) Structure of P7AC-*b*-PNIPAM and MoS<sub>2</sub>. (D) The change of photoluminescence spectra of BCP-MoS<sub>2</sub> nanocomposites by NIR light-induced heating. (E) Illustration of BCP-MoS<sub>2</sub>-film upon NIR irradiation through a photomask. (F) Optical microscope image of the BCP-MoS<sub>2</sub>-film visualized using the photomask. (G) Fluorescence image of selective local heating by NIR irradiation. (H) Recovered fluorescence image at 2 min after laser shutdown. Adapted with permission from ref. 114. Copyright 2017 WILEY-VCH Verlag GmbH & Co. KGaA.

and offers sensing within a wide range of temperature between 25 to 45 °C.<sup>110</sup>

Furthermore, they exchanged the quenching GO nanosheet with green emitting QDs. They used surface-initiated RAFT polymerization and synthesized PNIPAM-*b*-P7AC copolymers on QDs. The QD serves as the acceptor for the blue emitting P7AC block. If the temperature is above the LCST, the PNIPAM block collapses and the P7AC blocks enter suitable FRET ranges. As the result, the blue intensity drops distinctly while the green emission increases.<sup>288</sup>

RAFT polymers with pH-responsive conformation change, *i.e.*, PAA and poly(2-vinylpyridine), can also be used as a responsive spacer in energy transfer systems. Kim and co-workers demonstrated multiple colorimetric pH sensors based on QDs (donor) and GO (quencher) with two types of QDs linked to the surface of GO using RAFT polymer linkers with distinct pH-responsiveness.<sup>287</sup>

Plasmonic noble metal NPs can also be used as quencher in energy transfer processes like NSET. Li *et al.*<sup>165</sup> and Qiu *et al.*<sup>138</sup> have both fabricated AgNPs decorated with RAFT copolymers carrying a block binding drug molecules *via* a UV or pH-cleavable bond. Furthermore, these drugs are fluorescent and both interact with AgNPs *via* NSET when they are attached to the surface-bound polymer, causing fluorescence quenching. After drug release upon UV or pH stimuli, the fluorescent drug recovers its fluorescent emission, enabling real-time monitoring of the drug release process.

In summary, we can clearly see the flexible role of stimuli-responsive RAFT polymers as tunable soft nanospacers and molecule carriers for energy transfer-related applications. The precise structural design and anchoring group positioning make RAFT polymers a perfect tool for connecting and adjusting the interaction between diverse energy donors and acceptors in a smart and dynamic manner.

### 3.2. Creating precise nanoassemblies using RAFT polymers

Recent advances are being made in the development of novel polymer-functionalized nanomaterials with the aim beyond the conventional definition of functions. The introduction of asymmetry, chirality, and interaction between each nanocomponent has triggered a very intriguing self-assembly process according to the corresponding properties. Transferring these symmetrical and directional codes from functional polymers (molecular level) to the assembly of NPs (nanoscale) requires innovations in the polymer design and a deep understanding of the colloidal interactions between polymers, solvent and nanosurfaces.

Using RAFT polymers as structured linkers, well-defined hierarchical core-satellite nanoassemblies can also be created with tunable interparticle distance and interaction. The flexible polymerization technique and topological design enable the fabrication of polymer-capped core NPs with a strong ability to capture colloidal nanosatellite particles. By building





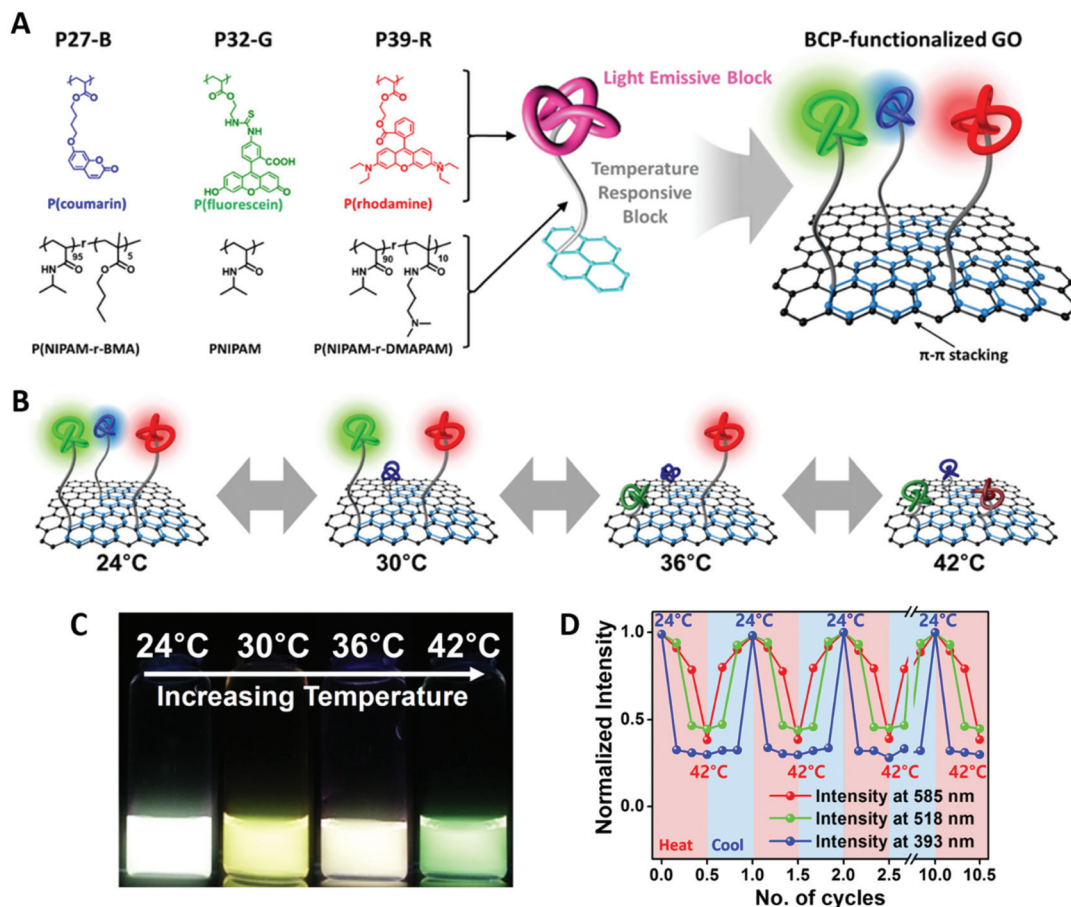


Fig. 9 (A) Structures of pyrene-functionalized RAFT BCP with different thermoresponsive and fluorescent blocks (27 °C-blue, 32 °C-green, 39 °C-red). Schematic illustration of the BCP-GO nanohybrid. (B and C) Illustration and the actual color change of BCP-GO with increasing temperature. (D) Reversibility of the thermoresponsive photoluminescent behavior at three different wavelengths of the BCP-GO nanostructure. Adapted with permission from ref. 110. Copyright 2016 American Chemical Society.

stimuli-responsiveness into the polymer linker, the core-satellite NPs can smartly switch their structure and properties.

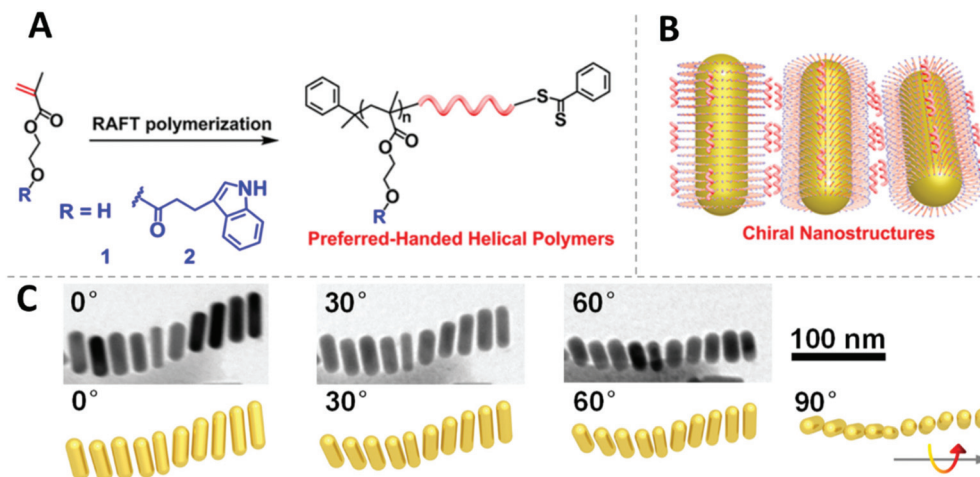
**3.2.1. Chiral self-assembly of NPs induced by RAFT polymer shells.** Plasmonic nanoassemblies with chirality have been of special interest for their exceptionally strong chiroptical properties.<sup>355,356</sup> These properties have very high potential in bioanalytical applications, for example, biosensing<sup>356,357</sup> and medical diagnosis.<sup>356</sup> The preparation of these chiral nanostructures was dominated by biomolecular linkers like amino acids,<sup>358</sup> peptides,<sup>359</sup> and DNA<sup>356,360–362</sup> due to their inherent chirality. Fabricating similar structures using non-biological polymers is a more systematic and flexible strategy, bringing access to all the functions and advantages synthetic polymers can offer.

The challenge for manipulating the chirality of nanoassemblies is to create the chiral interaction between NPs. On this point, Cheng *et al.* impressively demonstrated a method to endow AuNRs with chirality utilizing the RAFT polymers poly (methacrylate hydroxyethyl-3-indole propionate) (PIPEMA) and PHEMA (Fig. 10A).<sup>192</sup> Both polymers exhibit preferred-handed helical conformation, which could induce Au NRs to self-

assemble into a left-handed rotating side-by-side structure (Fig. 10B and C) at high pH conditions. The chiral nano-assembly of AuNRs showed strong circular dichroism signals in the Vis-NIR region. Notably, both monomers are achiral and no chiral reagents are involved in the polymerization process, the chirality is originated from the preferred-handed helical conformation of the polymer main chain caused by their high syndiotacticity and their bulky pendant side groups as demonstrated by MD simulation.

**3.2.2. Directional self-assembly of NPs with RAFT BCPs.** For the colloidal self-assembly process, BCPs have shown very flexible abilities for creating ordered nanostructures by tuning the structure of the polymer block and its interaction with the solvent.<sup>363–365</sup> Combining these features on the surface of single NPs can bring their self-assembly to the next level. The surface-bound morphological change of BCPs often induces inhomogeneous distribution of polymer brushes and the occurrence of attractive surface polymer regions (patches).<sup>169,196,366</sup> These surface-patterned polymer patches bring ordered anisotropic character to the surface of NPs which can be used as a template to guide directional self-



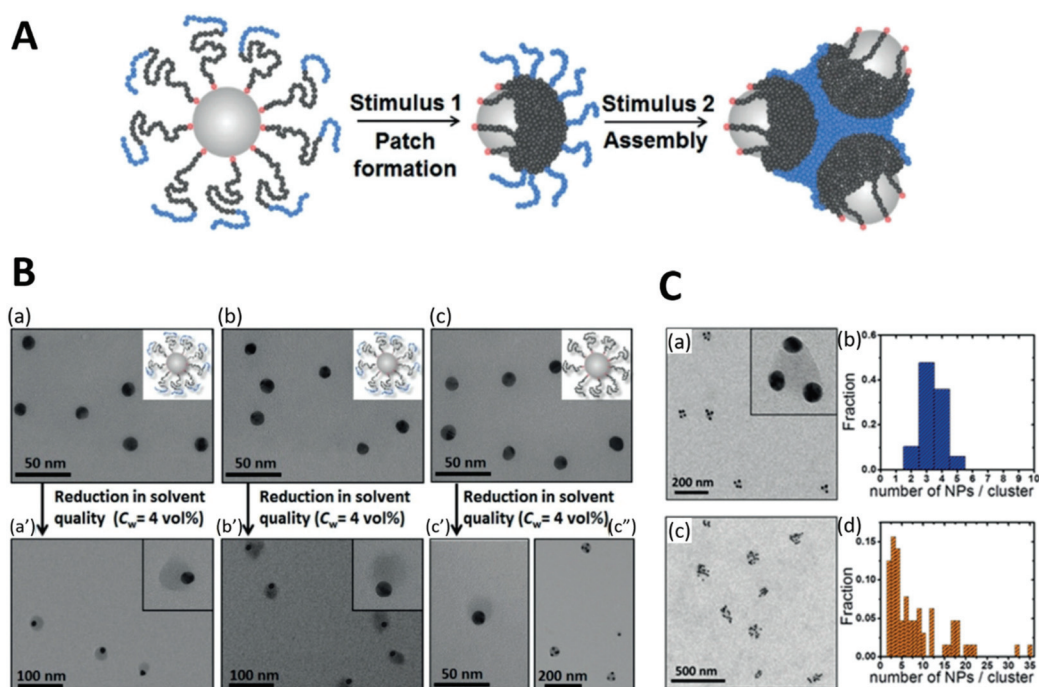


**Fig. 10** (A) Design and synthesis of the chiral RAFT polymer. (B) Illustration of chiral polymer capped AuNRs and the self-assembly of them. (C) The chiral arrangement of polymer capped AuNRs. Adapted with permission from ref. 192. Copyright 2019 American Chemical Society.

assembly processes. Furthermore, they have added some intriguing advantages in drug-delivery systems owing to their unique impact on the performance of nanocarriers.<sup>367</sup> The formation of surface polymer patches is controlled by thermodynamics: by reducing the solvent quality, the surface grafted polymer shell segregates into surface-pinned polymer patches.

