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Michael Malkoch *et al.*

Degradable high  $T_g$  sugar-derived polycarbonates from isosorbide and dihydroxyacetone



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## Degradable high $T_g$ sugar-derived polycarbonates from isosorbide and dihydroxyacetone†

Daniel Hult, Sandra García-Gallego, Tobias Ingverud, Oliver C. J. Andrén and Michael Malkoch\*

Polycarbonates from isosorbide and dihydroxyacetone (DHA) have been synthesised using organocatalytic step-growth polymerization of their corresponding diols and bis-carbonylimidazolidone monomers. By the choice of the feed ratio and monomer activation, either isosorbide or ketal protected DHA, random and alternating poly(iso-co-DHA) carbonates have been formed. Thermal properties by DSC and TGA were herein strongly correlated to the monomer composition. Dilution studies using  $^1\text{H-NMR}$  spectroscopy of a model compound DHA-diethyl carbonate in  $\text{CD}_3\text{CN}$  and  $\text{D}_2\text{O}$  highlighted the influence of  $\alpha$ -substituents on the keto/hydrate equilibrium of DHA. Further kinetics studies in the  $\text{pH}^*$  range of 4.7 to 9.6 serve to show the hydrolytic pH-profile of DHA-carbonates. The hydrolytic degradation of deprotected polymer pellets shows an increased degradation with increasing DHA content. Pellets with a random or alternating configuration show different characteristics in terms of mass loss and molecular weight loss profile over time.

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### Introduction

Aliphatic poly(carbonate)s (APCs) are still considered as an unexplored family of polymers. This has mainly been due to their slow degradability and low glass-transition temperatures ( $T_g$ ). In comparison with their aromatic analogues, the inherent properties of APCs are today seen as attractive features for biomedical applications.<sup>1,2</sup> In an effort to expand the uses of APCs, researchers are currently focusing on the sustainable development of poly(carbonate)s through the synthesis of renewable monomers and polymers with degradable features.<sup>3,4</sup> From a biomedical point of view, poly(carbonate)s exhibit appealing degradation properties. Unlike poly(ester)s, APCs do not cause a local lowering of pH, which can be problematic for pH-sensitive drugs and the surrounding tissue.<sup>2</sup> The degradation of APCs occurs by surface erosion *in vivo* rather than the bulk degradation behaviour exhibited by poly(ester)s.<sup>2</sup>

In another context, isosorbide is an attractive building-block being a rigid, chiral, non-toxic and renewable replacement for petrochemical monomers such as bis-phenol A.<sup>5–7</sup> It is a sugar-based diol derived from the hydrolysis of starch, sucrose or cellulose, and has an annual production in tens of thousand tons.<sup>5</sup> Isosorbide produces polymers with a high  $T_g$

and great mechanical and optical properties.<sup>6</sup> Efforts to make poly(isosorbide) carbonates degradable have been made through the incorporation of di-acidic co-monomers to produce poly(ester-carbonate)s capable of enzymatic degradation when used with a poly(ester) content higher than 50%.<sup>6</sup> An alternative approach to increase the degradability of various co-polymers is reported in a pioneering study by Putnam and co-workers,<sup>8</sup> using the sugar-derivative dihydroxyacetone (DHA), a metabolic intermediate in the glycolysis cycle.<sup>4</sup> DHA can be synthesized by the selective oxidation of glycerol, an abundant waste product from biodiesel production.<sup>9</sup> The multiple forms of DHA in solution, together with the high reactivity of its ketone, limit the number of available polymerization conditions. Nevertheless, successful synthetic procedures have been reported towards poly(carbonate)s and poly(carbonate-ester)s.<sup>4</sup> Among them, the semi-crystalline poly(DHA) carbonate has proven difficult to process given its high melting point coinciding with the degradation temperature and it required co-monomers such as caprolactone and lactide (LA) to decrease the melting temperature.<sup>8,10</sup> In terms of biodegradability, the presence of DHA in several materials – including random co-polymers of LA-DHA, alternating aliphatic poly(ester)s, block co-polymers with DHA and poly(ethylene)glycol (PEG) – can be related to their degradation profile.<sup>8,11–13</sup> Surprisingly, to date little is known in regard to the mechanism behind the improved hydrolytic degradation exhibited by DHA.<sup>3</sup>

Our group recently reported on a procedure to generate and tailor the final configuration of APCs from bis-carbonylimid-

KTH Royal Institute of Technology, Department of Fibre and Polymer Technology, Teknikringen 56-58, SE-100 44, Stockholm, Sweden. E-mail: malkoch@kth.se

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azolide monomers of 1,3- and higher diols.<sup>14</sup> The monomers were attained on multi-gram scales through the activation of diols by 1,1'-carbonyldiimidazole (CDI), which readily precipitate out of solution and can be isolated by simple filtration. The resulting monomers were then polymerized through Fluoride Promoted Carbonylation (FPC) step-growth polymerization to afford a library of functional APCs. During the development of FPC polymerization, a competing ring-closing depolymerization mechanism was observed, which resulted in scrambling and in some cases an under-representation of 1,3-diols in the final polymer. This phenomenon was especially prevalent for the less nucleophilic diol isosorbide, with a final composition of 76/24 isosorbide to 2,2-dimethylpropane-1,3-diol. Considering the scrambling phenomenon as an advantage, it is apparent that random co-polymers can be achieved with the similarity of what is achieved with ring-opening polymerization (ROP), using step-growth polymerization from simpler and more easily scalable monomers.

In the present study, we tailor the degradability of isosorbide poly(carbonate)s through their copolymerization with DHA. High  $T_g$  amorphous poly(carbonate)s are accomplished that can generate high-modulus materials that degrade in neutral to weakly alkaline solutions. A thorough analysis on the pH-profile of model compound degradation by <sup>1</sup>H-NMR spectroscopy, together with a polymer degradation study by SEC and gravimetric measurements, serves to shed some light on the behaviour of this class of copolymers.

