Deciphering the mechanism of O₂ reduction with electronically tunable non-heme iron enzyme model complexes†

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A homologous series of electronically tuned 2,2′,2″-nitriilotris(N-arylacetamide) pre-ligands (H₄L₈) were prepared (R = NO₂, CN, CF₃, F, Cl, Br, Et, Me, H, OMe, NMe₂) and some of their corresponding Fe and Zn species synthesized. The iron complexes react rapidly with O₂, the final products of which are diferric mu-oxo bridged species. The crystal structure of the oxidized product obtained from DMA solutions contain a structural motif found in some diiron proteins. The mechanism of iron mediated O₂ reduction was explored to the extent that allowed us to construct an empirically consistent rate law. A Hammett plot was constructed that enabled insightful information into the rate-determining step and hence allows for a differentiation between two kinetically equivalent O₂ reduction mechanisms.

The systematic electronic and steric tuning of synthetic enzyme models offers a potential solution to this dilemma.⁹⁻¹¹

One of the most powerful techniques used to investigate mechanism that takes advantage of systematic changes is the linear free energy relationship in the form of a Hammett plot.¹² While linear free energy relationships have been used to great success in understanding O₂ activation,¹⁰,¹¹ the specific use of the Hammett plot in inorganic and organometallic reactions is not as common and, to our knowledge, only a few reports have demonstrated the utility of the Hammett plot in O₂ reduction by synthetic non-heme iron complexes.¹⁴,¹⁵ The first step in O₂ activation at non-heme centers, namely the two limiting cases of inner vs. outer sphere reduction of O₂, has not been thoroughly addressed when compared to heme analogues that have been extensively studied.⁷,¹⁶⁻¹⁸ In fact, the discussion about O₂ binding and reduction in non-heme centers is predominantly described as an inner sphere process.⁶⁻¹² While an inner sphere reduction to form Fe(III)-superoxo species is reasonable and probably true in many cases, the alternative outer sphere description is equally plausible.

To this end, we report a systematically varied series of N-arylacetamide ligands that contain remote substituents for electronic tuning of metal-ligand bonding for the purpose of using a Hammett plot to gain insight into the rate-limiting step of O₂ reduction. Herein we report the synthesis and characterization of these new ligands in addition to the biologically relevant Fe and Zn metal complexes. Finally, the iron complexes react with molecular oxygen and the mechanism of this reaction was deciphered with the aid of a Hammett plot. To our knowledge, this study serves as the first kinetic analysis that...
Results and discussion

Synthesis of ligands

The new ligands described in this report are based on the tris-aceamidate ligands first used by Borovik and coworkers. They prepared a variety of aliphatic and aryl acetamidate ligands and used them in coordination chemistry studies with first-row transition metal complexes including O₂ activation and stabilization of unusual electronic and coordination environments. We adopted the synthetic strategy for the known 2,2',2''-nitrotris(N-(3,5-dimethylphenyl)acetamide) compound [H₃L₃imp] to prepare the new ligands, which involves heating a solution of nitritotriacetic acid in pyridine and triphenylphosphate with the appropriate aniline (Scheme 1). Eleven ligands H₃L₄ (R = NO₂, CN, CF₃, F, Cl, Br, Et, Me, H, OMe, NMe₂) were thus obtained in good yield and high purity. A plot of the ¹H-NMR acetamide N resonance vs. the Hammett parameters reveals a linear correlation (Fig. S1†) confirming electronic communication between the substituents and the aryaceticamide nitrogen atom that will serve as the donor to a transition metal ion.