On this topic, Rossner *et al.* reported an approach for fabricating patchy polymer shell-capped AuNPs using unfavorable

solvent interaction (1<sup>st</sup> stimulus) and subsequent self-assembly of these asymmetrical NPs *via* their functional polymer patches upon application of a 2<sup>nd</sup> stimulus ([Cu<sub>2</sub>(OAc)<sub>4</sub>] complex). The two-step orthogonal stimuli-responsiveness requires sophisticated design of the BCP (Fig. 11A). AuNPs were capped with RAFT poly(styrene-*block*-(4-vinylbenzoic acid)) [poly(St-*b*-4VBA)], with poly(St) as NP-adjacent block and poly(4VBA) as NP-remote block. Upon reducing solvent quality,



**Fig. 11** (A) Schematic illustration of the design principle for the stimuli-triggered surface patterning and self-assembly of AuNPs with block-copolymer ligands. (B) TEM images of AuNPs capped with BCP (a, b, distinct chain length) and poly(St) (c). a', b' and c' are the corresponding patchy NPs yielded from the poorer solvent condition. (C) Self-assembly of patchy NPs after 2<sup>nd</sup> stimulus with a statistic of the number of NPs in each cluster. Adapted with permission from ref. 169. Copyright 2019 WILEY-VCH Verlag GmbH & Co. KGaA.



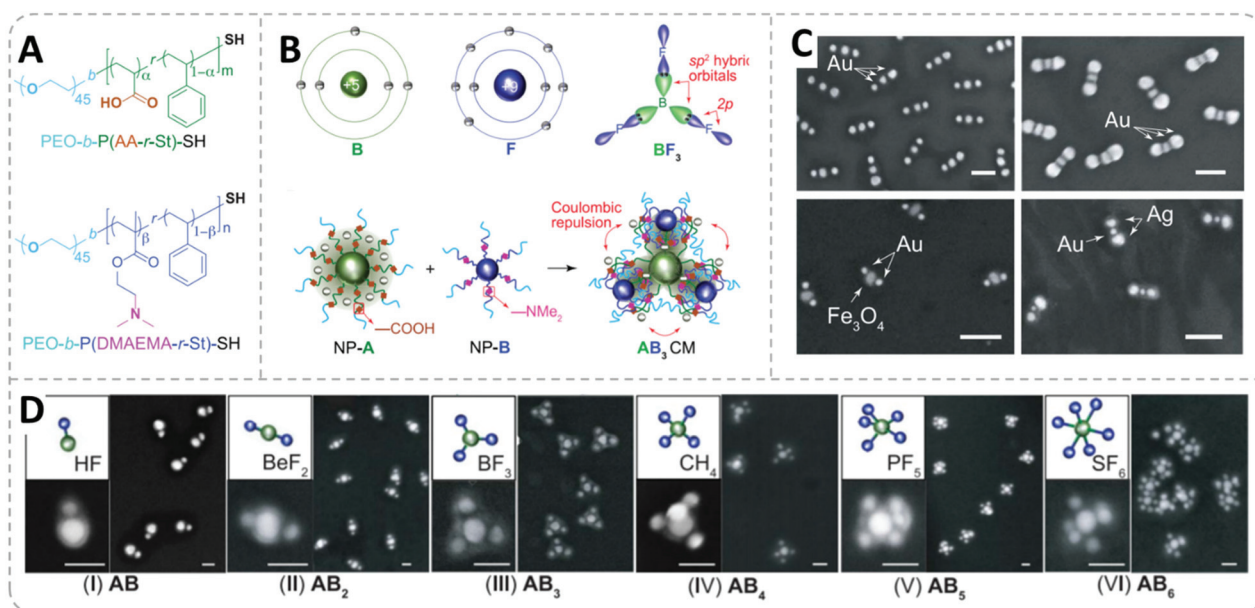
the Poly(St) block segregated to patches on the AuNP surface to minimize unfavorable polymer-solvent interaction, while the solvated Poly(4VBA) block stabilized the patchy NPs against aggregation. The subsequent self-assembly of these NPs is triggered by adding  $\text{Cu}^{2+}$  precursor complexes to the colloids: junction of patchy NPs was mediated *via* coordination of  $\text{Cu}_2^{4+}$  dimetal-ions by Poly(4VBA) blocks.<sup>368</sup> Moreover, the size and shape of the self-assembled nanocluster are highly dependent on the BCP design: the short Poly(4VBA) block leads to a small, well-defined cluster containing 3–4 NPs (Fig. 11C(a)); while longer Poly(4VBA) blocks form rather large irregularly shaped assemblies (Fig. 11C(c)).<sup>169</sup>

Yi *et al.* have impressively demonstrated the fabrication of a series of directional core-satellite-like NP assemblies utilizing the charge interaction on the grafted BCP.<sup>196</sup> In their work, gold, silver, and magnetite ( $\text{Fe}_3\text{O}_4$ ) were used as building blocks for diverse nanoassemblies. Both RAFT and ATRP polymerization were applied to fabricate the BCPs used to form dense polymer shells on the NPs: thiol-terminated BCPs synthesized from RAFT polymerization were grafted onto gold and silver surfaces *via* simply mixing them under sonication. For anchoring polymers onto  $\text{Fe}_3\text{O}_4$ , phosphonate-terminated BCP was synthesized from ATRP polymerization.

To simultaneously introduce complementary reactive bonding between core and satellite NPs and the repulsive interaction between the charged polymer patches bonds, an acid-base neutralization reaction between two types of NPs carrying polymer brushes with opposite acid-base groups was applied. Those acid/base functionalities are tuned with the

build-in moieties in the BCP using acrylic acid (AA) as acid and *N,N*-dimethylaminoethyl methacrylate (DMAEMA) as the base monomer. The molecular design of these acid/base polymers is illustrated in Fig. 12A, which is comprised of two blocks: the first polymer block consists of a statistic copolymer of acid/base and styrene monomers, the ratio of both monomers in this block was kept constant (~30%) while the chain length is varied to adjust the content of the acid/base moieties in the polymer shell. The second PEO block is acting as a steric stabilizer of the NPs.

Once the acid copolymer-capped NPs are mixed with the base copolymer-capped NPs, the interparticle neutralization reaction takes place. The neutralization yields a strong ionic attractive interaction which provides a powerful binding effect between the two types of NPs, resulting in “colloidal molecules” nanoassemblies (Fig. 12B). The charged polymer regimes also confer a repulsing interaction between the polymer brushes of adjacent satellite NPs, thus leading to a highly ordered geometry (Fig. 12C and D). Here, the Coulomb repulsion can be analogously compared with the valence electron pair model which can predict the geometry of simple molecules in  $\text{AB}_x$  form. Now the same principles and rules can be applied to the “colloidal molecules” nanoassemblies. To adjust the number of satellite NPs in each nanostructure, they varied the chain length of the polymer shell of the satellite. Since the ratio of the DMAEMA is kept the same in the polymer chain, a shorter polymer brush chain contains fewer base groups which resulted in a smaller neutralization capacity of the satellites, which then allowed the acid core NP to catch



**Fig. 12** (A) Molecular structure of the BCPs used for surface modification of AuNPs and AgNPs. (B) Schematic illustration of an  $\text{sp}^2$ -hybridized  $\text{BF}_3$  molecule. Illustration of directional bonding of NP-A and NP-B and the formation of an  $\text{AB}_3$  structure by the stoichiometric reaction between the complementary reactive polymer shells. (C) SEM images of  $\text{AB}_2$  nanoclusters formed from diverse types of NPs, scale bar = 100 nm. (D) SEM images of  $\text{AB}_x$  structures ( $x = 1$  to 6) assembled from 36 nm AuNPs (A) and 20 nm AuNPs (B). The change in the number and geometry of the satellite NPs is controlled by increasing the chain length of the poly(DMAEMA-*r*-St) block. Adapted with permission from ref. 196. Copyright 2020 American Association for the Advancement of Science.





more satellite NPs. As a result of the interaction brought by the reactive polymer shell, the 3D structure of the nano “colloidal molecules” including the arrangement and the number of the satellite particles can be precisely tuned by only engineering the chain length of the surface BCP brush.

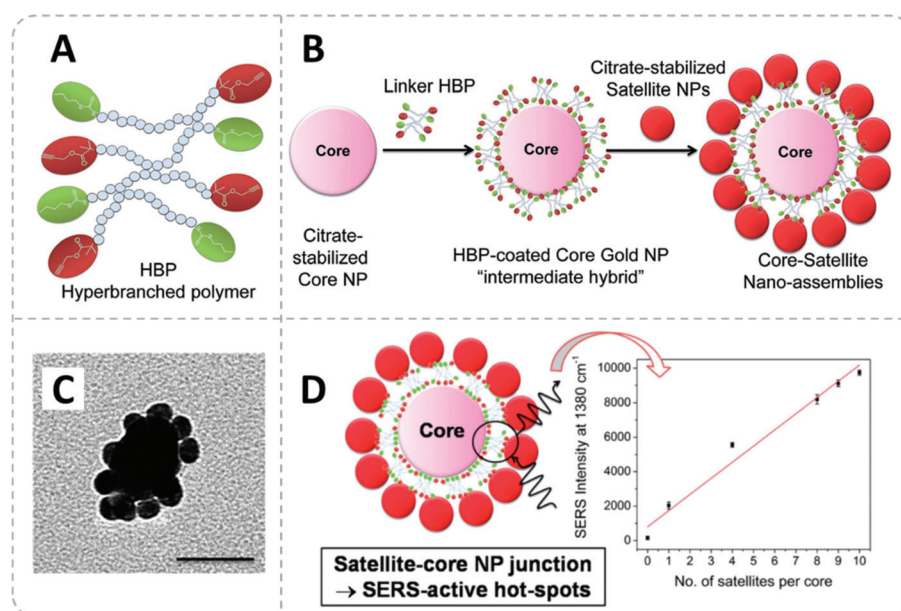
**3.2.3. Core-satellite nanoassemblies by RAFT polymer linkers.** Core-satellite (or planet-satellite) nanostructures have attracted enormous research interest in material science: simultaneously integrating distinct nanomaterials into one well-defined nanostructure often gives unique hybrid characteristics for novel applications in different fields including multimodal bioimaging,<sup>369,370</sup> cancer treatment,<sup>369,371</sup> SER spectroscopy,<sup>207,372</sup> fluorescence enhancement,<sup>373</sup> *etc.* To build such structures, planet and satellite NPs must be bound with a proper linker to form a colloiddally stable nanostructure. For the task of accomplishing a precise assembly of the planet-satellite nanostructure, various synthetic approaches based on small molecular linkers,<sup>374–376</sup> polymer linkers,<sup>109,113,154,168,175,207</sup> and DNA conjugation<sup>369,373,377,378</sup> have made significant progress recently.

Choosing appropriately designed synthetic polymers as a linker brings multiple advantages such as enforced colloidal stability owing to the steric shielding of a polymer shell (as we have seen in section 3.1), the tunable interparticle distance between planet-satellite,<sup>109,168,175</sup> and last but not least, its high efficiency and low cost to produce. As a linker for nanomaterials, the design flexibility in the macromolecular structure including topology, end-group functionality and the choice of monomers/BCPs offers very open access for creating hierarchical nanostructures with desired functions.

In general, the number of free anchoring end groups for satellite particles on the polymer-capped core NP surface is the key for the self-assembly process. For the case that both core and satellite are noble metal NPs, the multi-arm polymer with anchoring groups at each chain termini is a widely used approach. Dey *et al.* demonstrated a practical example to increase the free end groups using hyperbranched polymers carrying multiple anchoring RAFT and alkyne end groups (Fig. 13).<sup>207</sup> The nanostructure made of AuNPs presents a well-defined structural arrangement with a tunable number of satellite NPs. Due to both the structural nature of the hyperbranched polymer and its low molecular weight ( $\sim 10$  kg mol<sup>-1</sup>), the interparticle distance between core and satellites was very short. This close distance between the NPs induced a strong SER effect which can be easily tuned by the number of attached satellite AuNPs (Fig. 13D).