## Experimental

### Materials

1,1'-Carbonyldiimidazole (CDI), isosorbide (Iso) and dihydroxyacetone (DHA) were purchased from Carbosynth (Berkshire, UK). Isosorbide (98%) was purified by re-crystallization from acetone prior to use in polymerizations. All other chemicals were purchased from Sigma-Aldrich (St Louis, MO, USA) and used without further purification. HPLC grade solvents were used for all synthetic procedures, and dried over vacuum/heat-activated molecular sieves (3 Å) prior to use. Diazabicycloundecene (DBU) was dried and stored over 3 Å molecular sieves. All glassware and magnets were flame dried under vacuum and filled with argon prior to use. Dihydroxyacetone dimethylacetal ((MeO)<sub>2</sub>DHA) (**1**)<sup>15</sup> and isosorbide-biscarbonylimidazolide (Iso-bis-Im) (**2**)<sup>14</sup> were prepared according to published protocols, and complete structural characterization is reported in the ESI.†

### Methods

**NMR (<sup>1</sup>H and <sup>13</sup>C)** experiments were performed on a Bruker Avance 400 (100 MHz NMR instrument). <sup>1</sup>H-NMR spectra were referenced to the residual solvent peak of (CD<sub>3</sub>)<sub>2</sub>SO δ 2.50, CDCl<sub>3</sub> δ 7.26 and D<sub>2</sub>O δ 4.79, and <sup>13</sup>C-NMR spectra to (CD<sub>3</sub>)<sub>2</sub>SO δ 39.52 and CDCl<sub>3</sub> δ 77.16. Quantitative <sup>13</sup>C-NMR spectroscopy was performed with 200 mg sample in 0.8 mL (CD<sub>3</sub>)<sub>2</sub>SO with 20 mM chromium(III) acetylacetonate and a relaxation time of

10 s. 2D-NMR techniques including COSY, HSQC, HMBC, DEPT and NOESY were used to assign <sup>1</sup>H and <sup>13</sup>C shifts.

**Size exclusion chromatography (SEC)** was carried out on a TOSOH EcoSEC HLC-8320 GPC system equipped with an EcoSES RI detector and three columns from PSS GmbH (PSS PFG 5 μm; Microguard, 100 Å and 300 Å). A mobile phase of DMF with 0.01 M LiBr (0.2 mL min<sup>-1</sup>) at 50 °C was used together with a linear polymethylmethacrylate (PMMA) calibration method.

**Differential scanning calorimetry (DSC)** was performed on a Mettler Toledo DSC820, at a heating/cooling rate of 10 °C min<sup>-1</sup> under a nitrogen flow (50 mL min<sup>-1</sup>). The glass transition temperature ( $T_g$ ) was selected as the midpoint of transition on the second heating cycle (20 °C to 200 °C).

**Thermogravimetric analysis (TGA)** was performed on a Mettler Toledo TGA/DSC1, using the STARe software to process the data. A sample of 5 mg was placed in a 70 μL ceramic crucible and heated from 30 °C to 500 °C at a rate of 10 °C min<sup>-1</sup> under a nitrogen flow (50 mL min<sup>-1</sup>).

### Synthesis of compounds

**(MeO)<sub>2</sub>DHA-biscarbonylimidazolide ((MeO)<sub>2</sub>DHA-bis-Im) (**4**).** CDI (14.9 g, 91.8 mmol) was suspended in ethyl acetate (35 mL). (MeO)<sub>2</sub>DHA (**1**) (5.0 g, 36.7 mmol) was added portionwise to avoid cyclization of 1,3-diol. After the addition of (MeO)<sub>2</sub>DHA, the suspension turned into a yellow solution, followed by product precipitation. Solids were collected in a glass filter and washed with diethyl ether to afford a pure product as a white powder, 4.0 g (34%). δH (400 MHz; CDCl<sub>3</sub>) 8.16 (s, 2H, H5), 7.43 (s, 2H, H7), 7.10 (s, 2H, H6), 4.51 (s, 4H, CH<sub>2</sub> 1 and 3), 3.37 (s, 6H, COCH<sub>3</sub> 4' and 4''). δC (100 MHz; CDCl<sub>3</sub>) 148.1 (C8), 137.3 (C5), 131.2 (C6), 117.2 (C7), 98.4 (C2), 62.9 (C1 and C3), 49.0 (C4' and C4'').

**Ethyl-1-imidazolecarboxylate.** CDI (10.0 g, 61.7 mmol) was added to a mixture of ethanol (3.6 mL, 61.7 mmol) and 60 mL ethyl acetate and allowed to be stirred for 1 h. The resulting solution was filtered through a silica plug and eluted with excess ethyl acetate. The filtrate was concentrated *in vacuo* to afford the pure product as a transparent oil (5.15 g, 60%). Structural characterization of the product was in agreement with the previously published protocols (see the ESI†).<sup>16</sup>

**(MeO)<sub>2</sub>DHA-diethyl carbonate ((MeO)<sub>2</sub>DHA-(EC)<sub>2</sub>).** (MeO)<sub>2</sub>DHA (**1**) (2.0 g, 14.7 mmol), cesium fluoride (112 mg, 0.734 mmol) and ethyl-1-imidazolecarboxylate (5.15 g, 36.7 mmol) were mixed under neat conditions and allowed to be stirred overnight. Precipitated imidazole was filtered off and the product was purified by flash column chromatography (ethyl acetate/heptane), using an ethyl acetate gradient from 0% to 30%. The product was isolated as a transparent oil (3.49 g, 85%). δH (400 MHz; CDCl<sub>3</sub>) 4.21 (s, 4H, CCH<sub>2</sub>O 1 and 3), 4.20 (q, 4H, <sup>3</sup>J = 7.1 Hz, OCH<sub>2</sub>CH<sub>3</sub> 5' and H5''), 3.28 (s, 6H, OCH<sub>3</sub> 4' and 4''), 1.31 (t, 6H, <sup>3</sup>J = 7.1 Hz, OCH<sub>2</sub>CH<sub>3</sub> 6' and 6''). δC (100 MHz; CDCl<sub>3</sub>) 154.7 (O(CO)O 7' and 7''), 98.6 (C2), 64.4 (C5' and C5''), 63.0 (C1 and C3), 48.6 (C4' and C4''), 14.3 (C6' and 6'').