Metal complex synthesis and characterization

The ligands can be deprotonated in dimethylacetamide (DMA) solvent with three equiv. of KH to afford the respective ligand salt. These are then treated with M(OAc)₂ (M = Fe or Zn) followed by two equiv. of [Me₄N][OAc]. The resulting KOAc is easily removed by filtration and subsequent recrystallization of the complexes afford [Me₄N]₂[ML(R)(OAc)] salts. The [Me₄N]₂[ML(NO₂)(OAc)] and [Me₄N]₂[ML(OAc)] (M = Fe, Zn) salts were characterized by XRD. The [Me₄N]₂[ML(NO₂)(OAc)] (M = Zn, Fe) species have distorted six-coordinate geometries (Fig. 1 and S3, Table S1†). When M = Fe, the acetato ligand adopts a bidentate orientation for R = NO₂ (1NO₂) rather than the monodentate mode in the R = H molecules (1H), possibly indicating a more electron deficient metal. The acetate ligation in the respective Zn complexes (R = H, NO₂) is similar to the iron complexes except that the long Zn–O in [ZnL(NO₂)(OAc)] is about 0.4 Å longer. The solution state-structure of the two Zn complexes (R = H and NO₂) was probed by ¹H-NMR spectroscopy. With the exception of a broadened peak for the acetato ligand, which might indicate fluxional ligation or exchange, the peaks are sharp and reveal a C₃ symmetric coordination-geometry on the NMR time scale (Fig. 2 and S2†). Hence, we assume that the solution-state structure of the Fe(II) ions is somewhat similar to the Zn complexes. The salt [Me₄N]₂[Fe(NO₂)(OAc)] was characterized by Mössbauer spectroscopy in the solid state and has parameters consistent with an S = 2 species. This is in agreement with the room temperature solution NMR Evans’ method magnetic moment of μeff = 4.91 μB.

Bulk oxidation of [Fe(NO₂)(OAc)]²⁻ with O₂

The iron complexes react rapidly with molecular oxygen (pure O₂ or in air) forming a red compound (2B). Preparative scale reactions performed using O₂ and 1NO₂ in DMA or MeCN resulted in good isolated yield (≥88%) of [Me₄N]₂[Fe(NO₂)(OAc)] (1NO₂), [μ-O]-[μ-K₂(O,O')-OAc] (2a) or [Me₄N]₂[Fe(NO₂)₂·μ-O] (2b), respectively (Scheme 2). In contrast to previous studies using O₂ on similar platforms, the final products isolated herein are specifically attempts to address the outer vs. inner sphere hypothesis in non-heme iron enzyme model complexes.
dimeric μ-oxido complexes rather than mononuclear complexes with terminal hydroxido ligands. The lack of steric protection is the probable cause for this difference since the bulky aliphatic-acetamide ligand L^{1PV} stabilizes the terminal hydroxido K[FeL^{1PV}(OH)]. Another major difference between 2a/b and monomeric ferric hydroxido complexes with similar ligands is that one of the ligand arms in 2a/b has an altered binding mode, having undergone tautomerization. As such, each iron center in 2a/b contains one anionic oxygen donor from one of the acetimidate moieties and two anionic nitrogen donors binding in the usual fashion from the other two acetamide arms. The assignment of the charges on the donor groups is supported by the number of counter ions in the unit cell and the substantial differences in the C=N bond lengths (Table 1). For example, the [K(L_{dmp}FeNO)]_{7} complex has C=N bond distances of 1.34 Å, which is comparable to 1^{NO2} and 1^{H} and consistent with the acetamidate ligation. Similarly, two of the C=N bonds in each of the crystallographically related halves of the 2a and 2b molecules are 1.34 and 1.35 Å, respectively, further indicating acetamidate ligation. The remaining C=N bond distance in 2a and 2b is shorter, 1.30 Å, and supports the assignment of acetamidate ligation.

A key difference between the 2a and 2b is the coordination number at the iron centers (Fig. 3). 2b attains a five coordinate geometry with loss of local three-fold symmetry in the primary coordination sphere due to the tautomerization and hence an asymmetrical binding mode. In contrast, the iron centers in 2a are six-coordinate due to the additional ligation of the bridging acetate ligand. The synthetic procedure for 2a and 2b differ only in the solvent used (DMA and MeCN, respectively). Thus, the formation of two similar dimeric μ-oxido complexes is likely caused by the greater extent to which DMA can stabilize the trianionic 2a during crystallization.

Interestingly, the structure of 2a resembles carboxylate-oxo-bridged diiron enzymes that are important in a number of biological transformations that use molecular oxygen. It is well established that the bridging ligands strongly influence the magnetic properties of these active sites and influence chemistry. Similarly here, the binding of acetate to the diferric core appears to influence the magnetic properties of the complex. For example, 2b has a magnetic moment of 2.26 μB (DMSO, room temperature) that is similar to other μ-oxido differferic complexes and 2a has a higher magnetic moment of 3.01 μB. However, little can be said about these differences because the solution speciation of 2a appears to be complicated. For instance, the UV-vis spectra of 2a and 2b are essentially identical in DMA and suggest that the binding is minimal in solution. In fact, treatment of a solution of 2b in DMA with 0.10, 1.0, 10 and 30 equivalents of [Me_{4}N][OAc] causes a shift in the UV-vis spectrum to lower energy to occur with no isosbestic point implicating multiple binding modes or complicated equilibria (Fig. S6†).