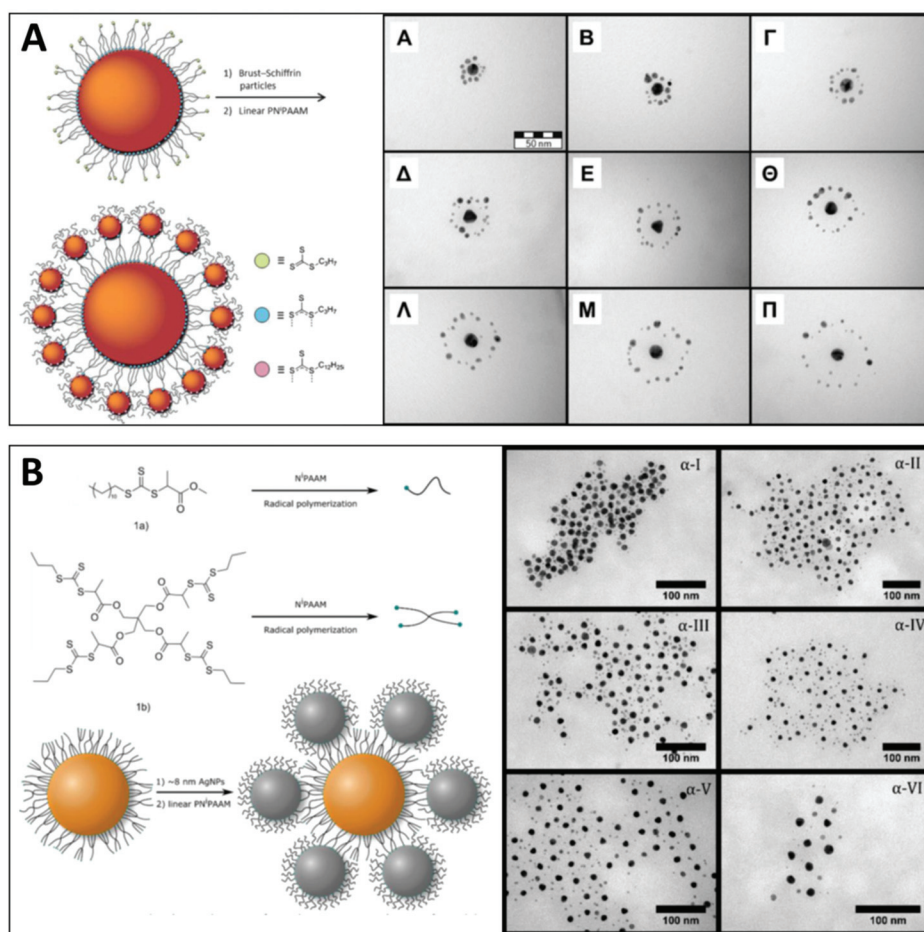
To form a connection between two NPs, at least two end groups are required. However, if both end groups have attached to the surface of the core NP, the linking effect becomes unavailable: linear bi-end-functionalized polymers have shown diminished performance of forming core-satellite nanostructures compared with star polymers containing more than three arms.<sup>379</sup> Gooding and coworkers have reported a method to first saturate the planet AuNPs with thiol end-functionalized polymers carrying a carboxylic acid group on the other terminus.<sup>113,156</sup> An additional step was applied to convert the outer carboxylic acid group into thiol groups which allows for self-assembly of additional satellite AuNPs.

Our group has reported a method to prepare noble metal core-satellite nanostructures based on multi-arm star PNIPAM



**Fig. 13** (A) Structural design of hyperbranched polymer. (B) Schematic illustration of the fabrication strategy of core-satellite nanostructures using hyperbranched polymer linkers. (C) TEM micrograph of core-satellite nanostructure with a satellite/core ratio of 10. Scale bar = 50 nm. (D) Increasing SER scattering intensity with growing number of satellites per core. Adapted with permission from ref. 207. Copyright 2014 The Royal Society of Chemistry.





**Fig. 14** (A) Synthetic strategy (left) and TEMicrograph of gold-planet–gold-satellite nanostructures. Adapted with permission from ref. 109. Copyright 2014 WILEY-VCH Verlag GmbH & Co. KGaA. (B) Structure of star and linear RAFT agents, synthetic scheme, and TEMicrograph of gold-planet–silver-satellite nanostructures. Adapted with permission from ref. 175. Copyright 2016 American Chemical Society.

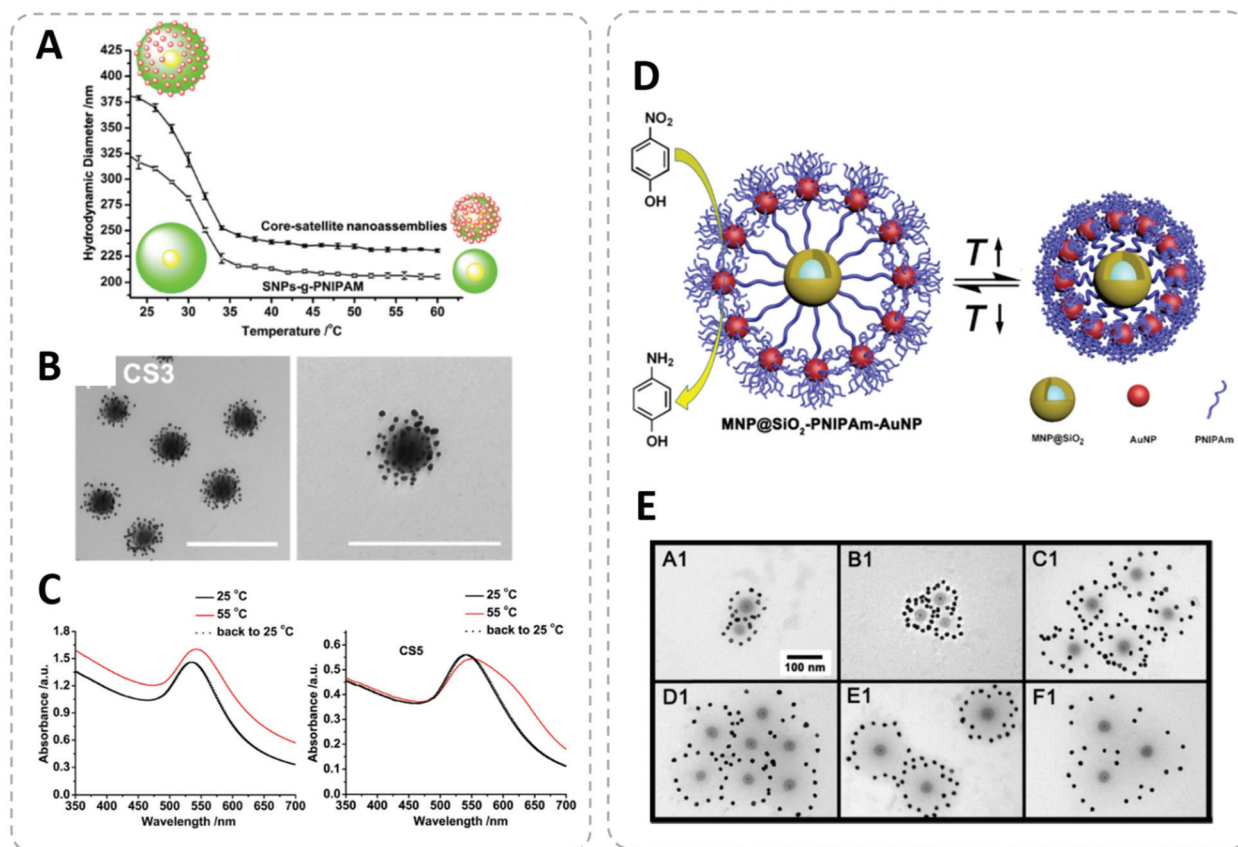
with RAFT end groups as linkers (Fig. 14).<sup>109,175</sup> The planet is constructed by self-assembly of star RAFT polymers onto citrate-capped AuNPs using the grafting-to approach. The free (not surface-bound) polymer arms of the grafted polymer can capture satellite NPs.<sup>379</sup> The same planet nanostructure is able to capture both gold (Fig. 14A)<sup>109</sup> and silver (Fig. 14B)<sup>175</sup> satellite NPs. Notably, the well-defined structure of the star polymer offers full control over the arrangement of the satellite nano-components: the average distance between planet satellites NPs can be precisely tuned between approximately 3 to 20 nm by varying the molecular weight of the linker.

Sulfur-terminated polymers with suitable topological designs have been proven to be a powerful platform for the self-assembly of hierarchical nanostructures made from noble-metal nanomaterials. However, for constructing structures with non-metal NPs, other approaches are inevitably required. As building material for hierarchical nanostructures, silica brings very intriguing potential owing to the intensively studied silica coating methods which allow almost every nanomaterial to be introduced as a planet. To bring polymer brushes onto the silica surface, the well-established silane

chemistry offers a great variety of protocols.<sup>380,381</sup> However, creating well-defined silica planets with a dense polymer shell requires an effective route and careful treatment of silica NPs for their tendency to form aggregation. For this task, the surface-initiated RAFT polymerization *via* grafting-from has been proven to be suitable.<sup>95,99,264</sup> By designing the surface-initiated RAFT polymerization using an *R*-approach, the RAFT moieties will be positioned at the outer termini of the polymer shell which are capable for attaching to satellite NPs. Based on this principle, Wu *et al.* and Tian *et al.* have both reported protocols for fabricating AuNPs decorated silica-RAFT PNIPAM core-satellite nanostructures (Fig. 15).<sup>154,168</sup> The nanoassemblies showed a well-defined structure with full control over the number of satellite NPs<sup>154</sup> and interparticle distance.<sup>168</sup> Tian *et al.* also demonstrated the access of introducing further  $\text{Fe}_3\text{O}_4$  NPs in the core position using the silica coating technique.<sup>168</sup> These complicated nanostructures can be fabricated using a simple synthetic method and a very efficient route.

Due to the close distance between the NPs brought by the polymer linker, the nanostructure often shows a strong surface plasmonic coupling between noble metal NPs, which has the





**Fig. 15** (A) Temperature dependence of the hydrodynamic diameter of PNIPAM-functionalized silica-core-gold-satellite nanoassemblies and PNIPAM-capped silica NPs (core). (B) TEM micrograph of silica-core-gold-satellite nanoassemblies. (Scale bar = 500 nm) (C) UV-vis spectra of silica-core-gold-satellite nanostructures at different temperatures. The structure for the right spectrum has a higher gold/silica ratio. The additional absorption originates from the surface plasmonic coupling in the shrunken state. A–C: Adapted with permission from ref. 154. Copyright 2017 WILEY-VCH Verlag GmbH & Co. KGaA. (D) Schematic illustration of the fabrication strategy of  $\text{Fe}_3\text{O}_4\text{NP@silica-PNIPAM-AuNPs}$  nanostructures. (E) TEM micrograph of  $\text{Fe}_3\text{O}_4\text{NP@silica-PNIPAM-AuNPs}$  nanostructure with increasing molecular weight of the PNIPAM linker. D–E: Adapted with permission from ref. 168. Copyright 2019 American Chemical Society.

potential to be used for SER applications.<sup>113,207</sup> Moreover, a stimuli-responsive polymer linker, *e.g.*, PNIPAM, also brings its ability to these core-satellite nanostructures as a thermo-responsive colloidal actuator reversibly switching the structure between hydrated and shrunken state.<sup>113,154,171</sup> The temperature-triggered switch of the interparticle distance in the nanostructure also induces a shift in the optical properties of the colloid, which may render them as flexible candidates for nanosensors.

Notably, the versatile silica-coating technique enables a wide choice of nanomaterials for the role of the core and makes this strategy a general protocol to fabricate core-satellite nanostructures with tunable distance and interactions.

## 4. Conclusions and outlook

It was demonstrated that RAFT polymers can nicely be used to decorate and/or connect nanoparticles of different nature in order to arrive at complex and functional nanocomposites.

The success of RAFT polymerization in this field is clearly due to the great versatility in terms of polymerization reaction conditions as well as in terms of polymer-to-surface grafting strategies. It has for instance been shown that RAFT polymer with its thiocarbonylthio-moieties as reactive end-groups can often directly be used in grafting-to approaches without preceding end-group transformation. This clearly simplifies nanoengineering with these reactive building blocks. Another great advantage of RAFT is its good tolerance against water-based systems, which is key for bio-related applications.

While we observe that RAFT polymers and the nanocomposites formed from them are constantly finding new fields of application, the question arises as to what the next big goals are in order to achieve a higher level in quality and function. While we can't see into the future, we think the following goals will be relevant and hopefully solved in the near future:

1. The combination of different types of polymers and different types of particles within one nanocomposite will become important. Some first initial work into this direction





has been performed, but increasing the control over different types of nanocomponents will enhance the functionality. By tuning the interplay of components, *e.g.*, *via* different stimulus sensitivity and or *via* orthogonal (surface) chemistry, the functionality of such nanohybrids can be greatly enhanced and more than one function, *e.g.*, imaging and disease treatment at the same time, may become possible.

2. In the early days of this scientific field, polymer was mainly applied to the surface of inorganic materials using grafting-from methods. However, progress in surface chemistry using polymers has shown that similarly good end results can be achieved with grafting-to methods, while the synthetic effort is greatly reduced. With this synthetic simplification, however, the complexity of the overall system can be increased at the same time while the overall expenditure remains the same. These functional nanohybrids, after all, need to remain accessible in short times and with an acceptable spending of resources. Efforts must therefore be made to further simplify the grafting-to methods for a large number of surfaces, including those that are less reactive. An effective grafting-to method for silica surfaces would, *e.g.*, be a huge step forward.

3. Most of the studies so far have addressed the action of these functional nanocomposites in solution, where they have been formed. Little is known so far about the behavior of these nanoarchitectures when cast to a surface or how their function can be exploited in a dry bulk material state. Ordering behavior and regularity control in the dry state will be future challenges to exploit these nanohybrids in the design of novel functional materials.

It is fascinating to see, how RAFT polymerization evolved during the last years from a chain-transfer effect that is capable of controlling radical polymerization to a powerful tool for constructing functional nanocomposites. It is very likely, that this is not the end of the story and that many exciting new developments using RAFT polymers will emerge.