### DHA-diethylcarbonate (DHA-(EC)<sub>2</sub>, model-compound A).

Deprotection was adapted from procedures reported by Barton *et al.*<sup>17</sup> (MeO)<sub>2</sub>DHA-(EC)<sub>2</sub> (3.0 g, 10.7 mmol) and triphenylcarbenium tetrafluoroborate (Ph<sub>3</sub>CBF<sub>4</sub>) (3.53 g, 10.7 mmol) were added to 200 mL of dichloromethane and allowed to be stirred for 1 h. Aqueous NaHCO<sub>3</sub> (10%) was added to the stirring reaction mixture and later it was washed with distilled water. The organic phase was dried over Na<sub>2</sub>SO<sub>4</sub> and purified by flash column chromatography (ethyl acetate/heptane), using an ethyl acetate gradient from 0% to 35%. The product was isolated as a white powder (2.36 g, 94%). Structural characterization of the product was in agreement with the previously published protocols (see the ESI†).<sup>15</sup>

### Polymerization of poly(Iso-co-(MeO)<sub>2</sub>DHA)

All polymerizations were performed in 50–100 mL round-bottom flasks, fitted with magnetic stir bars and flame dried under vacuum. Varying feed ratios of (MeO)<sub>2</sub>DHA (**1**), bis-carbonylimidazolide (**2** or **4**) and isosorbide (**3**) were added in powder form (1 : 1 eq. diol to bis-carbonylimidazolide), sealed and cycled with vacuum/argon three times. Stock solutions of DBU and solvent (CHCl<sub>3</sub> or CH<sub>3</sub>CN) were dried overnight on vacuum/heat-activated 3 Å molecular sieves, and then added to the reaction vessel using a syringe. Final catalytic loading was 1 mol% DBU with regard to bis-carbonylimidazolide. Polymerisations in solvent were either conducted at room temperature or at 50 °C under magnetic stirring for 5 h to 16 h. Neat polymerization was conducted at 100 °C to 120 °C, by adding the catalyst directly to the reaction vessel and reacting for 3 h to 4 h. The DBU was subsequently quenched with 5 eq. of acetic acid and precipitated from CHCl<sub>3</sub> into methanol, forming a white powder precipitate, in scales of 2 to 10 g and yields of 87–97%.  $\delta$ H (400 MHz; CDCl<sub>3</sub>) 5.05–5.11 (2H, br m, H7 and H10), 4.89 (1H, br t, H9), 4.53 (1H, br d, H8), 4.29–4.17 (4H, br m, CH<sub>2</sub> 1 and 3), 4.10–3.97 (2H, br m, CH<sub>2</sub> 6), 3.96–3.85 (2H, br m, CH<sub>2</sub> 11), 3.27 (6H, br s, CH<sub>3</sub> 4).  $\delta$ C (100 MHz; CDCl<sub>3</sub>) 154.1–153.2 (m, O(CO)O), 98.3 (C2), 85.8 (C8), 81.6 (C7), 81.1 (C9), 77.3 (C10), 73.2 (C6), 70.6 (C11), 63.3 (CH<sub>2</sub>(CO) CH<sub>2</sub> 1 and 3), 48.6 (CH<sub>3</sub> 4).

### Deprotection of poly(Iso-co-(MeO)<sub>2</sub>DHA)

Deprotection of dimethyl ketal was performed using trityl tetrafluoroborate Ph<sub>3</sub>CBF<sub>4</sub> (1 eq.) in CH<sub>2</sub>Cl<sub>2</sub>.<sup>10,17</sup> To a round-bottom flask fitted with a magnetic stir bar, 300 mL of CH<sub>2</sub>Cl<sub>2</sub>, poly(Iso-co-(MeO)<sub>2</sub>DHA) (3.00 g, 8.98 mmol ketal units), Ph<sub>3</sub>CBF<sub>4</sub> (2.96 g, 8.98 mmol, 1 eq. per (MeO)<sub>2</sub>DHA repeating unit) and H<sub>2</sub>O (180  $\mu$ L, 8.98 mmol) were added. The reaction was allowed to be stirred at room temperature for 1 h. The solvent was subsequently removed *in vacuo* and purified by precipitation into methanol, affording poly(Iso-co-DHA) as a white powder (2.1 g, 82%).  $\delta$ H (400 MHz; (CD<sub>3</sub>)<sub>2</sub>SO) 5.07 (1H, br m, H6), 5.01 (1H, br m, H9), 4.97 (4H, br m, CH<sub>2</sub> 1 and 3), 4.85 (1H, br m, H7), 4.78 (1H, br m, H8), 3.99 (1H, br m, H10), 3.85 (3H, br m, H5, H5' and H10').  $\delta$ C (100 MHz; (CD<sub>3</sub>)<sub>2</sub>SO) 197.6 (C2), 153.5–153.2 (C4, C4'), 85.2 (C8), 81.2 (C9), 80.6 (C7), 77.3 (C6), 72.3 (C10), 70.3 (C5), 68.8 (C1 and C3).

### pH-Titration of the model compound (A)

For the hydrolysis study of the model compound DHA-diethyl carbonate (A), acetate and phosphate buffers were prepared in D<sub>2</sub>O between pH\* 3.8 and 9.0, where the pH\* notation refers to the pH value recorded using a standard glass electrode pH-probe in D<sub>2</sub>O. This is called the “cancel-out” approach, whereby the constant term (pD = pH\* + 0.4) used to convert the pH to pD is assumed to cancel out the increase in the pK<sub>a</sub> of acids in D<sub>2</sub>O.<sup>18,19</sup> Due to the limited solubility of A in D<sub>2</sub>O, experiments were carried out in a mixture of D<sub>2</sub>O and CD<sub>3</sub>CN. As a consequence of this, a second assumption was made, where it has been shown that pH scales linearly with the volume fraction of MeCN ( $\phi_{\text{MeCN}}$ ) added to an aqueous buffer through the function  $m_{\text{pH}}$  (eqn (1), see the ESI†).<sup>20</sup>

$${}^s_{\text{w}}\text{pH} - {}^w_{\text{w}}\text{pH} = m_{\text{pH}}\phi_{\text{MeCN}} \quad (1)$$

where  ${}^w_{\text{w}}\text{pH}$  refers to the pH in water and  ${}^s_{\text{w}}\text{pH}$  refers to the pH of the organic solvent and water mixture. A mixture of D<sub>2</sub>O (60% v/v) and CD<sub>3</sub>CN (40% v/v) with 50 mM acetate or phosphate buffers affords  $m_{\text{pH}}$  values of 2.28 or 1.71, respectively, to give the final pH\* range of 4.7 to 9.6.<sup>20</sup>

### Degradation study

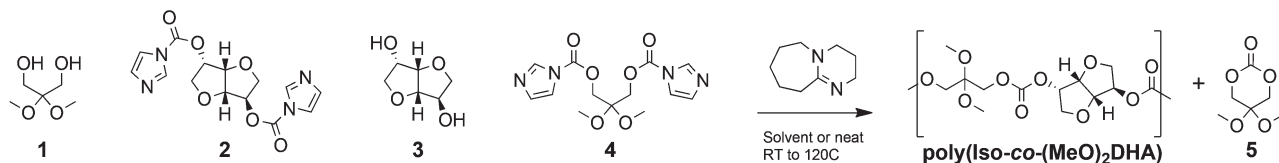
For each material, 100 mg of a finely ground polymer was pressed into a pellet and cut into four equal pieces of approximately 25 mg. The polymer samples were then submerged into 50 mM phosphate buffer (pH 8.0) and incubated at 37 °C for a week. Duplicates of the samples were isolated at different time intervals, lyophilized and analysed both gravimetrically and by SEC.