The formation of the mu-oxo species 2^{NO2} likely forms from the condensation of O_{2} derived {Fe(n)[OH]}_{n} (n = 1 or 2) species. To test this premise, we quantified the water produced in the reaction between 1^{NO2} and O_{2} in bulk oxidations using ^{19}F-NMR spectroscopy and the water sensitive reagent iodosobenzene difluoride (PhIF_{2}). Specifically, the volatiles from a solution of freshly prepared 2b were transferred to a clean, dry flask via trap-to-trap vacuum distillation on a high-vacuum line. The distillate was transferred into a glove box and treated with freshly prepared PhIF_{2} in solvent dried with newly activated alumina and the solution contents were analyzed by ^{19}F-NMR spectroscopy. Any water in solution reacts with PhIF_{2} to form the [FHF] anion, which can be quantified using BF_{4} internal standard. It was found that about 0.5 equivalent H_{2}O formed per molecule of 1^{NO2} used (three runs, 54%, 37%, and 30% yield H_{2}O based on iron).

Hence, it is reasonable to assume that “Fe(n)[OH]” moieties form in the reaction, likely through a C-H bond cleavage reaction. To further test this hypothesis, we included 10 equiv. of dihydroanthracene (DHA) in a bulk oxidation reaction, but we did not observe anthracene as a product. A likely reactive intermediate in the oxidation of 1^{S} is a superoxo species with an accessible active site that is exposed to free solvent; for such an intermediate, DHA may not be able to compete kinetically with solvent molecules in a bimolecular reaction. Recently, it has been shown that enzymatic and synthetic iron-superoxo species...
are competent for such transformations, but other intermediate species (e.g., oxo) are also possible.

Mechanism of iron mediated O2 reduction

The kinetics of the reactions of [FeLH(OAc)]2− (1H), [FeL(OAc)]2− (1Cl), and 1NO2 with O2 in DMA were investigated with UV-vis spectroscopy. The reactions with O2 and 1H in DMA were carried out in Schlenk UV-vis cuvettes that were degassed and equilibrated at 20 °C prior to exposure to 0.75 atm of pure, dry O2. The dissolution of O2 initially causes complication in the kinetic analysis and has been described before as prohibitive to mechanistic studies. However, we have conducted a mass transfer analysis that accounts for this complication and is further enabled by the fact that O2 saturation occurs early enough that we can determine first order rate constants (see ESI†). A representative plot of spectra obtained by treatment of 1H in DMA with O2 is shown in Fig. 4. In the case of 1NO2, the UV-vis spectrum of the final species (designated 2NO2) in low concentration experiments is almost identical to 2b with added acetate in solution (Fig. S6†). Specifically, the λmax of the final product is shifted by 10 nm from 2b and has a lower extinction coefficient. Considering the complicated equilibrium between 2b and [Me4N][OAc], we propose that the final product generated in UV-vis cuvettes is an isomer of 2a that converts into 2a upon crystallization at higher concentrations. To avoid complications from incomplete knowledge about the speciation

Table 1 Crystallographic bond metrics for the Fe complexes with comparative examplesa