## Abbreviations

AA	Acrylic acid
ATRP	Atom transfer radical polymerization
BCP	Block copolymer
CNT	Carbon nanotube
CSIRO	Commonwealth scientific and industrial research organization
CTA	Chain transfer agent
CTAB	Cetyltrimethylammonium bromide
DBM	Dibromomaleimide
DMAEMA	<i>N,N</i> -Dimethylaminoethyl methacrylate
DOX	Doxorubicin
FRET	Förster resonance energy transfer
GMA	Glycidyl methacrylate
GO	Graphene oxide
GQD	Graphene quantum dot
IONP	Iron oxid nanoparticle
LA-HEMA	Lipoic acid-substituted hydroxyethyl methacrylate

LCST	Lower critical solution temperature
MPC	Methacrylamide phosphorylcholine
NHS	<i>N</i> -Hydroxysuccinimide
NIR	Near-infrared
NP	Nanoparticle
NR	Nanorod
NSET	Nanometal surface energy transfe
P7AC	Poly(7-(4-(acryloyloxy)butoxy)coumarin)
PA	Phosphonate acrylate
PAA	Polyacrylic acid
PEG	Polyethylene glycol
PHEMA	Poly(2-hydroxyethyl methacrylate)
PIPEMA	Poly(methacrylate hydroxyethyl-3-indole propionate)
PNIPAM	Poly( <i>N</i> -isopropylacrylamide)
POEGA	Poly(oligoethylene glycol) methyl ether acrylate
Poly (4VBA)	Poly(4-vinylbenzoic acid)
Poly(St)	Poly(styrene)
PRET	Plasmon resonance energy transfer
PVP	Polyvinylpyrrolidone
QD	Quantum dot
RAFT	Reversible addition–fragmentation chain-transfer
RDRP	Reversible deactivation radical polymerization
SER	Surface-enhanced Raman
TEOS	Tetraethylorthosilicate
TOAB	Tetraoctylammonium bromide
UCNP	Upconverting nanoparticle

## Conflicts of interest

There are no conflicts to declare.

## Acknowledgements

Financial support of our work by the Fonds der Chemischen Industrie is gratefully acknowledged.

## References

- 1 J. Chieffari, Y. K. B. Chong, F. Ercole, J. Krstina, J. Jeffery, T. P. T. Le, R. T. A. Mayadunne, G. F. Meijs, C. L. Moad, G. Moad, E. Rizzardo and S. H. Thang, *Macromolecules*, 1998, **31**, 5559–5562.
- 2 S. Perrier, *Macromolecules*, 2017, **50**, 7433–7447.
- 3 D. J. Keddie, G. Moad, E. Rizzardo and S. H. Thang, *Macromolecules*, 2012, **45**, 5321–5342.
- 4 G. Moad, E. Rizzardo and S. H. Thang, *Polym. Int.*, 2011, **60**, 9–25.
- 5 A. Skandalis, T. Sentoukas, D. Giaouzi, M. Kafetzi and S. Pispas, *Polymers*, 2021, **13**, 1698.
- 6 D. J. Keddie, *Chem. Soc. Rev.*, 2014, **43**, 496–505.



- 7 M. Semsarilar and V. Abetz, *Macromol. Chem. Phys.*, 2021, **222**, 2000311.
- 8 J. M. Ren, T. G. McKenzie, Q. Fu, E. H. H. Wong, J. Xu, Z. An, S. Shanmugam, T. P. Davis, C. Boyer and G. G. Qiao, *Chem. Rev.*, 2016, **116**, 6743–6836.
- 9 D. Kuckling and A. Wycisk, *J. Polym. Sci., Part A: Polym. Chem.*, 2013, **51**, 2980–2994.
- 10 Y. Zheng, S. Li, Z. Weng and C. Gao, *Chem. Soc. Rev.*, 2015, **44**, 4091–4130.
- 11 S. I. Bhat, Y. Ahmadi and S. Ahmad, *Ind. Eng. Chem. Res.*, 2018, **57**, 10754–10785.
- 12 Z. Li, M. Tang, S. Liang, M. Zhang, G. M. Biesold, Y. He, S.-M. Hao, W. Choi, Y. Liu, J. Peng and Z. Lin, *Prog. Polym. Sci.*, 2021, 101387.
- 13 G. Moad, *Polym. Chem.*, 2016, **8**, 177–219.
- 14 C. Lu and M. W. Urban, *Prog. Polym. Sci.*, 2018, **78**, 24–46.
- 15 C. Zhao, Z. Ma and X. X. Zhu, *Prog. Polym. Sci.*, 2019, **90**, 269–291.
- 16 N. Deirram, C. Zhang, S. S. Kermaniyan, A. P. R. Johnston and G. K. Such, *Macromol. Rapid Commun.*, 2019, **40**, 1800917.
- 17 B. D. Fairbanks, P. A. Gunatillake and L. Meagher, *Adv. Drug Delivery Rev.*, 2015, **91**, 141–152.
- 18 A. Zhang, K. Jung, A. Li, J. Liu and C. Boyer, *Prog. Polym. Sci.*, 2019, **99**, 101164.
- 19 M. Ahmed and R. Narain, *Prog. Polym. Sci.*, 2013, **38**, 767–790.
- 20 P. Gurnani and S. Perrier, *Prog. Polym. Sci.*, 2020, **102**, 101209.
- 21 X. Jin, P. Sun, G. Tong and X. Zhu, *Biomaterials*, 2018, **178**, 738–750.
- 22 R. Yang, X. Wang, S. Yan, A. Dong, S. Luan and J. Yin, *Prog. Polym. Sci.*, 2021, **118**, 101409.
- 23 G. Moad, E. Rizzardo and S. H. Thang, *Chem. – Asian J.*, 2013, **8**, 1634–1644.
- 24 F. D'Agosto, J. Rieger and M. Lansalot, *Angew. Chem., Int. Ed.*, 2020, **59**, 8368–8392.
- 25 S. L. Canning, G. N. Smith and S. P. Armes, *Macromolecules*, 2016, **49**, 1985–2001.
- 26 J. Yeow and C. Boyer, *Adv. Sci.*, 2017, **4**, 1700137.
- 27 M. J. Derry, L. A. Fielding and S. P. Armes, *Prog. Polym. Sci.*, 2016, **52**, 1–18.
- 28 J. O. Zoppe, N. C. Ataman, P. Mocny, J. Wang, J. Moraes and H.-A. Klok, *Chem. Rev.*, 2017, **117**, 1105–1318.
- 29 J. Gaitzsch, X. Huang and B. Voit, *Chem. Rev.*, 2016, **116**, 1053–1093.
- 30 C. K. Wong, X. Qiang, A. H. E. Müller and A. H. Gröschel, *Prog. Polym. Sci.*, 2020, **102**, 101211.
- 31 K. Nagase, T. Okano and H. Kanazawa, *Nano-Struct. Nano-Objects*, 2018, **16**, 9–23.
- 32 K. Jung, N. Corrigan, M. Ciftci, J. Xu, S. E. Seo, C. J. Hawker and C. Boyer, *Adv. Mater.*, 2020, **32**, 1903850.
- 33 Z. Zhang, N. Corrigan, A. Bagheri, J. Jin and C. Boyer, *Angew. Chem., Int. Ed.*, 2019, **58**, 17954–17963.
- 34 G. Palui, F. Aldeek, W. Wang and H. Mattoussi, *Chem. Soc. Rev.*, 2015, **44**, 193–227.
- 35 X. Huang, J. Hu, Y. Li, F. Xin, R. Qiao and T. P. Davis, *Biomacromolecules*, 2019, **20**, 4243–4257.
- 36 N. Zhao, L. Yan, X. Zhao, X. Chen, A. Li, D. Zheng, X. Zhou, X. Dai and F.-J. Xu, *Chem. Rev.*, 2019, **119**, 1666–1762.
- 37 S. O. Pereira, A. Barros-Timmons and T. Trindade, *Polymers*, 2018, **10**, 189.
- 38 X. Bouju, É. Duguet, F. Gauffre, C. R. Henry, M. L. Kahn, P. Mélinon and S. Ravaine, *Adv. Mater.*, 2018, **30**, 1706558.
- 39 M. A. Boles, M. Engel and D. V. Talapin, *Chem. Rev.*, 2016, **116**, 11220–11289.
- 40 M. Grzelczak, L. M. Liz-Marzán and R. Klajn, *Chem. Soc. Rev.*, 2019, **48**, 1342–1361.
- 41 K. Deng, Z. Luo, L. Tan and Z. Quan, *Chem. Soc. Rev.*, 2020, **49**, 6002–6038.
- 42 W. B. Rogers, W. M. Shih and V. N. Manoharan, *Nat. Rev. Mater.*, 2016, **1**, 1–14.
- 43 C. Yi, S. Zhang, K. T. Webb and Z. Nie, *Acc. Chem. Res.*, 2017, **50**, 12–21.
- 44 S. Samanta, S. L. Banerjee, K. Bhattacharya and N. K. Singha, *ACS Appl. Mater. Interfaces*, 2021, **13**, 36307–36319.
- 45 N. Penfold, J. Yeow, C. Boyer and S. P. Armes, *ACS Macro Lett.*, 2019, **8**, 1029–1054.
- 46 C. Liu, C.-Y. Hong and C.-Y. Pan, *Polym. Chem.*, 2020, **11**, 3673–3689.
- 47 X. Wang and Z. An, *Macromol. Rapid Commun.*, 2019, **40**, 1800325.
- 48 S. Y. Khor, J. F. Quinn, M. R. Whittaker, N. P. Truong and T. P. Davis, *Macromol. Rapid Commun.*, 2019, **40**, 1800438.
- 49 J. Cao, Y. Tan, Y. Chen, L. Zhang and J. Tan, *Macromol. Rapid Commun.*, 2021, 2100498.
- 50 E. J. Cornel, J. Jiang, S. Chen and J. Du, *CCS Chem.*, 2021, **3**, 2104–2125.
- 51 G. Moad, E. Rizzardo and S. H. Thang, *Aust. J. Chem.*, 2005, **58**, 379–410.
- 52 J. Cuthbert, S. V. Wanasinghe, K. Matyjaszewski and D. Konkolewicz, *Macromolecules*, 2021, **54**, 8331–8340.
- 53 U. C. Palmiero, M. Sponchioni, N. Manfredini, M. Maraldi and D. Moscatelli, *Polym. Chem.*, 2018, **9**, 4084–4099.
- 54 M. S. Messina, K. M. M. Messina, A. Bhattacharya, H. R. Montgomery and H. D. Maynard, *Prog. Polym. Sci.*, 2020, **100**, 101186.
- 55 F. L. Hatton, *Polym. Chem.*, 2020, **11**, 220–229.
- 56 M. D. Nothling, Q. Fu, A. Reyhani, S. Allison-Logan, K. Jung, J. Zhu, M. Kamigaito, C. Boyer and G. G. Qiao, *Adv. Sci.*, 2020, **7**, 2001656.
- 57 G. Moad and E. Rizzardo, *Polym. Int.*, 2020, **69**, 658–661.
- 58 M. Destarac, *Polym. Chem.*, 2018, **9**, 4947–4967.
- 59 C. Barner-Kowollik, *Handbook of RAFT Polymerization*, John Wiley & Sons, 2008.
- 60 H. Willcock and R. K. O'Reilly, *Polym. Chem.*, 2010, **1**, 149–157.
- 61 G. Moad, Y. K. Chong, A. Postma, E. Rizzardo and S. H. Thang, *Polymer*, 2005, **46**, 8458–8468.
- 62 G. Moad, E. Rizzardo and S. H. Thang, *Aust. J. Chem.*, 2006, **59**, 669–692.