## Results and discussion

### Synthesis of poly(carbonate)s

With the aim of evaluating poly(Iso-co-DHA) carbonates and the effect the final architecture has on their properties, we relied on the step-growth polymerization of bis-carbonylimidazolides and diols to afford a library of rigid polycarbonates (Scheme 1 and Table 1). While the former study unfolded FPC chemistry using cesium fluoride (CsF) as an inorganic polymerization catalyst,<sup>14</sup> herein we evaluate the highly potent organocatalyst DBU commonly used for ring-opening polymerization.<sup>21</sup> In this regard, DBU was chosen in an effort to maximize the incorporation of DHA 1,3-diol into the final polymers. Kinetic experiments for the polymerization of bis-carbonylimidazolide and diols showed a satisfactory rate increase with DBU concentration, indicating that it is a viable catalyst (Fig. S1†). Polymerization reactions were carried out with a catalytic loading of 1 mol% with respect to diol, in either solvent (1.0 M monomer) or under neat conditions. Compared to CsF used in the previous study, polymerizations with DBU behaved in a similar manner, as seen for polymers **I** and **II** (Table 1). DHA's propensity to cyclize was most apparent in polar solvents such as acetone and acetonitrile (Fig. S2†). A





**Scheme 1** Step-growth polymerization using monomers **1** to **4** to form poly(Iso-co-(MeO)<sub>2</sub>DHA) carbonates, with the formation of cyclic monomer **5**.

**Table 1** Polymerization of diols (**1** and **3**) and bis-carbonylimidazolides (**2** and **4**) under various conditions

Polymer	Solvent <sup>a</sup>	Monomers	Time (h)	Cyclization ( <b>5</b> ) <sup>e</sup> (%)	DHA feed/final <sup>f</sup> (%)	$M_n$ (g mol <sup>-1</sup> )	$D$	Yield (%)
<b>I</b>	Neat <sup>b</sup>	<b>1</b> + <b>2</b>	4	6	50/48	9356	1.78	87
<b>II</b>	Neat <sup>c</sup>	<b>1</b> + <b>2</b>	3	8	50/47	7292	1.76	91
<b>III</b>	(CH <sub>3</sub> ) <sub>2</sub> CO	<b>1</b> + <b>2</b>	16	11	50/45	4556	1.53	87
<b>IV</b>	CH <sub>3</sub> CN	<b>1</b> + <b>2</b>	16	10	50/45	9231	1.99	90
<b>V</b>	CHCl <sub>3</sub>	<b>1</b> + <b>2</b>	16	4	50/48	7857	1.83	97
<b>VI</b>	CHCl <sub>3</sub> <sup>d</sup>	<b>1</b> + <b>2</b> + <b>3</b>	4	10	25/21	5378	1.84	94
<b>VII</b>	CHCl <sub>3</sub>	<b>3</b> + <b>4</b>	16	2	50/50	7153	1.65	92
<b>VIII</b>	CHCl <sub>3</sub>	<b>1</b> + <b>3</b> + <b>4</b>	16	4	75/74	5790	1.49	88

<sup>a</sup> Unless otherwise stated, reactions were carried out at room temperature using DBU (1 mol%) as a catalyst. <sup>b</sup> Reaction was carried out at 120 °C using DBU as a catalyst. <sup>c</sup> Reaction was carried out at 100 °C using CsF as a catalyst. <sup>d</sup> Reaction was conducted at 50 °C. <sup>e</sup> <sup>1</sup>H-NMR integration of  $\delta$  4.28 and 3.31 ppm. <sup>f</sup> <sup>1</sup>H-NMR integration of the Iso-segment at  $\delta$  4.88 and the DHA-segment at 4.21 ppm.

similar behaviour was also observed in the case of high isosorbide content polymer **VI** at an elevated temperature (50 °C) in chloroform.

A closer look at the configuration of polymers using <sup>13</sup>C-NMR spectroscopy reveals significant scrambling when using monomer **1** (Fig. 1). Dyad analysis of carbonyls with <sup>13</sup>C-NMR spectra indicates that the reaction of monomers **3** and **4** to generate polymer **VII** shows a strict alternating sequence with two observed shifts at  $\delta$  154.0 and 153.8 ppm, corresponding to the *endo*- and *exo*-configuration of isosorbide. However, the integration of carbonyl shifts using the inverse-gated decoupling <sup>13</sup>C-NMR spectra of polymers **V** and **VI**, accomplished from monomers **1**, **2** and **3**, indicates statistically random co-polymers (Table S1†). This competing depolymerization mechanism was observed for both catalysts when a terminal 1,3-diol was present in the system.<sup>14</sup> The highest degree of incorporation of residual six-membered monomer **5** was observed when using chloroform. With this in mind, the final properties of polymers can be tailored to fit the specific needs of the application.

Deprotection of methyl-ketal in poly(Iso-co-(OMe)<sub>2</sub>DHA) was evaluated through the previously reported acetal deprotection protocols.<sup>10,15</sup> Mixtures of TFA and water led to significant degrees of deprotection with concomitant hydrolysis as reported by Putnam and co-workers.<sup>15</sup> The most satisfactory results were achieved with trityl tetrafluoroborate Ph<sub>3</sub>CBF<sub>4</sub> and water (1:1) in dichloromethane.<sup>10,17</sup> Trityl-deprotection limited the acid-catalyzed hydrolysis of carbonates and showed the visual precipitation of the polymer with increasing DHA-content. This procedure afforded 80–90% deprotection, confirmed by <sup>1</sup>H-NMR and <sup>13</sup>C-NMR spectra. After precipitation into cold methanol, poly(Iso-co-DHA) carbonates were isolated

as glassy white powders with  $M_n$  in the range of 4500–9350 and PDI values in the range of 1.49–1.99.