<table>
<thead>
<tr>
<th>M8</th>
<th>1H</th>
<th>1NO2</th>
<th>2a</th>
<th>2b</th>
<th>K[LiPrFeIIIOH]b</th>
<th>[K[LiPrFeNO]]7 c</th>
</tr>
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<tbody>
<tr>
<td>Fe-Namine (Å)</td>
<td>2.235(2)</td>
<td>2.231(1)</td>
<td>2.251(2)</td>
<td>2.245(2)</td>
<td>2.194(3)</td>
<td>2.198(2)</td>
</tr>
<tr>
<td>Fe-Nimidate ave (Å)</td>
<td>2.015</td>
<td>2.158</td>
<td>2.114</td>
<td>2.027</td>
<td>2.022</td>
<td>2.026</td>
</tr>
<tr>
<td>Cimidate−Nimidate</td>
<td>—</td>
<td>—</td>
<td>1.304(4)</td>
<td>1.295(4)</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>Cimidate−Nimidate</td>
<td>1.338(3)</td>
<td>1.355(2)</td>
<td>1.339(4)</td>
<td>1.349(4)</td>
<td>1.307(6)</td>
<td>1.339(3)</td>
</tr>
<tr>
<td>1.339(3)</td>
<td>1.324(2)</td>
<td>1.341(3)</td>
<td>1.350(4)</td>
<td>1.314(5)</td>
<td>1.339(3)</td>
<td></td>
</tr>
<tr>
<td>1.331(2)</td>
<td>1.335(2)</td>
<td>—</td>
<td>—</td>
<td>1.321(4)</td>
<td>1.341(3)</td>
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</tr>
<tr>
<td>Fe-Oimidate</td>
<td>—</td>
<td>—</td>
<td>2.071(2)</td>
<td>1.977(2)</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>Fe-X (Å) (X = OAc)</td>
<td>2.046(1)</td>
<td>2.111(1) (X = OAc)</td>
<td>1.800 (X = O)</td>
<td>1.782(1) (X = O)</td>
<td>1.876(3) (X = OH)</td>
<td>1.748(2) (X = NO)</td>
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<tr>
<td>Fe-OFe (Å)</td>
<td>—</td>
<td>128.1</td>
<td>180</td>
<td>—</td>
<td>—</td>
<td>—</td>
</tr>
</tbody>
</table>

a Counterion is Me4N+ unless otherwise noted. b Shorter C–N bond lengths due to aliphatic trisacetamidate ligand L3Fe. c From ref. 25.

Fig. 3 Molecular structures of [Me4N]3[FeIII(LNO2)_2-μ-O-μ-O-μ-O-μ-O(OAc)] (2a, top) and [Me4N]2[FeIII(LNO2)_2-μ-O] (2b, bottom); solvent and counter ion molecules not shown. Ellipsoids drawn at 50% probability and H-atoms removed for clarity. Color scheme: orange = iron; blue = nitrogen; red = oxygen; grey = carbon. Fe−Fe distance 3.2371(5) Å for 2a and 3.5647(7) Å for 2b.

Fig. 4 Representative UV-vis spectra monitoring the oxidation of 1H (0.1 mM) with O2 (0.75 atm) in DMA (20 °C) (blue spectrum = 1H at t = 0; red spectrum = final product at t = 1000). Inset: 400 nm trace with 10 seconds intervals between spectra.
of $2^R$, we performed our kinetic analysis by following the consumption of $1^R$ by method of extent of reaction.

For the three complexes $1^R$, $1^C$, and $1^{NO_2}$ it was determined using log–log plots and flooding methods that the reaction is first order in iron and has a complicated dependence on acetate and $O_2$ (Fig. S8–S11†). Taken together, we propose the following mechanism (Scheme 3): (1) reversible acetate dissociation is followed by (2) a rate limiting $O_2$ binding step (step 2-III) or outer sphere electron transfer (step 2-II and 2-II); (3-4) reduction of $O_2$ is then followed by several fast steps to form an iron(II) compound $2^R$. A steady-state approximation of the proposed mechanism with the mono anionic $[Fe(n)L^R]$ serving as the intermediate gives a single term rate law of the following form (see ESI† for derivation):

$$\frac{d[2^R]}{dt} = \frac{k_1k_2[FeL^R(OAc)]O_2}{k_{-1}(OAc) + k_5[O_2]}$$

This rate law simplifies further to $k_{obs}[FeL^R(OAc)]$ (eqn S1–S5†). Following consumption of $1^R$ as a function of time provides first order plots with a $k_{obs} = 0.017 \pm 0.004$ s$^{-1}$ for $R = H$ (Table S2†) that is effectively independent of $[FeL^R(OAc)]$. To further test the rate law, we kept iron concentration constant and varied the concentration of $O_2$ in the presence of additional acetate (20, 30, and 40 equiv. $[Me_4N][OAc]$) and plotted $1/k_{obs}$ against $1/[O_2]$ (Fig. S10†). The plots with different acetate concentration each furnish a horizontal region with a $y$ intercept of 1/k$_1$ providing a value of $k_1 = 0.024 \pm 0.006$ M$^{-1}$ s$^{-1}$ (R = H). The values of $k_2$, $k_{eq}$, and $k_{-1}$ are 0.39 M$^{-1}$ s$^{-1}$, 0.08, and 0.27 M$^{-1}$ s$^{-1}$, respectively, were obtained through solving a system of equations (eqn S11–S14†). These values are an estimate that is accurate to the order or magnitude presented due to the tolerance set in the MATALAB code we used to solve the system of equations.