- 63 G. Moad, E. Rizzardo and S. H. Thang, *Aust. J. Chem.*, 2009, **62**, 1402–1472.
- 64 G. Moad, E. Rizzardo and S. H. Thang, *Aust. J. Chem.*, 2012, **65**, 985–1076.
- 65 M. Mazloomi-Rezvani, M. Salami-Kalajahi and H. Roghani-Mamaqani, *J. Phys. Chem. Solids*, 2018, **123**, 183–190.
- 66 F.-W. Shen, K.-C. Zhou, H. Cai, Y.-N. Zhang, Y.-L. Zheng and J. Quan, *Colloids Surf., B*, 2019, **173**, 504–511.
- 67 W. Shen, Q. Qiu, Y. Wang, M. Miao, B. Li, T. Zhang, A. Cao and Z. An, *Macromol. Rapid Commun.*, 2010, **31**, 1444–1448.
- 68 C. Wu, Y. Zhou, H. Wang, J. Hu and X. Wang, *J. Saudi Chem. Soc.*, 2019, **23**, 1080–1089.
- 69 P. J. Roth, C. Boyer, A. B. Lowe and T. P. Davis, *Macromol. Rapid Commun.*, 2011, **32**, 1123–1143.
- 70 M. J. Kade, D. J. Burke and C. J. Hawker, *J. Polym. Sci., Part A: Polym. Chem.*, 2010, **48**, 743–750.
- 71 Y. K. Chong, G. Moad, E. Rizzardo and S. H. Thang, *Macromolecules*, 2007, **40**, 4446–4455.
- 72 S. Perrier, P. Takolpuckdee and C. A. Mars, *Macromolecules*, 2005, **38**, 2033–2036.
- 73 A. Postma, T. P. Davis, G. Moad and M. S. O'Shea, *Macromolecules*, 2005, **38**, 5371–5374.
- 74 M. H. Stenzel, *ACS Macro Lett.*, 2013, **2**, 14–18.
- 75 P. De, M. Li, S. R. Gondi and B. S. Sumerlin, *J. Am. Chem. Soc.*, 2008, **130**, 11288–11289.
- 76 L. Tao, C. S. Kaddis, R. R. O. Loo, G. N. Grover, J. A. Loo and H. D. Maynard, *Macromolecules*, 2009, **42**, 8028–8033.
- 77 K. L. Heredia, G. N. Grover, L. Tao and H. D. Maynard, *Macromolecules*, 2009, **42**, 2360–2367.
- 78 G. N. Grover, S. N. S. Alconcel, N. M. Matsumoto and H. D. Maynard, *Macromolecules*, 2009, **42**, 7657–7663.
- 79 L. Tao, J. Liu and T. P. Davis, *Biomacromolecules*, 2009, **10**, 2847–2851.
- 80 C. Boyer, J. Liu, V. Bulmus and T. P. Davis, *Aust. J. Chem.*, 2009, **62**, 830–847.
- 81 V. Bulmus, *Polym. Chem.*, 2011, **2**, 1463–1472.
- 82 M. Scherger, H. J. Räder and L. Nuhn, *Macromol. Rapid Commun.*, 2021, **42**, 2000752.
- 83 K. L. Heredia and H. D. Maynard, *Org. Biomol. Chem.*, 2006, **5**, 45–53.
- 84 C. Chen, D. Y. W. Ng and T. Weil, *Prog. Polym. Sci.*, 2020, **105**, 101241.
- 85 Y. Wang and C. Wu, *Biomacromolecules*, 2018, **19**, 1804–1825.
- 86 X. Liu and W. Gao, *Angew. Chem., Int. Ed.*, 2021, **60**, 11024–11035.
- 87 E. M. Pelegri-O'Day and H. D. Maynard, *Acc. Chem. Res.*, 2016, **49**, 1777–1785.
- 88 R. A. Olson, A. B. Korpusik and B. S. Sumerlin, *Chem. Sci.*, 2020, **11**, 5142–5156.
- 89 C. I. Biggs, M. Walker and M. I. Gibson, *Biomacromolecules*, 2016, **17**, 2626–2633.
- 90 J. L. M. Gonçalves, E. J. Castanheira, S. P. C. Alves, C. Baleizão and J. P. Farinha, *Polymers*, 2020, **12**, 2175.
- 91 R. Falatach, C. McGlone, M. S. Al-Abdul-Wahid, S. Averick, R. C. Page, J. A. Berberich and D. Konkolewicz, *Chem. Commun.*, 2015, **51**, 5343–5346.
- 92 M. Mazloomi-Rezvani, M. Salami-Kalajahi, H. Roghani-Mamaqani and A. Pirayesh, *Appl. Organomet. Chem.*, 2018, **32**, e4079.
- 93 J. E. Gargari, A. Shakeri, H. S. Kalal, A. Khanchi and H. Rashedi, *Polym. Adv. Technol.*, 2017, **28**, 1884–1891.
- 94 A. J. Chancellor, B. T. Seymour and B. Zhao, *Anal. Chem.*, 2019, **91**, 6391–6402.
- 95 K. Ohno, Y. Ma, Y. Huang, C. Mori, Y. Yahata, Y. Tsujii, T. Maschmeyer, J. Moraes and S. Perrier, *Macromolecules*, 2011, **44**, 8944–8953.
- 96 E. Roeven, A. R. Kuzmyn, L. Scheres, J. Baggerman, M. M. Smulders and H. Zuilhof, *Langmuir*, 2020, **36**, 10187–10199.
- 97 S. Ghasemi and S. Karim, *Mater. Chem. Phys.*, 2018, **205**, 347–358.
- 98 G. Conzatti, S. Cavalié, C. Combes, J. Torrisani, N. Carrere and A. Tourrette, *Colloids Surf., B*, 2017, **151**, 143–155.
- 99 C. Li, J. Han, C. Y. Ryu and B. C. Benicewicz, *Macromolecules*, 2006, **39**, 3175–3183.
- 100 J. C. Foster, S. C. Radzinski and J. B. Matson, *J. Polym. Sci., Part A: Polym. Chem.*, 2017, **55**, 2865–2876.
- 101 J. Moehrke and P. Vana, *Macromol. Chem. Phys.*, 2017, **218**, 1600506.
- 102 P. Eskandari, Z. Abousalman-Rezvani, H. Roghani-Mamaqani, M. Salami-Kalajahi and H. Mardani, *Adv. Colloid Interface Sci.*, 2019, **273**, 102021.
- 103 C. Boyer, M. H. Stenzel and T. P. Davis, *J. Polym. Sci., Part A: Polym. Chem.*, 2011, **49**, 551–595.
- 104 J. E. Rauschendorfer, K. M. Thien, M. Denz, S. Köster, F. Ehlers and P. Vana, *Macromol. Mater. Eng.*, 2021, **306**, 2000595.
- 105 H. Mardani, H. Roghani-Mamaqani, K. Khezri and M. Salami-Kalajahi, *Appl. Phys. A*, 2020, **126**, 251.
- 106 M. A. Macchione, C. Biglione and M. Strumia, *Polymers*, 2018, **10**, 527.
- 107 H. Liu, M. Ding, Z. Ding, C. Gao and W. Zhang, *Polym. Chem.*, 2017, **8**, 3203–3210.
- 108 X. Li, J. Iocozzia, Y. Chen, S. Zhao, X. Cui, W. Wang, H. Yu, S. Lin and Z. Lin, *Angew. Chem., Int. Ed.*, 2018, **57**, 2046–2070.
- 109 C. Rossner and P. Vana, *Angew. Chem., Int. Ed.*, 2014, **53**, 12639–12642.
- 110 J. Lee, H. Yang, C. H. Park, H.-H. Cho, H. Yun and B. J. Kim, *Chem. Mater.*, 2016, **28**, 3446–3453.
- 111 R. Qiao, L. Esser, C. Fu, C. Zhang, J. Hu, P. Ramírez-arcía, Y. Li, J. F. Quinn, M. R. Whittaker, A. K. Whittaker and T. P. Davis, *Biomacromolecules*, 2018, **19**, 4423–4429.
- 112 M. Tasso, E. Giovanelli, D. Zala, S. Bouccara, A. Fragola, M. Hanafi, Z. Lenkei, T. Pons and N. Lequeux, *ACS Nano*, 2015, **9**, 11479–11489.
- 113 F. Han, A. H. Soeriyadi and J. J. Gooding, *Macromol. Rapid Commun.*, 2018, **39**, 1800451.



- 114 C. H. Park, H. Yun, H. Yang, J. Lee and B. J. Kim, *Adv. Funct. Mater.*, 2017, **27**, 1604403.
- 115 M. Brust, D. Bethell, C. J. Kiely and D. J. Schiffrin, *Langmuir*, 1998, **14**, 5425–5429.
- 116 G. Schmid and B. Corain, *Eur. J. Inorg. Chem.*, 2003, **2003**, 3081–3098.
- 117 A. N. Shipway, E. Katz and I. Willner, *ChemPhysChem*, 2000, **1**, 18–52.
- 118 C. E. Talley, J. B. Jackson, C. Oubre, N. K. Grady, C. W. Hollars, S. M. Lane, T. R. Huser, P. Nordlander and N. J. Halas, *Nano Lett.*, 2005, **5**, 1569–1574.
- 119 J. N. Anker, W. P. Hall, O. Lyandres, N. C. Shah, J. Zhao and R. P. Van Duyne, in *Nanoscience And Technology: A Collection of Reviews from Nature Journals*, World Scientific, 2010, pp. 308–319.
- 120 J. Sun, Y. Lu, L. He, J. Pang, F. Yang and Y. Liu, *TrAC, Trends Anal. Chem.*, 2020, **122**, 115754.
- 121 N. Lopez, T. V. W. Janssens, B. S. Clausen, Y. Xu, M. Mavrikakis, T. Bligaard and J. K. Nørskov, *J. Catal.*, 2004, **223**, 232–235.
- 122 A. Bej, K. Ghosh, A. Sarkar and D. W. Knight, *RSC Adv.*, 2016, **6**, 11446–11453.
- 123 I. Favier, D. Pla and M. Gómez, *Chem. Rev.*, 2020, **120**, 1146–1183.
- 124 S. Linic, P. Christopher and D. B. Ingram, *Nat. Mater.*, 2011, **10**, 911.
- 125 K. L. McGilvray, M. R. Decan, D. Wang and J. C. Scaiano, *J. Am. Chem. Soc.*, 2006, **128**, 15980–15981.
- 126 S. Kruss, T. Wolfram, R. Martin, S. Neubauer, H. Kessler and J. P. Spatz, *Adv. Mater.*, 2010, **22**, 5499–5506.
- 127 J. F. Hainfeld, D. N. Slatkin, T. M. Focella and H. M. Smilowitz, *Br. J. Radiol.*, 2006, **79**, 248–253.
- 128 S. A. Bansal, V. Kumar, J. Karimi, A. P. Singh and S. Kumar, *Nanoscale Adv.*, 2020, **2**, 3764–3787.
- 129 P. Ghosh, G. Han, M. De, C. K. Kim and V. M. Rotello, *Adv. Drug Delivery Rev.*, 2008, **60**, 1307–1315.
- 130 M. Sharifi, F. Attar, A. A. Saboury, K. Akhtari, N. Hooshmand, A. Hasan, M. A. El-Sayed and M. Falahati, *J. Controlled Release*, 2019, **311–312**, 170–189.
- 131 R. R. Arvizo, S. Bhattacharyya, R. A. Kudgus, K. Giri, R. Bhattacharya and P. Mukherjee, *Chem. Soc. Rev.*, 2012, **41**, 2943–2970.
- 132 B. Kłębowski, J. Depciuch, M. Parlińska-Wojtan and J. Baran, *Int. J. Mol. Sci.*, 2018, **19**, 4031.
- 133 Y. Vlamidis and V. Voliani, *Front. Bioeng. Biotechnol.*, 2018, **6**, 143.
- 134 Y. Xue, X. Li, H. Li and W. Zhang, *Nat. Commun.*, 2014, **5**, 4348.
- 135 H. Grönbeck, A. Curioni and W. Andreoni, *J. Am. Chem. Soc.*, 2000, **122**, 3839–3842.
- 136 A.-S. Duwez, P. Guillet, C. Colard, J.-F. Gohy and C.-A. Fustin, *Macromolecules*, 2006, **39**, 2729–2731.
- 137 I. Blakey, T. L. Schiller, Z. Merican and P. M. Fredericks, *Langmuir*, 2010, **26**, 692–701.
- 138 L. Qiu, J.-W. Li, C.-Y. Hong and C.-Y. Pan, *ACS Appl. Mater. Interfaces*, 2017, **9**, 40887–40897.
- 139 B. Ebeling and P. Vana, *Macromolecules*, 2013, **46**, 4862–4871.
- 140 J. Wagner, W. Peng and P. Vana, *Polymers*, 2018, **10**, 407.
- 141 M. I. Majeed, J. Guo, W. Yan and B. Tan, *Polymers*, 2016, **8**, 392.
- 142 A. Zengin, U. Tamer and T. Caykara, *Anal. Chim. Acta*, 2014, **817**, 33–41.
- 143 X. He, X. Wu, X. Cai, S. Lin, M. Xie, X. Zhu and D. Yan, *Langmuir*, 2012, **28**, 11929–11938.
- 144 H. T. T. Duong, Y. Chen, S. A. Tawfik, S. Wen, M. Parviz, O. Shimoni and D. Jin, *RSC Adv.*, 2018, **8**, 4842–4849.
- 145 W. Zhang, B. Peng, F. Tian, W. Qin and X. Qian, *Anal. Chem.*, 2014, **86**, 482–489.
- 146 D. M. Montana, M. Nasilowski, W. R. Hess, M. Saif, J. A. Carr, L. Nienhaus and M. G. Bawendi, *ACS Appl. Mater. Interfaces*, 2020, **12**, 35845–35855.
- 147 G. Marcelo, F. Burns, T. Ribeiro, J. M. G. Martinho, M. P. Tarazona, E. Saiz, M. G. Moffitt and J. P. S. Farinha, *Langmuir*, 2017, **33**, 8201–8212.
- 148 M. S. Amini-Fazl, M. Barzegarzadeh and R. Mohammadi, *J. Inorg. Organomet. Polym. Mater.*, 2021, **31**, 2959–2970.
- 149 H. Roghani-Mamaqani and K. Khezri, *Appl. Surf. Sci.*, 2016, **360**, 373–382.
- 150 J. Wang, W. Zhang and J. Wei, *J. Mater. Chem. A*, 2019, **7**, 2055–2065.
- 151 P. Ding, J. Zhang, N. Song, S. Tang, Y. Liu and L. Shi, *Composites, Part A*, 2015, **69**, 186–194.
- 152 H. Hosseinzadeh, S. Pashaei, S. Hosseinzadeh, Z. Khodaparast, S. Ramin and Y. Saadat, *Sci. Total Environ.*, 2018, **640–641**, 303–314.
- 153 B. Zhang, Y. Chen, L. Xu, L. Zeng, Y. He, E.-T. Kang and J. Zhang, *J. Polym. Sci., Part A: Polym. Chem.*, 2011, **49**, 2043–2050.
- 154 L. Wu, U. Glebe and A. Böker, *Adv. Mater. Interfaces*, 2017, **4**, 1700092.
- 155 M. S. Strozyk, M. Chanana, I. Pastoriza-Santos, J. Pérez-Juste and L. M. Liz-Marzán, *Adv. Funct. Mater.*, 2012, **22**, 1436–1444.
- 156 F. Han, S. R. C. Vivekchand, A. H. Soeriyadi, Y. Zheng and J. J. Gooding, *Nanoscale*, 2018, **10**, 4284–4290.
- 157 C. K. Adokoh, S. Quan, M. Hitt, J. Darkwa, P. Kumar and R. Narain, *Biomacromolecules*, 2014, **15**, 3802–3810.
- 158 K. Matsuura, K. Ohno, S. Kagaya and H. Kitano, *Macromol. Chem. Phys.*, 2007, **208**, 862–873.
- 159 H. Yun, Y. J. Lee, M. Xu, D. C. Lee, G. E. Stein and B. J. Kim, *ACS Nano*, 2020, **14**, 9644–9651.
- 160 M. S. Yavuz, Y. Cheng, J. Chen, C. M. Cobley, Q. Zhang, M. Rycenga, J. Xie, C. Kim, K. H. Song, A. G. Schwartz, L. V. Wang and Y. Xia, *Nat. Mater.*, 2009, **8**, 935–939.
- 161 X. Chen, J. Lawrence, S. Parelkar and T. Emrick, *Macromolecules*, 2013, **46**, 119–127.
- 162 K. Ohno, K.-m. Koh, Y. Tsujii and T. Fukuda, *Macromolecules*, 2002, **35**, 8989–8993.
- 163 E. G. Westbrook and P. Zhang, *Dalton Trans.*, 2018, **47**, 8638–8645.