### Thermal properties

Thermal analysis of the co-polymers revealed the stark effect each sugar-derived monomer has on the final performance of the material. The keto-functional DHA and the bicyclic isosorbide provide a roughly a 100 °C difference in  $T_g$  for the two homo-polymers, 68 °C and 169 °C respectively.<sup>8,14</sup> Differential scanning calorimetry revealed that all co-polymers were amorphous with a single glass transition. Deprotection of poly(Iso-co-(OMe)<sub>2</sub>DHA) led to an increase in  $T_g$ . For the deprotected poly(Iso-co-DHA) carbonates, an increasing trend in  $T_g$  was observed in the range from 80 °C to 127 °C, going from 25% (**VIII-d**) to 75% (**VI-d**) isosorbide content (Fig. 2 and Table 2). Deprotected polymers showed some instability at temperatures above the  $T_g$  which is consistent with thermal degradation. This was further verified by thermogravimetric analysis.

In terms of thermal degradation, TGA revealed an increasing trend in the onset of degradation,  $T_{d-5\%}$  values (corresponding to 5% mass loss), with increasing isosorbide content (Fig. 3). Looking at the 75% DHA polymer (**VIII-d**) a sharper decrease was observed at lower temperatures leading to a maximum rate of degradation ( $T_{d-max}$ ) of 237 °C from the more thermally labile blocky DHA configuration. This is consistent with the results from Simon *et al.* for low crystallinity copolymers with  $\epsilon$ -caprolactone and DHA, having lower  $T_{d-5\%}$  values for higher DHA content.<sup>10</sup> For the 50/50 random co-polymer (**V-d**) a bimodal degradation behaviour was observed with two  $T_{d-max}$  values at 288 °C and 368 °C (Fig. 3). This behaviour is presumably due to its random configuration yielding short blocks of the two monomers. A comparison between the





the hydrolysis of esters has been discussed to a great extent in the literature.<sup>22–26</sup> Several studies have shown that the intramolecular attack of hydrated  $\beta$ -carbonyls can significantly increase the rate of hydrolysis.<sup>26</sup>

### Hydrolysis of the model-compound DHA(EC)<sub>2</sub>

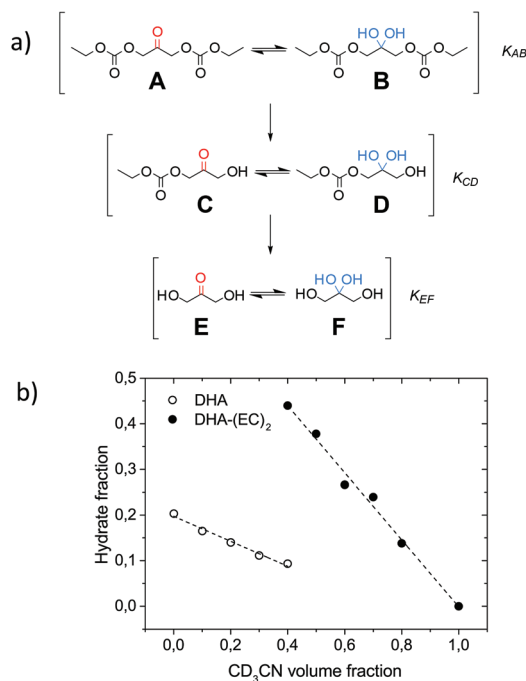
To better understand the behavior of DHA-containing poly(carbonate)s, the diethyl carbonate analogue of DHA (**A**) was studied using <sup>1</sup>H-NMR spectroscopy in D<sub>2</sub>O and CD<sub>3</sub>CN (Fig. 4a). DHA is known to co-exist in its keto, hydrate and enol forms.<sup>27</sup> When mixed in D<sub>2</sub>O/CD<sub>3</sub>CN both DHA (**E**) and DHA-(EC)<sub>2</sub> (**A**) instantly form an equilibrium between the keto- and hydrate forms. The CH<sub>2</sub>  $\alpha$ -protons of the various DHA-forms in the <sup>1</sup>H-NMR spectra show a downward shift of about  $\delta$  0.8 ppm between their keto- and hydrate forms.<sup>27</sup> This is also observed in the case of diethyl carbonate **A** ( $\delta$  4.82 ppm) to **B** ( $\delta$  4.04 ppm), and DHA **E** ( $\delta$  4.26 ppm) to **F** ( $\delta$  3.42 ppm) (see the ESI† for assignments). No enol-form could be observed through <sup>1</sup>H-NMR spectroscopy and this was further confirmed by HSQC and HMBC 2D-NMR analysis (assignments of NMR spectra can be found in the ESI†).

In order to investigate the influence that the overall polarity of the medium has on the keto/hydrate equilibrium, a series of dilution tests were performed using D<sub>2</sub>O and CD<sub>3</sub>CN. Fig. 4b indicates that DHA (**E**), being predominantly water soluble, decreases its concentration upon dilution with CD<sub>3</sub>CN in D<sub>2</sub>O, going from a ratio of 4 : 1 as reported by Davis<sup>27</sup> to a ratio of

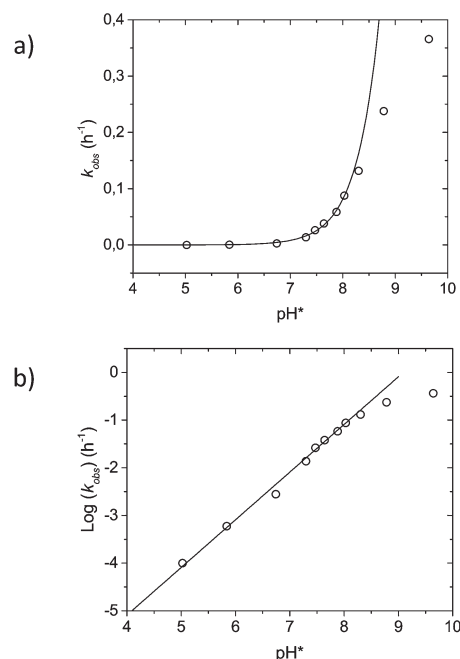
9 : 1, affording the equilibrium constant  $K_{EF} = [F]/[E] = 0.11$ , in D<sub>2</sub>O/CD<sub>3</sub>CN ( $\theta_{CD_3CN} = 0.4$ ). The predominantly acetonitrile soluble DHA-(EC)<sub>2</sub> (**A**) increases its concentration upon dilution with D<sub>2</sub>O in CD<sub>3</sub>CN, from the 100% keto-form in CD<sub>3</sub>CN to the 44% hydrate form in D<sub>2</sub>O/CD<sub>3</sub>CN ( $\theta_{CD_3CN} = 0.4$ ),  $K_{AB} = [B]/[A] = 0.80$ . The major difference between the two equilibria is the presence of two electron-withdrawing carbonate groups in diethyl carbonate (**A**), while DHA (**E**) contains two electron-donating hydroxyl groups, meaning that the carbons in **A** are significantly more electron-poor than those in **E**. The keto/hydrate equilibrium was not affected by alterations in the pH\* range studied from 4.7 to 9.6, using 50 mM acetate- and phosphate-buffer solutions in D<sub>2</sub>O/CD<sub>3</sub>CN ( $\theta_{CD_3CN} = 0.4$ ).