Taking advantage of the fact that the rate law can be approximated by $k_1[FeL^R(OAc)]$ in the absence of free acetate ($k_1 = k_{obs}$), the rate constant was measured over the temperature range from −10 to 70 °C. Unfortunately, the Eyring plot (see Table S3†) contains a large degree of scatter because the rate has a negligible dependence on temperature; $k_{obs}$ has a value of 0.023 s$^{-1} \pm 0.004$ from the range of −10 to 70 °C (Table S3†). There also appears to be an inflection point near 10 °C, but the large scatter makes this Eyring plot difficult to interpret and possibly not informative outside the context of other similar studies.

Iron centers that bind $O_2$ often have small enthalpy of activation, reflecting the fact that $O_2$ is a poor ligand.30,39 Our lack of a clear relationship between temperature and rate may also reflect a small entropic contribution. This is only speculative however and there might be other factors such as competing pathways with relatively similar barriers. For instance, Busch observed complicated parabolic dependence of the rate constant with temperature for $O_2$ with myoglobin, hemoglobin, and cyclidene complexes.40 Busch’s interpretation of the temperature dependence relied on competing inner and outer sphere $O_2$ reduction pathways in addition to competitive ligand binding – all of which are possible in the system studied here.

These two possible $O_2$ reduction pathways, one involving rate limiting inner sphere $O_2$ binding and reduction (step 2-III in Scheme 3) and the other rate limiting outer sphere electron transfer followed by rapid superoxide coordination (step 2-III and 2-II, respectively, in Scheme 3), provide the same rate law and are difficult to distinguish. Herein lies the advantage of the Hammett plot to decipher reaction mechanisms. The first order rate constants for five of the $1^R$ complexes were plotted against the Hammett parameter (σ) and from this plot a negative slope was obtained (Fig. 5 and S7†). An even better fit was obtained when we used the Swain–Lupton correlation that takes into account both inductive and resonance effects.41 The negative slope in these plots indicates positive charge build up in the transition state and is expected for an outer sphere electron transfer event. An alternative interpretation is positive charge build up from loss of acetate ligand. However, the rate-determining step is not acetate loss and so we surmise that the data are most consistent with a rate-determining outer sphere electron transfer event.

$$y = -1.562x + 0.019$$

$$R^2 = 0.997$$

Fig. 5 Swain–Lupton plot (variation of a Hammett plot) for reaction between $1^R$ and $O_2$ in DMA at 20 °C. The rate constants obtained in triplicate for each substituent are the following: OMe = 0.042 ± 0.01; Et = 0.023 ± 0.005; H = 0.015 ± 0.003; F = 0.018 ± 0.004; NO$_2$ = 0.0038 ± 0.0006. See Fig. S7† for additional Hammett and Swain–Lupton plots.

Scheme 3 Proposed $O_2$ reduction mechanism with Fe$^L$$^R$. 

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To put our work in context, Sun and coworkers have studied the \( \text{O}_2 \) reduction dioxygenase model reaction with a six-coordinate non-heme iron complex.\(^{12}\) Assuming that \( \text{O}_2 \) reduction is rate limiting in their reaction, the negative slope in their Hammett plot also indicates that an outer sphere mechanism is operative. This is expected for a six-coordinate iron species. However, Que and coworkers reported a Hammett plot with a positive slope that is consistent with an outer sphere reduction of \( \text{O}_2 \) in the equilibrium constant of \( k \). The dependence of the rate on the mechanism of the formation of \( \text{O}_2 \) reduction step in non-heme iron metallocentres in non-haem iron(II)-dependent enzymes, Models, and Intermediates, Biochem. Soc. Trans., 2008, 36, 1180–1186.


15. Y.-J. Sun, Q.-Q. Huang and J.-J. Zhang, Set of Fe(n)-3-Hydroxyflavonolate Enzyme-Substrate Model Complexes of Atypically Coordinated Mononuclear Non-Heme Fe(n)-Dependent Quercetin 2,4-Dioxygenase, *ACS Omega*, 2017, 2, 5850–5860.


33. Autoxidation (dissociation of superoxide from ferric center) is also possible. In this case, solvent reduces free superoxide into water that further reacts with the ferric centers to produce the same final products.