- 164 S. Nasri, G. R. Bardajee and M. Bayat, *Colloids Surf., B*, 2018, **171**, 544–552.
- 165 J.-Y. Li, L. Qiu, X.-F. Xu, C.-Y. Pan, C.-Y. Hong and W.-J. Zhang, *J. Mater. Chem. B*, 2018, **6**, 1678–1687.
- 166 D. Huebner, C. Rossner and P. Vana, *Polymer*, 2016, **107**, 503–508.
- 167 D. Rohleder and P. Vana, *Biomacromolecules*, 2021, **22**, 1614–1624.
- 168 J. Tian, B. Huang and W. Zhang, *Langmuir*, 2019, **35**, 266–275.
- 169 C. Rossner, E. B. Zhulina and E. Kumacheva, *Angew. Chem., Int. Ed.*, 2019, **58**, 9269–9274.
- 170 K. Hendrich, W. Peng and P. Vana, *Polymers*, 2020, **12**, 1214.
- 171 C. Rossner, O. Glatter and P. Vana, *Macromolecules*, 2017, **50**, 7344–7350.
- 172 C. Rossner, B. Ebeling and P. Vana, *ACS Macro Lett.*, 2013, **2**, 1073–1076.
- 173 C. Rossner, O. Glatter, O. Saldanha, S. Köster and P. Vana, *Langmuir*, 2015, **31**, 10573–10582.
- 174 F. Han, T. Armstrong, A. Andres-Arroyo, D. Bennett, A. Soeriyadi, A. A. Chamazketi, P. Bakthavathsalam, R. D. Tilley, J. J. Gooding and P. J. Reece, *Nanoscale*, 2020, **12**, 1680–1687.
- 175 W. Peng, C. Rossner, V. Roddatis and P. Vana, *ACS Macro Lett.*, 2016, **5**, 1227–1231.
- 176 D. Miyamoto, M. Oishi, K. Kojima, K. Yoshimoto and Y. Nagasaki, *Langmuir*, 2008, **24**, 5010–5017.
- 177 Y. Nagasaki, *Chem. Lett.*, 2008, **37**, 564–569.
- 178 J. Huang, D. Li, H. Liang and J. Lu, *Macromol. Rapid Commun.*, 2017, **38**, 1700202.
- 179 R. Poupart, A. Benlahoues, B. Le Droumaguet and D. Grande, *ACS Appl. Mater. Interfaces*, 2017, **9**, 31279–31290.
- 180 T. Klein, J. Parkin, P. A. J. M. de Jongh, L. Esser, T. Sepehrizadeh, G. Zheng, M. De Veer, K. Alt, C. E. Hagemeyer, D. M. Haddleton, T. P. Davis, M. Thelakkat and K. Kempe, *Macromol. Rapid Commun.*, 2019, **40**, e1800911.
- 181 C. Boyer, P. Priyanto, T. P. Davis, D. Pissuwan, V. Bulmus, M. Kavallaris, W. Y. Teoh, R. Amal, M. Carroll, R. Woodward and T. St Pierre, *J. Mater. Chem.*, 2010, **20**, 255–265.
- 182 G. Kahraman, D.-Y. Wang, J. von Irmer, M. Gallei, E. Hey-Hawkins and T. Eren, *Polymers*, 2019, **11**, 613.
- 183 Z. Xie, X. Deng, B. Liu, S. Huang, P. Ma, Z. Hou, Z. Cheng, J. Lin and S. Luan, *ACS Appl. Mater. Interfaces*, 2017, **9**, 30414–30425.
- 184 K. W. Fan and A. M. Granville, *Polymers*, 2016, **8**, 81.
- 185 H. Kakwere, M. E. Materia, A. Curcio, M. Prato, A. Sathya, S. Nitti and T. Pellegrino, *Nanoscale*, 2018, **10**, 3930–3944.
- 186 J. Yu, R. J. Nap, I. Szleifer and J. Y. Wong, *Langmuir*, 2019, **35**, 15864–15871.
- 187 H. Kakwere, M. P. Leal, M. E. Materia, A. Curcio, P. Guardia, D. Niculaes, R. Marotta, A. Falqui and T. Pellegrino, *ACS Appl. Mater. Interfaces*, 2015, **7**, 10132–10145.
- 188 W. Liu, A. B. Greytak, J. Lee, C. R. Wong, J. Park, L. F. Marshall, W. Jiang, P. N. Curtin, A. Y. Ting, D. G. Nocera, D. Fukumura, R. K. Jain and M. G. Bawendi, *J. Am. Chem. Soc.*, 2010, **132**, 472–483.
- 189 J. H. Dunlap, A. F. Loszko, R. A. Flake, Y. Huang, B. C. Benicewicz and A. B. Greytak, *J. Phys. Chem. C*, 2018, **122**, 26756–26763.
- 190 J. Liu, W. Yang, L. Tao, D. Li, C. Boyer and T. P. Davis, *J. Polym. Sci., Part A: Polym. Chem.*, 2010, **48**, 425–433.
- 191 Y. Xu, H. Bai, G. Lu, C. Li and G. Shi, *J. Am. Chem. Soc.*, 2008, **130**, 5856–5857.
- 192 G. Cheng, D. Xu, Z. Lu and K. Liu, *ACS Nano*, 2019, **13**, 1479–1489.
- 193 E. Pensa, E. Cortés, G. Corthey, P. Carro, C. Vericat, M. H. Fonticelli, G. Benítez, A. A. Rubert and R. C. Salvarezza, *Acc. Chem. Res.*, 2012, **45**, 1183–1192.
- 194 R. G. Nuzzo, L. H. Dubois and D. L. Allara, *J. Am. Chem. Soc.*, 1990, **112**, 558–569.
- 195 L. Kankate, A. Turchanin and A. Götzhäuser, *Langmuir*, 2009, **25**, 10435–10438.
- 196 C. Yi, H. Liu, S. Zhang, Y. Yang, Y. Zhang, Z. Lu, E. Kumacheva and Z. Nie, *Science*, 2020, **369**, 1369–1374.
- 197 W. Yu, L.-L. Lou, S. Li, T. Ma, L. Ouyang, L. Feng and S. Liu, *RSC Adv.*, 2017, **7**, 751–757.
- 198 L. E. Wilkins, M. Hasan, A. E. R. Fayter, C. Biggs, M. Walker and M. I. Gibson, *Polym. Chem.*, 2019, **10**, 2986–2990.
- 199 Y. Kwon, Y. Choi, J. Jang, S. Yoon and J. Choi, *Pharmaceutics*, 2020, **12**, 204.
- 200 S. Slavin, A. H. Soeriyadi, L. Voorhaar, M. R. Whittaker, C. R. Becer, C. Boyer, T. P. Davis and D. M. Haddleton, *Soft Matter*, 2012, **8**, 118–128.
- 201 Y.-X. Wang, Y. Li, S.-H. Qiao, J. Kang, Z.-L. Shen, N.-N. Zhang, Z. An, X. Wang and K. Liu, *J. Phys. Chem. Lett.*, 2021, **12**, 4713–4721.
- 202 H.-J. Li, P.-Y. Li, L.-Y. Li, A. Haleem and W.-D. He, *Molecules*, 2018, **23**, 921.
- 203 P. Dey, K. J. Thurecht, P. M. Fredericks and I. Blakey, *Macromolecules*, 2020, **53**, 7469–7478.
- 204 B. Huang, J. Tian, D. Jiang, Y. Gao and W. Zhang, *Biomacromolecules*, 2019, **20**, 3873–3883.
- 205 A. Ruiu, B. Bauer-Siebenlist, M. Senila, T. Jänisch, D. Foix, K. Seaudeau-Pirouley and P. Lacroix-Desmazes, *J. CO<sub>2</sub> Util.*, 2020, **41**, 101232.
- 206 S. Zhang, K. L. Chandra and C. B. Gorman, *J. Am. Chem. Soc.*, 2007, **129**, 4876–4877.
- 207 P. Dey, S. Zhu, K. J. Thurecht, P. M. Fredericks and I. Blakey, *J. Mater. Chem. B*, 2014, **2**, 2827–2837.
- 208 S. Moulay, *Polym. Rev.*, 2014, **54**, 436–513.
- 209 M. Monteil, H. Moustauoui, G. Picardi, F. Aouidat, N. Djaker, M. L. de La Chapelle, M. Lecouvey and J. Spadavecchia, *J. Colloid Interface Sci.*, 2018, **513**, 205–213.
- 210 S. W. Han, S. J. Lee and K. Kim, *Langmuir*, 2001, **17**, 6981–6987.