Degradation studies by <sup>1</sup>H-NMR spectroscopy in D<sub>2</sub>O/CD<sub>3</sub>CN ( $\theta_{CD_3CN} = 0.4$ ) revealed a third equilibrium constant between mono-ethyl carbonates **C** and **D**,  $K_{CD} = [D]/[C] = 0.26$ . Under the conditions used, adding one carbonate to DHA doubles the concentration of hydrate in solution. On the other hand, having two electron-withdrawing carbonates (**A**) will quadruple the amount of hydrate formed in solution, as compared to DHA (**E**).

To investigate further which form of DHA causes rapid hydrolysis, kinetic studies were performed on the diethyl carbonate analogue (**A**) of DHA in the pH\* range of 4.7 to 9.6. Kinetic data for the formation of ethanol fit well with a pseudo first-order reaction with increasing rates going from neutral to alkaline conditions (Fig. 5a), with a first order dependence on [OD<sup>-</sup>] (Fig. 5b). If the carbonate hydrolysis were to go through



**Fig. 4** (a) Keto/hydrate equilibria of the model compound (**A/B**) and degradation products monoethyl carbonate (**C/D**) and DHA (**E/F**). (b) Amount of hydrate as a function of CD<sub>3</sub>CN fraction in D<sub>2</sub>O (lines are included as a visual guide). Equilibrium constants in D<sub>2</sub>O/CD<sub>3</sub>CN ( $\theta_{CD_3CN} = 0.4$ , 17 mM) were found to be 0.80 ( $K_{AB}$ ), 0.26 ( $K_{CD}$ ) and 0.11 ( $K_{EF}$ ).

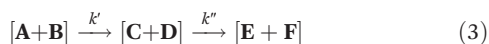
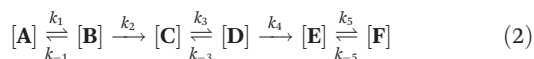


**Fig. 5** pH-Profile for the hydrolysis of DHA-(EC)<sub>2</sub> (**A**) in D<sub>2</sub>O/CD<sub>3</sub>CN ( $\theta_{CD_3CN} = 0.4$ ). (a) Observed pseudo first-order rate constants vs. pH\*, (b) log–log plot of observed pseudo first-order rate constants, showing a second-order overall rate constant  $k_{EtOH} = 0.82 \times 10^5$  (M<sup>-1</sup> h<sup>-1</sup>) for the formation of ethanol.



an intermediate enol-form, significant deuteration of CH<sub>2</sub> protons in the  $\alpha$ -position would be observed.<sup>28</sup> This was not the case, suggesting that the seemingly hydrolytically unstable nature of these compounds likely stems from the hydrated form.

For pH\* in the range of 5.0 to 8.0, reactions conform well to an equilibrium approximation, meaning that the rates of the irreversible steps ( $k_2$  and  $k_4$ , eqn (2)) can be seen as significantly slower than those of the reversible hydration reactions. This means that the kinetics can be seen as a two-step consecutive reaction (eqn (3)), where the concentration at any given point in time is given by  $[A]_0 = [A + B]_t + [C + D]_t + [E + F]_t$ , using the equilibrium constants ( $K_{AB}$ ,  $K_{CD}$  and  $K_{EF}$ ) to solve for each individual component. At pH\* values above 8.0, reactions deviated from equilibrium over time.



The rate of the first irreversible step ( $k'$ ) is taken from the rate consumption of diethyl carbonates A and B, with known initial concentrations  $[A]_0$  (see the ESI†). Using the observed rate constants ( $k_{obs}$  for A and B), a second-order overall rate constant can be determined with respect to  $[OD^-]$  and  $[A + B]$ ,  $k' = 1.31 \times 10^5 \text{ (M}^{-1} \text{ h}^{-1})$  (Fig. S3†). The second irreversible step was evaluated by fitting the integrated rate law to the experimental data (Fig. S4†). The best fit to the data suggests that the second step ( $k''$ ) has a slower rate than the first step ( $k'$ ). This is consistent with the equilibrium of the mono-ethyl-carbonate being shifted towards the keto-form, making  $k''$  the rate limiting step in the formation of E.

Overall, these results would suggest that the hydrolytic instability is highly dependent on the ability of substrates to become hydrated. Neighbouring group participation has been showcased for poly(glycerol) carbonates, where the poly(1,2-glycerol) carbonate with a pendant primary hydroxyl group increases the degradation rates in solution.<sup>29,30</sup> In a recent study by Ricapito *et al.* on poly(ethylene glycol) (PEG) and poly(DHA) carbonate hydrogels, it was shown that having an extra methylene group on each side, in the case of 1,5-dihydroxypentan-3-one, provided significantly slower degradation compared to hydrogels formed with PEG and DHA-carbonates.<sup>12</sup> As the dilution study suggested, decreasing the polarity of the surrounding medium and decreasing the electron-withdrawing effect on the  $\alpha$ -carbons limit the ability to form hydrates, which subsequently affects the hydrolytic degradation of the compound. In the polymer case, being confined to a hydrophobic micro-environment together with limited conformational mobility, chemical degradation rates are expected to be less pronounced. The hydrolytic degradation of the polymers is assessed in the following section.