- 211 W. Gan, B. Xu and H.-L. Dai, *Angew. Chem., Int. Ed.*, 2011, **50**, 6622–6625.
- 212 A. B. Lowe, B. S. Sumerlin, M. S. Donovan and C. L. McCormick, *J. Am. Chem. Soc.*, 2002, **124**, 11562–11563.
- 213 G. Corthey, A. A. Rubert, A. L. Picone, G. Casillas, L. J. Giovanetti, J. M. Ramallo-López, E. Zelaya, G. A. Benitez, F. G. Requejo, M. José-Yacamán, R. C. Salvarezza and M. H. Fonticelli, *J. Phys. Chem. C*, 2012, **116**, 9830–9837.
- 214 P. Carro, G. Corthey, A. A. Rubert, G. A. Benitez, M. H. Fonticelli and R. C. Salvarezza, *Langmuir*, 2010, **26**, 14655–14662.
- 215 A. Isakova, P. D. Topham and A. J. Sutherland, *Macromolecules*, 2014, **47**, 2561–2568.
- 216 B. Fan, J. Wan, Y. Liu, W. W. Tian and S. H. Thang, *Polym. Chem.*, 2021, **12**, 3015–3025.
- 217 L. Xiao, J. Li, D. F. Brougham, E. K. Fox, N. Feliu, A. Bushmelev, A. Schmidt, N. Mertens, F. Kiessling, M. Valldor, B. Fadeel and S. Mathur, *ACS Nano*, 2011, **5**, 6315–6324.
- 218 J. Park, N. R. Kadasala, S. A. Abouelmagd, M. A. Castanares, D. S. Collins, A. Wei and Y. Yeo, *Biomaterials*, 2016, **101**, 285–295.
- 219 K. Ulbrich, K. Holá, V. Šubr, A. Bakandritsos, J. Tuček and R. Zbořil, *Chem. Rev.*, 2016, **116**, 5338–5431.
- 220 I. Andreu, E. Natividad, L. Solozábal and O. Roubeau, *ACS Nano*, 2015, **9**, 1408–1419.
- 221 R. Bazak, M. Houri, S. El Achy, S. Kamel and T. Refaat, *J. Cancer Res. Clin. Oncol.*, 2015, **141**, 769–784.
- 222 M. Lévy, C. Wilhelm, J.-M. Siaugue, O. Horner, J.-C. Bacri and F. Gazeau, *J. Phys.: Condens. Matter*, 2008, **20**, 204133.
- 223 K. Turcheniuk, A. V. Tarasevych, V. P. Kukhar, R. Boukherroub and S. Szunerits, *Nanoscale*, 2013, **5**, 10729–10752.
- 224 C. Xu, K. Xu, H. Gu, R. Zheng, H. Liu, X. Zhang, Z. Guo and B. Xu, *J. Am. Chem. Soc.*, 2004, **126**, 9938–9939.
- 225 E. Amstad, A. U. Gehring, H. Fischer, V. V. Nagaiyanallur, G. Hähner, M. Textor and E. Reimhult, *J. Phys. Chem. C*, 2011, **115**, 683–691.
- 226 E. Amstad, T. Gillich, I. Bilecka, M. Textor and E. Reimhult, *Nano Lett.*, 2009, **9**, 4042–4048.
- 227 J. Xie, C. Xu, Z. Xu, Y. Hou, K. L. Young, S. X. Wang, N. Pourmand and S. Sun, *Chem. Mater.*, 2006, **18**, 5401–5403.
- 228 D. Burnand, C. A. Monnier, A. Redjem, M. Schaefer, B. Rothen-Rutishauser, A. Kilbinger and A. Petri-Fink, *J. Magn. Magn. Mater.*, 2015, **380**, 157–162.
- 229 K. Ohno, C. Mori, T. Akashi, S. Yoshida, Y. Tago, Y. Tsujii and Y. Tabata, *Biomacromolecules*, 2013, **14**, 3453–3462.
- 230 Y. Huang, K. Ohno and R. Ishige, *Langmuir*, 2014, **31**, 1172–1179.
- 231 L. Sandiford, A. Phinikaridou, A. Protti, L. K. Meszaros, X. Cui, Y. Yan, G. Frodsham, P. A. Williamson, N. Gaddum, R. M. Botnar, P. J. Blower, M. A. Green and R. T. M. de Rosales, *ACS Nano*, 2013, **7**, 500–512.
- 232 A. J. McGrath, C. Dolan, S. Cheong, D. A. J. Herman, B. Naysmith, F. Zong, P. Galvosas, K. J. Farrand, I. F. Hermans, M. Brimble, D. E. Williams, J. Jin and R. D. Tilley, *J. Magn. Magn. Mater.*, 2017, **439**, 251–258.
- 233 R. Bilan, F. Fleury, I. Nabiev and A. Sukhanova, *Bioconjugate Chem.*, 2015, **26**, 609–624.
- 234 O. T. Bruns, T. S. Bischof, D. K. Harris, D. Franke, Y. Shi, L. Riedemann, A. Bartelt, F. B. Jaworski, J. A. Carr, C. J. Rowlands, M. W. B. Wilson, O. Chen, H. Wei, G. W. Hwang, D. M. Montana, I. Coropceanu, O. B. Achorn, J. Kloepper, J. Heeren, P. T. C. So, D. Fukumura, K. F. Jensen, R. K. Jain and M. G. Bawendi, *Nat. Biomed. Eng.*, 2017, **1**, 1–11.
- 235 T. M. Inerbaev, A. E. Masunov, S. I. Khondaker, A. Dobrinescu, A.-V. Plamadă and Y. Kawazoe, *J. Chem. Phys.*, 2009, **131**, 044106.
- 236 H. R. Chandan, J. D. Schiffman and R. G. Balakrishna, *Sens. Actuators, B*, 2018, **258**, 1191–1214.
- 237 P. N. Goswami, D. Mandal and A. K. Rath, *Nanoscale*, 2018, **10**, 1072–1080.
- 238 T. D. Nguyen, H. Vu-Quang, T. S. Vo, D. C. Nguyen, D.-V. N. Vo, D. H. Nguyen, K. T. Lim, D. L. Tran and L. G. Bach, *Polymers*, 2019, **11**, 77.
- 239 S. Jeong, M. Achermann, J. Nanda, S. Ivanov, V. I. Klimov and J. A. Hollingsworth, *J. Am. Chem. Soc.*, 2005, **127**, 10126–10127.
- 240 M. Green, *J. Mater. Chem.*, 2010, **20**, 5797–5809.
- 241 Y. Zheng, Y. Tang, J. Yu, L. Xie, H. Dong, R. Deng, F. Jia, B. Liu, L. Gao and J. Duan, *Materials*, 2020, **13**, 440.
- 242 Y. Liu, Y. Inoue and K. Ishihara, *Colloids Surf., B*, 2015, **135**, 490–496.
- 243 L. Zhang, D. Jin and M. H. Stenzel, *Biomacromolecules*, 2021, **22**, 3168–3201.
- 244 D. Yang, P. Ma, Z. Hou, Z. Cheng, C. Li and J. Lin, *Chem. Soc. Rev.*, 2015, **44**, 1416–1448.
- 245 V. Muhr, S. Wilhelm, T. Hirsch and O. S. Wolfbeis, *Acc. Chem. Res.*, 2014, **47**, 3481–3493.
- 246 A. Bagheri, H. Arandiyani, C. Boyer and M. Lim, *Adv. Sci.*, 2016, **3**, 1500437.
- 247 A. Sedlmeier and H. H. Gorris, *Chem. Soc. Rev.*, 2015, **44**, 1526–1560.
- 248 A. Bagheri, H. Arandiyani, N. N. M. Adnan, C. Boyer and M. Lim, *Macromolecules*, 2017, **50**, 7137–7147.
- 249 A. Kavand, C. Blanck, F. Przybilla, Y. Mély, N. Anton, T. Vandamme, C. A. Serra and D. Chan-Seng, *Polym. Chem.*, 2020, **11**, 4313–4325.
- 250 A. Bagheri, Z. Sadrearhami, N. N. M. Adnan, C. Boyer and M. Lim, *Polymer*, 2018, **151**, 6–14.
- 251 J. Chen, Q. Chen and Q. Ma, *J. Colloid Interface Sci.*, 2012, **370**, 32–38.
- 252 W. Zhao, Z. Hou, Z. Yao, X. Zhuang, F. Zhang and X. Feng, *Polym. Chem.*, 2015, **6**, 7171–7178.
- 253 Y.-S. Ye, Y.-N. Chen, J.-S. Wang, J. Rick, Y.-J. Huang, F.-C. Chang and B.-J. Hwang, *Chem. Mater.*, 2012, **24**, 2987–2997.



- 254 A. Guerrero-Martínez, J. Pérez-Juste and L. M. Liz-Marzán, *Adv. Mater.*, 2010, **22**, 1182–1195.
- 255 C. Graf, D. L. J. Vossen, A. Imhof and A. van Blaaderen, *Langmuir*, 2003, **19**, 6693–6700.
- 256 W. Stöber, A. Fink and E. Bohn, *J. Colloid Interface Sci.*, 1968, **26**, 62–69.
- 257 Y. Han, Z. Lu, Z. Teng, J. Liang, Z. Guo, D. Wang, M.-Y. Han and W. Yang, *Langmuir*, 2017, **33**, 5879–5890.
- 258 H. L. Ding, Y. X. Zhang, S. Wang, J. M. Xu, S. C. Xu and G. H. Li, *Chem. Mater.*, 2012, **24**, 4572–4580.
- 259 A. M. El-Toni, S. Yin and T. Sato, *J. Colloid Interface Sci.*, 2006, **300**, 123–130.
- 260 K. Nozawa, H. Gailhanou, L. Raison, P. Panizza, H. Ushiki, E. Sellier, J. P. Delville and M. H. Delville, *Langmuir*, 2005, **21**, 1516–1523.
- 261 Y. Kobayashi, H. Inose, T. Nakagawa, K. Gonda, M. Takeda, N. Ohuchi and A. Kasuya, *J. Colloid Interface Sci.*, 2011, **358**, 329–333.
- 262 Y. Han, J. Jiang, S. S. Lee and J. Y. Ying, *Langmuir*, 2008, **24**, 5842–5848.
- 263 M. A. López-Quintela, C. Tojo, M. C. Blanco, L. G. Rio and J. R. Leis, *Curr. Opin. Colloid Interface Sci.*, 2004, **9**, 264–278.
- 264 Y. Cai, W. Peng, S. Demeshko, J. Tian and P. Vana, *Macromol. Rapid Commun.*, 2018, **39**, 1800226.
- 265 A. Pourjavadi, M. Kohestanian and C. Streb, *Mater. Sci. Eng., C*, 2020, **108**, 110418.
- 266 D. Huebner, V. Koch, B. Ebeling, J. Mechau, J. Steinhoff and P. Vana, *J. Polym. Sci., Part A: Polym. Chem.*, 2015, **53**, 103–113.
- 267 S. Ghasemi and S. Karim, *Colloid Polym. Sci.*, 2018, **296**, 1323–1332.
- 268 B. Sahoo, K. S. P. Devi, R. Banerjee, T. K. Maiti, P. Pramanik and D. Dhara, *ACS Appl. Mater. Interfaces*, 2013, **5**, 3884–3893.
- 269 F. Chen, J. Wang, H. Chen, R. Lu and X. Xie, *Appl. Surf. Sci.*, 2018, **435**, 247–255.
- 270 A. Liberman, N. Mendez, W. C. Trogler and A. C. Kummel, *Surf. Sci. Rep.*, 2014, **69**, 132–158.
- 271 Y. Zheng, Y. Huang, Z. M. Abbas and B. C. Benicewicz, *Polym. Chem.*, 2016, **7**, 5347–5350.
- 272 K. C. Bentz and D. A. Savin, *Polym. Chem.*, 2018, **9**, 2059–2081.
- 273 X. Huang, D. Appelhans, P. Formanek, F. Simon and B. Voit, *Macromolecules*, 2011, **44**, 8351–8360.
- 274 M. Dhara, S. Rudra, N. Mukherjee and T. Jana, *Polym. Chem.*, 2021, **12**, 3976–3991.
- 275 G. Moad, *Polym. Int.*, 2015, **64**, 15–24.
- 276 H. Ma, A. Li, Y. Xu, W. Zhang and J. Liu, *Eur. Polym. J.*, 2015, **72**, 212–221.
- 277 W. Li, X. Cai, C. Kim, G. Sun, Y. Zhang, R. Deng, M. Yang, J. Chen, S. Achilefu, L. V. Wang and Y. Xia, *Nanoscale*, 2011, **3**, 1724–1730.
- 278 I. Blakey, Z. Merican and K. J. Thurecht, *Langmuir*, 2013, **29**, 8266–8274.
- 279 T. Honold, D. Skrybeck, K. G. Wagner and M. Karg, *Langmuir*, 2017, **33**, 253–261.
- 280 X. Liao, L. Guo, J. Chang, S. Liu, M. Xie and G. Chen, *Macromol. Rapid Commun.*, 2015, **36**, 1492–1497.
- 281 Z.-L. Wang, J.-T. Xu, B.-Y. Du and Z.-Q. Fan, *J. Colloid Interface Sci.*, 2011, **360**, 350–354.
- 282 P. E. Williams, S. T. Jones, Z. Walsh, E. A. Appel, E. K. Abo-Hamed and O. A. Scherman, *ACS Macro Lett.*, 2015, **4**, 255–259.
- 283 J. Liu, C. Detrembleur, M.-C. De Pauw-Gillet, S. Mornet, E. Duguet and C. Jérôme, *Polym. Chem.*, 2014, **5**, 799–813.
- 284 B. Liao, X. Liu, S. Liao, W. Liu, S. Yi, Q. Liu and B. He, *Polym. J.*, 2020, **52**, 1289–1298.
- 285 T. Kim, M. Xu, Y. J. Lee, K. H. Ku, D. J. Shin, D. C. Lee, S. G. Jang, H. Yun and B. J. Kim, *Small*, 2021, **17**, 2101222.
- 286 J. Zhou, K. Mishra, V. Bhagat, A. Joy and M. L. Becker, *Polym. Chem.*, 2015, **6**, 2813–2816.
- 287 K. Paek, H. Yang, J. Lee, J. Park and B. J. Kim, *ACS Nano*, 2014, **8**, 2848–2856.
- 288 C. H. Park, H. Yang, J. Lee, H.-H. Cho, D. Kim, D. C. Lee and B. J. Kim, *Chem. Mater.*, 2015, **27**, 5288–5294.
- 289 T. Ribeiro, E. Coutinho, A. S. Rodrigues, C. Baleizão and J. P. S. Farinha, *Nanoscale*, 2017, **9**, 13485–13494.
- 290 M. M. Khani, Z. M. Abbas and B. C. Benicewicz, *J. Polym. Sci., Part A: Polym. Chem.*, 2017, **55**, 1493–1501.
- 291 T. Nohara, K. Koseki, K. Tabata, R. Shimada, Y. Suzuki, K. Umemoto, M. Takeda, R. Sato, S. Rodbuntum, T. Arita and A. Masuhara, *ACS Sustainable Chem. Eng.*, 2020, **8**, 14674–14678.
- 292 K. Shito, J. Matsui, Y. Takahashi, A. Masuhara and T. Arita, *Chem. Lett.*, 2018, **47**, 9–12.
- 293 A. Pourjavadi, S. Rahemipoor and M. Kohestanian, *Compos. Sci. Technol.*, 2020, **188**, 107951.
- 294 C. Durand-Gasselín, R. Koerin, J. Rieger, N. Lequeux and N. Sanson, *J. Colloid Interface Sci.*, 2014, **434**, 188–194.
- 295 Z. Cao, N. N. M. Adnan, G. Wang, A. Rawal, B. Shi, R. Liu, K. Liang, L. Zhao, J. J. Gooding, C. Boyer and Z. Gu, *J. Colloid Interface Sci.*, 2018, **521**, 242–251.
- 296 M. J. Roberts, M. D. Bentley and J. M. Harris, *Adv. Drug Delivery Rev.*, 2012, **64**, 116–127.
- 297 K. Knop, R. Hoogenboom, D. Fischer and U. S. Schubert, *Angew. Chem., Int. Ed.*, 2010, **49**, 6288–6308.
- 298 S. Lowe, N. M. O'Brien-Simpson and L. A. Connal, *Polym. Chem.*, 2015, **6**, 198–212.
- 299 Â. Serrano, O. Sterner, S. Mieszkina, S. Zürcher, S. Tosatti, M. E. Callow, J. A. Callow and N. D. Spencer, *Adv. Funct. Mater.*, 2013, **23**, 5706–5718.
- 300 Z. Wu, H. Chen, X. Liu, Y. Zhang, D. Li and H. Huang, *Langmuir*, 2009, **25**, 2900–2906.
- 301 Z. G. Estephan, P. S. Schlenoff and J. B. Schlenoff, *Langmuir*, 2011, **27**, 6794–6800.
- 302 Q. Shao and S. Jiang, *Adv. Mater.*, 2015, **27**, 15–26.
- 303 J. Manson, D. Kumar, B. J. Meenan and D. Dixon, *Gold Bull.*, 2011, **44**, 99–105.
- 304 J. Tournebise, A. Boudier, A. Sapin-Minet, P. Maincent, P. Leroy and R. Schneider, *ACS Appl. Mater. Interfaces*, 2012, **4**, 5790–5799.