### Hydrolytic degradation of poly(Iso-co-DHA) polycarbonates

Based on the insights gained from model compound studies, further degradation investigations were carried out on poly-

mers in their solid state. Previous studies on LA-DHA co-poly (ester-carbonate)s have indicated an increased degradation behaviour with increasing DHA content, with more surface-like erosion characteristics.<sup>8</sup> Pellets of co-polymers poly(Iso-*ran*-DHA) with varying DHA contents from 25% to 75% (V-d, VI-d and VIII-d) and the alternating version of poly(Iso-*alt*-DHA) containing 50% DHA (VII-d) were exposed to an accelerated aging study in 50 mM phosphate buffer at pH 8.0 and 37 °C. Being first order in  $[OH^-]$ , going from physiological pH 7.4 to 8.0, the chemical degradation rates are expected to increase by a factor of 4.

Within 1 day the 75% DHA containing (VIII-d) poly(Iso-*co*-DHA) had lost most of its mass (Fig. 6a). After 5 hours of immersion in aqueous buffer VIII-d became insoluble in DMF, affording swollen transparent gel-like pieces. The random poly(Iso-*ran*-DHA) 50% (V-d) and 25% (VI-d) DHA containing

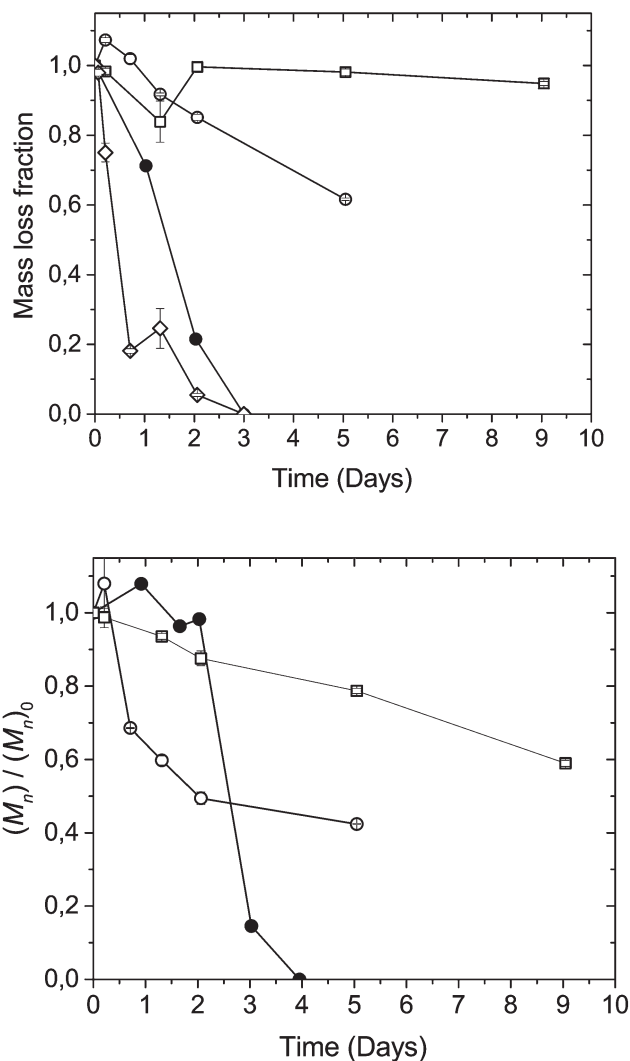


Fig. 6 Fractional mass loss (a) and  $M_n$  loss by SEC (b) for the hydrolytic degradation of poly(Iso-*co*-DHA); VI-d 75–25 (□), V-d 50–50 (○), VIII-d 25–75 (◇), and the alternating VII-d 50–50 (●), in 50 mM phosphate buffer at pH 8 and 37 °C.





pellets remained physically intact with low mass loss over the first 2 days. The 50% co-polymer **V-d** showed a faster trend in terms of molecular weight loss compared to its 25% (**VI-d**) counterpart, indicating a traditional surface erosion type of degradation.<sup>2,6</sup>

The alternating poly(Iso-*alt*-DHA) (**VII-d**) displayed a different type of degradation behaviour. Pellets of **VII-d** showed little molecular weight loss in the first 2 days (Fig. 6a). However, pellets showed continuous mass loss, followed by rapid degradation upon swelling in the buffer solution. After 2 days the pellets had completely disintegrated, presumably due to increases in stress and crack formation due to water uptake, leading to increased plasticization and rapid hydrolysis. Endo and co-workers have reported radical differences in the solubility of polystyrene with pendant tri-carbonyls upon the formation of its hydrate form.<sup>31</sup> The alternating analogue (**VII-d**) consists of one type of carbonate linkage, namely from DHA to isosorbide, with a more limited conformational mobility compared to its blocky random counterpart (**V-d**). The alternating analogue should thus be less prone to an intramolecular nucleophilic attack, given that each DHA-unit is anchored to the more rigid isosorbide. The random co-polymers contain three types of carbonates: DHA–DHA, DHA–isosorbide and isosorbide–isosorbide (Fig. 1). These three types of linkages are expected to have different probabilities of hydrolysis by chemical degradation. The DHA–DHA carbonate linkages should be more prone to random chain-scission, having greater conformational mobility and two adjacent  $\beta$ -carbonyls per carbonate, according to what was observed related to thermal stability in TGA measurements (Fig. 3). Previous studies have shown that DHA polycarbonates pose high surface energy, with an especially high polar contribution, presumably from the ketone.<sup>15</sup> These findings would suggest that a higher DHA content provides a more polar microenvironment. Comparing the high isosorbide content polymer **VI-d** (25% DHA) with the alternating **VII-d** (50% DHA), with little to no repeating DHA–DHA carbonates, it would seem that the higher isosorbide content co-polymer is less prone to water uptake, as can be seen in Fig. 6, both in terms of mass loss and molecular weight changes over time. The results suggest that not only does the amount of the labile DHA monomer affect the degradation behavior of the final polymer construct, but also that the distribution along the polymer chain will greatly affect water-uptake as a consequence. Furthermore, scrambled polymers with dyad sequences similar to those achieved by ROP protocols show a greater stability over time in a weakly alkaline buffer.<sup>8,32</sup>

## Conclusions

Rigid, amorphous and degradable poly(carbonate)s have been produced from the sugar-based isosorbide and the sugar metabolic derivative dihydroxyacetone (DHA). In order to tailor the final properties, the bi-cyclic chiral isosorbide diol proved to be excellent for achieving high  $T_g$  amorphous co-polymers.

The keto-functional DHA was also shown to provide the hydrolytic instability necessary to make the poly(carbonate)s degradable. Hydrolytic instability presented by the DHA co-monomer likely stems from carbonates having  $\beta$ -carbonyl neighbouring groups, capable of forming hydrates. Model compound studies indicate that ketones with two adjacent carbonates show a large degree of hydration, with the loss of one carbonate leading to the equilibrium being shifted towards the keto form. The pH-profile of the hydrolysis reaction indicates that it is first order in  $[\text{OH}^-]$  affording substantial hydrolysis going from neutral to weakly alkaline conditions. In terms of polymer degradation of solid pellets, the rate of random chain-scission increases with DHA content, providing degradation rates greater than conventional poly(carbonate)s and even poly(ester)s. Changing the monomer configuration had a marked effect on the degradation profiles, as the alternating configuration indicated little initial chain scission, with greater initial water uptake. This behaviour may be attributed to the fact that chains have limited mobility, with an even distribution of the polar DHA, capable of forming hydrates along the polymer. Rapid hydrolysis was only observed after the pellets had swollen and disintegrated. The co-polymers containing more blocky regions of isosorbide showed less initial mass loss due to water uptake followed by a gradual molecular weight loss over time. These rigid, amorphous and degradable poly(carbonate)s are seen as good candidates for the bone restoration field. In this field, it is necessary to provide initial mechanical support and ideally with high-modulus properties that match the surrounding mineral tissue followed by induced macroporosity for potential tissue ingrowth.

## Conflicts of interest

There are no conflicts to declare.

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## Notes and references

- 1 J. Xu, E. Feng and J. Song, *J. Appl. Polym. Sci.*, 2014, **131**, 39822.
- 2 S. Tempelaar, L. Mespouille, O. Coulembier, P. Dubois and A. P. Dove, *Chem. Soc. Rev.*, 2013, **42**, 1312–1336.
- 3 D. K. Schneiderman and M. A. Hillmyer, *Macromolecules*, 2017, **50**, 3733–3749.
- 4 N. G. Ricapito, C. Ghobril, H. Zhang, M. W. Grinstaff and D. Putnam, *Chem. Rev.*, 2016, **116**, 2664–2704.



- 5 C. Dussenne, T. Delaunay, V. Wiatz, H. Wyart, I. Suisse and M. Sauthier, *Green Chem.*, 2017, **19**, 5332–5344.
- 6 F. Fenouillot, A. Rousseau, G. Colomines, R. Saint-Loup and J. P. Pascault, *Prog. Polym. Sci.*, 2010, **35**, 578–622.
- 7 M. Rose and R. Palkovits, *ChemSusChem*, 2012, **5**, 167–176.
- 8 J. R. Weiser, P. N. Zawaneh and D. Putnam, *Biomacromolecules*, 2011, **12**, 977–986.
- 9 R. M. Painter, D. M. Pearson and R. M. Waymouth, *Angew. Chem., Int. Ed.*, 2010, **49**, 9456–9459.
- 10 J. Simon, J. V. Olsson, H. Kim, I. F. Tenney and R. M. Waymouth, *Macromolecules*, 2012, **45**, 9275–9281.
- 11 J. N. Korley, S. Yazdi, K. McHugh, J. Kirk, J. Anderson and D. Putnam, *Biomaterials*, 2016, **98**, 41–52.
- 12 N. G. Ricipito, J. Mares, D. Petralia and D. Putnam, *Macromol. Chem. Phys.*, 2016, **217**, 1917–1925.
- 13 P. N. Zawaneh, S. P. Singh, R. F. Padera, P. W. Henderson, J. A. Spector and D. Putnam, *Proc. Natl. Acad. Sci. U. S. A.*, 2010, **107**, 11014–11019.
- 14 J. V. Olsson, D. Hult, S. García-Gallego and M. Malkoch, *Chem. Sci.*, 2017, **8**, 4853–4857.
- 15 A. N. Zelikin, P. N. Zawaneh and D. Putnam, *Biomacromolecules*, 2006, **7**, 3239–3244.
- 16 M. Lecouvey, C. Dufau, D. El Manouni and Y. Leroux, *Nucleosides Nucleotides*, 1999, **18**, 2109–2120.
- 17 D. H. R. Barton, P. D. Magnus, G. Smith, G. Streckert and D. Zurr, *J. Chem. Soc., Perkin Trans. 1*, 1972, 542–552.
- 18 K. H. Scheller, V. Scheller-Krattiger and R. B. Martin, *J. Am. Chem. Soc.*, 1981, **103**, 6833–6839.
- 19 A. Krężel and W. Bal, *J. Inorg. Biochem.*, 2004, **98**, 161–166.
- 20 X. Subirats, M. Rosés and E. Bosch, *Sep. Purif. Rev.*, 2007, **36**, 231–255.
- 21 N. E. Kamber, W. Jeong, R. M. Waymouth, R. C. Pratt, B. G. G. Lohmeijer and J. L. Hedrick, *Chem. Rev.*, 2007, **107**, 5813–5840.
- 22 J. A. Walder, R. S. Johnson and I. M. Klotz, *J. Am. Chem. Soc.*, 1978, **100**, 5156–5159.
- 23 W. N. Washburn and E. R. Cook, *J. Am. Chem. Soc.*, 1986, **108**, 5962–5964.
- 24 M. L. Bender and M. S. Silver, *J. Am. Chem. Soc.*, 1962, **84**, 4589–4590.
- 25 F. Ramirez, B. Hansen and N. B. Desai, *J. Am. Chem. Soc.*, 1962, **84**, 4588–4588.
- 26 K. Bowden, *Chem. Soc. Rev.*, 1995, **24**, 431–435.
- 27 L. Davis, *Bioorg. Chem.*, 1973, **2**, 197–201.
- 28 S. J. Reynolds, D. W. Yates and C. I. Pogson, *Biochem. J.*, 1971, **122**, 285–297.
- 29 H. Zhang and M. W. Grinstaff, *J. Am. Chem. Soc.*, 2013, **135**, 6806–6809.
- 30 J. Geschwind and H. Frey, *Macromolecules*, 2013, **46**, 3280–3287.
- 31 T. Dei, K. Morino, A. Sudo and T. Endo, *J. Polym. Sci., Part A: Polym. Chem.*, 2011, **49**, 2245–2251.
- 32 J. R. Weiser, A. Yueh and D. Putnam, *Acta Biomater.*, 2013, **9**, 8245–8253.