- 305 E. Giovanelli, E. Muro, G. Sitbon, M. Hanafi, T. Pons, B. Dubertret and N. Lequeux, *Langmuir*, 2012, **28**, 15177–15184.
- 306 D. Roy, W. L. A. Brooks and B. S. Sumerlin, *Chem. Soc. Rev.*, 2013, **42**, 7214.
- 307 K. J. Hogan and A. G. Mikos, *Polymer*, 2020, **211**, 123063.
- 308 A. Bordat, T. Boissenot, J. Nicolas and N. Tsapis, *Adv. Drug Delivery Rev.*, 2019, **138**, 167–192.
- 309 E. Reimhult, M. Schroffenegger and A. Lassenberger, *Langmuir*, 2019, **35**, 7092–7104.
- 310 O. Veiseh, J. W. Gunn and M. Zhang, *Adv. Drug Delivery Rev.*, 2010, **62**, 284–304.
- 311 S. Shen, B. Ding, S. Zhang, X. Qi, K. Wang, J. Tian, Y. Yan, Y. Ge and L. Wu, *Nanomedicine*, 2017, **13**, 1607–1616.
- 312 M. A. Ward and T. K. Georgiou, *Polymers*, 2011, **3**, 1215–1242.
- 313 I. Yildiz and B. S. Yildiz, *J. Nanomater.*, 2016, **16**, 357.
- 314 J. Lü, Y. Yang, J. Gao, H. Duan and C. Lü, *Langmuir*, 2018, **34**, 8205–8214.
- 315 G. Liu, D. Wang, F. Zhou and W. Liu, *Small*, 2015, **11**, 2807–2816.
- 316 Q. Xiao, Y. Li, F. Li, M. Zhang, Z. Zhang and H. Lin, *Nanoscale*, 2014, **6**, 10179–10186.
- 317 Y. Zheng, A. H. Soeriyadi, L. Rosa, S. H. Ng, U. Bach and J. J. Gooding, *Nat. Commun.*, 2015, **6**, 1–8.
- 318 S. Maji, B. Cesur, Z. Zhang, B. G. De Geest and R. Hoogenboom, *Polym. Chem.*, 2016, **7**, 1705–1710.
- 319 R. Cheng, F. Meng, C. Deng, H.-A. Klok and Z. Zhong, *Biomaterials*, 2013, **34**, 3647–3657.
- 320 X. Liu, Y. Yang and M. W. Urban, *Macromol. Rapid Commun.*, 2017, **38**, 1700030.
- 321 R. Pelton, *J. Colloid Interface Sci.*, 2010, **348**, 673–674.
- 322 S. T. Jones, Z. Walsh-Korb, S. J. Barrow, S. L. Henderson, J. del Barrio and O. A. Scherman, *ACS Nano*, 2016, **10**, 3158–3165.
- 323 D. Fava, M. A. Winnik and E. Kumacheva, *Chem. Commun.*, 2009, 2571–2573.
- 324 H. Han, J. Y. Lee and X. Lu, *Chem. Commun.*, 2013, **49**, 6122–6124.
- 325 J. Niskanen and H. Tenhu, *Polym. Chem.*, 2017, **8**, 220–232.
- 326 F. Liu and S. Agarwal, *Macromol. Chem. Phys.*, 2015, **216**, 460–465.
- 327 X. Cai, L. Zhong, Y. Su, S. Lin and X. He, *Polym. Chem.*, 2015, **6**, 3875–3884.
- 328 Z. Dong, J. Mao, D. Wang, M. Yang, W. Wang, S. Bo and X. Ji, *Macromol. Chem. Phys.*, 2014, **215**, 111–120.
- 329 G. K. Such, Y. Yan, A. P. R. Johnston, S. T. Gunawan and F. Caruso, *Adv. Mater.*, 2015, **27**, 2278–2297.
- 330 A.-H. Ranneh, H. Takemoto, S. Sakuma, A. Awaad, T. Nomoto, Y. Mochida, M. Matsui, K. Tomoda, M. Naito and N. Nishiyama, *Angew. Chem., Int. Ed.*, 2018, **57**, 5057–5061.
- 331 H. Tang, W. Zhao, J. Yu, Y. Li and C. Zhao, *Molecules*, 2018, **24**, 4.
- 332 X. Lv, J. Zhang, D. Yang, J. Shao, W. Wang, Q. Zhang and X. Dong, *J. Mater. Chem. B*, 2020, **8**, 10700–10711.
- 333 H. Akiyama, K. Tamada, J. Nagasawa, K. Abe and T. Tamaki, *J. Phys. Chem. B*, 2003, **107**, 130–135.
- 334 R. Klajn, *Chem. Soc. Rev.*, 2014, **43**, 148–184.
- 335 P. K. Kundu, D. Samanta, R. Leizrowice, B. Margulis, H. Zhao, M. Börner, T. Udayabhaskararao, D. Manna and R. Klajn, *Nat. Chem.*, 2015, **7**, 646–652.
- 336 T. Bian, Z. Chu and R. Klajn, *Adv. Mater.*, 2020, **32**, 1905866.
- 337 L. Zhang, L. Dai, Y. Rong, Z. Liu, D. Tong, Y. Huang and T. Chen, *Langmuir*, 2015, **31**, 1164–1171.
- 338 J. Shi, F. Tian, J. Lyu and M. Yang, *J. Mater. Chem. B*, 2015, **3**, 6989–7005.
- 339 Z. Gueroui and A. Libchaber, *Phys. Rev. Lett.*, 2004, **93**, 166108.
- 340 S. Jiang and Y. Zhang, *Langmuir*, 2010, **26**, 6689–6694.
- 341 C. Chen and N. Hildebrandt, *TrAC, Trends Anal. Chem.*, 2020, **123**, 115748.
- 342 C. S. Yun, A. Javier, T. Jennings, M. Fisher, S. Hira, S. Peterson, B. Hopkins, N. O. Reich and G. F. Strouse, *J. Am. Chem. Soc.*, 2005, **127**, 3115–3119.
- 343 P. F. Gao, Y. F. Li and C. Z. Huang, *TrAC, Trends Anal. Chem.*, 2020, **124**, 115805.
- 344 G. L. Liu, Y.-T. Long, Y. Choi, T. Kang and L. P. Lee, *Nat. Methods*, 2007, **4**, 1015–1017.
- 345 Y. Choi, T. Kang and L. P. Lee, *Nano Lett.*, 2009, **9**, 85–90.
- 346 C. Chen and N. Hildebrandt, *TrAC, Trends Anal. Chem.*, 2020, **123**, 115748.
- 347 E. Morales-Narváez, B. Pérez-López, L. B. Pires and A. Merkoçi, *Carbon*, 2012, **50**, 2987–2993.
- 348 C. Zhu, Z. Zeng, H. Li, F. Li, C. Fan and H. Zhang, *J. Am. Chem. Soc.*, 2013, **135**, 5998–6001.
- 349 F. Prins, A. J. Goodman and W. A. Tisdale, *Nano Lett.*, 2014, **14**, 6087–6091.
- 350 A. Kasry, A. A. Ardakani, G. S. Tulevski, B. Menges, M. Copel and L. Vyklicky, *J. Phys. Chem. C*, 2012, **116**, 2858–2862.
- 351 J. Saha, A. D. Roy, D. Dey, D. Bhattacharjee and S. A. Hussain, *Mater. Today: Proc.*, 2018, **5**, 2306–2313.
- 352 K. Paek, S. Chung, C.-H. Cho and B. J. Kim, *Chem. Commun.*, 2011, **47**, 10272–10274.
- 353 M. Dadsetan, K. E. Taylor, C. Yong, Ž. Bajzer, L. Lu and M. J. Yaszemski, *Acta Biomater.*, 2013, **9**, 5438–5446.
- 354 H. Yang, K. Paek and B. J. Kim, *Nanoscale*, 2013, **5**, 5720–5724.
- 355 M. Hwang and B. Yeom, *Chem. Mater.*, 2021, **33**, 807–817.
- 356 W. Ma, H. Kuang, L. Xu, L. Ding, C. Xu, L. Wang and N. A. Kotov, *Nat. Commun.*, 2013, **4**, 1–8.
- 357 X. Wu, L. Xu, L. Liu, W. Ma, H. Yin, H. Kuang, L. Wang, C. Xu and N. A. Kotov, *J. Am. Chem. Soc.*, 2013, **135**, 18629–18636.
- 358 Y. Zhou, R. L. Marson, G. Van Anders, J. Zhu, G. Ma, P. Ercius, K. Sun, B. Yeom, S. C. Glotzer and N. A. Kotov, *ACS Nano*, 2016, **10**, 3248–3256.
- 359 A. D. Merg, J. C. Boatatz, A. Mandal, G. Zhao, S. Mokashi-Punekar, C. Liu, X. Wang, P. Zhang, P. C. van der Wel and N. L. Rosi, *J. Am. Chem. Soc.*, 2016, **138**, 13655–13663.





- 360 W. Yan, L. Xu, C. Xu, W. Ma, H. Kuang, L. Wang and N. A. Kotov, *J. Am. Chem. Soc.*, 2012, **134**, 15114–15121.
- 361 Q. Jiang, Q. Liu, Y. Shi, Z.-G. Wang, P. Zhan, J. Liu, C. Liu, H. Wang, X. Shi, L. Zhang, J. Sun, B. Ding and M. Liu, *Nano Lett.*, 2017, **17**, 7125–7130.
- 362 X. Lan, X. Lu, C. Shen, Y. Ke, W. Ni and Q. Wang, *J. Am. Chem. Soc.*, 2015, **137**, 457–462.
- 363 C. Li, Q. Li, Y. V. Kaneti, D. Hou, Y. Yamauchi and Y. Mai, *Chem. Soc. Rev.*, 2020, **49**, 4681–4736.
- 364 T. N. Hoheisel, K. Hur and U. B. Wiesner, *Prog. Polym. Sci.*, 2015, **40**, 3–32.
- 365 A. H. Gröschel, A. Walther, T. I. Löbbling, F. H. Schacher, H. Schmalz and A. H. E. Müller, *Nature*, 2013, **503**, 247–251.
- 366 R. M. Choueiri, E. Galati, H. Thérien-Aubin, A. Klinkova, E. M. Larin, A. Querejeta-Fernández, L. Han, H. L. Xin, O. Gang, E. B. Zhulina, M. Rubinstein and E. Kumacheva, *Nature*, 2016, **538**, 79–83.
- 367 J.-M. Rabanel, V. Adibnia, S. F. Tehrani, S. Sanche, P. Hildgen, X. Banquy and C. Ramassamy, *Nanoscale*, 2019, **11**, 383–406.
- 368 N. D. Knöfel, H. Rothfuss, C. Barner-Kowollik and P. W. Roesky, *Polym. Chem.*, 2019, **10**, 86–93.
- 369 M. Sun, L. Xu, W. Ma, X. Wu, H. Kuang, L. Wang and C. Xu, *Adv. Mater.*, 2016, **28**, 898–904.
- 370 A. Qu, L. Xu, M. Sun, L. Liu, H. Kuang and C. Xu, *Adv. Funct. Mater.*, 2017, **27**, 1703408.
- 371 L. He, M. Brasino, C. Mao, S. Cho, W. Park, A. P. Goodwin and J. N. Cha, *Small*, 2017, **13**, 1700504.
- 372 Y. Zheng, T. Thai, P. Reineck, L. Qiu, Y. Guo and U. Bach, *Adv. Funct. Mater.*, 2013, **23**, 1519–1526.
- 373 D. Sun, Y. Tian, Y. Zhang, Z. Xu, M. Y. Sfeir, M. Cotlet and O. Gang, *ACS Nano*, 2015, **9**, 5657–5665.
- 374 N. Gandra, A. Abbas, L. Tian and S. Singamaneni, *Nano Lett.*, 2012, **12**, 2645–2651.
- 375 J. H. Yoon, J. Lim and S. Yoon, *ACS Nano*, 2012, **6**, 7199–7208.
- 376 Y. Yang, J. Zhu, J. Zhao, G.-J. Weng, J.-J. Li and J.-W. Zhao, *ACS Appl. Mater. Interfaces*, 2019, **11**, 3617–3626.
- 377 R. Schreiber, J. Do, E.-M. Roller, T. Zhang, V. J. Schüller, P. C. Nickels, J. Feldmann and T. Liedl, *Nat. Nanotechnol.*, 2014, **9**, 74–78.
- 378 L. Y. T. Chou, K. Zagorovsky and W. C. W. Chan, *Nat. Nanotechnol.*, 2014, **9**, 148–155.
- 379 C. Rossner, Q. Tang, O. Glatter, M. Müller and P. Vana, *Langmuir*, 2017, **33**, 2017–2026.
- 380 J. Moraes, K. Ohno, T. Maschmeyer and S. Perrier, *Chem. Commun.*, 2013, **49**, 9077–9088.
- 381 H. Zou, S. Wu and J. Shen, *Chem. Rev.*, 2008, **108**, 3893–3957.

