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Latest perspectives of orally bioavailable 2,4-diarylaminopyrimidine analogues (DAAPalogues) as anaplastic lymphoma kinase inhibitors: discovery and clinical developments

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The course of anaplastic lymphoma kinase (ALK)-rearranged non-small-cell lung cancer (NSCLC) therapy has improved impressively. The Food and Drug Administration (FDA) has approved crizotinib (Xalkori, Pfizer) as a first-in-class tyrosine kinase inhibitor (TKI) that demonstrated a substantial objective response rate (ORR) and remarkable progression-free survival (PFS). However, acquired resistance to crizotinib is still a major concern especially as the central nervous system (CNS) remains the most common sites of relapse. To combat disease resistance, limited PFS and poor CNS exposure exhibited by crizotinib (Xalkori, Pfizer) led to the discovery of numerous next generation ALK-TKIs and surprisingly most of them are 2,4-Diarylaminopyrimidine Analogues (DAAPalogues). To date, DAAPalogues have been investigated extensively to display their superior potency against numerous kinase targets especially ALK/ROS1. This review describes hit-to-drug evolution strategies, activity spectra, milestones related to medicinal chemistry discovery efforts and scalable synthetic pathways of clinically emerging DAAPalogues which are either progressing as investigational or preclinical candidates. In addition, the significance of DAAPalogues to treat the patients with ALK⁺-NSCLC in clinical settings has been detailed. This review is beneficial for medicinal chemists and researchers contributing to discovering ALK-TKIs to overcome existing issues related to DAAPalogues in the drug discovery process.

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1 Introduction

Lung cancer is a fatal malignancy that has a higher mortality rate than colorectal, breast, and prostate malignancies.^{1–5} Lung cancer is associated with cancer related deaths globally with a truncated survival rate of about 15%. Smoking addiction is reported to be the predominant risk factor.^{6–10} Historically, various classifications of lung cancer have been reported but clinically, lung cancers are categorized into small-cell lung cancer (SCLC) and non-small cell lung cancer (NSCLC). Reports have revealed that SCLC is responsible for 10–15% while NSCLC is accountable for 85–90% of lung malignancies. NSCLC, typically includes squamous cell carcinoma (10–15%), adenocarcinoma (25–30%) and large cell carcinoma (10–15%) as its three key subtypes. Among these the adenocarcinoma accounts for about 40% of NSCLC cases whereas squamous cell carcinoma is

reported to be responsible for about 25% of cases.^{11–14} Despite the devoted investigations to improve the diagnosis of lung carcinoma and treatment, the five year survival rate for patients detected with NSCLC remains very low (10–15%). In spite of technological advancements in diagnostics and its availability, more than 50% patients are diagnosed at an advanced stage. It is pertinent to note that the therapy demonstrates a palliative aspect instead of a curative intent.^{15–17} The lung cancer therapy, especially NSCLC has been revolutionized remarkably due to the emergence of molecular targeted therapy (MTT).¹⁸ The emergence of activated gene rearrangements in ALK as an oncogenic driver of NSCLC patients (3–7%) have made the ALK a potential therapeutic target in medicinal chemistry discovery campaigns around the globe. The purpose of current article is to review hit-to-drug evolution strategies, activity spectrum, milestones related to medicinal chemistry discovery efforts and

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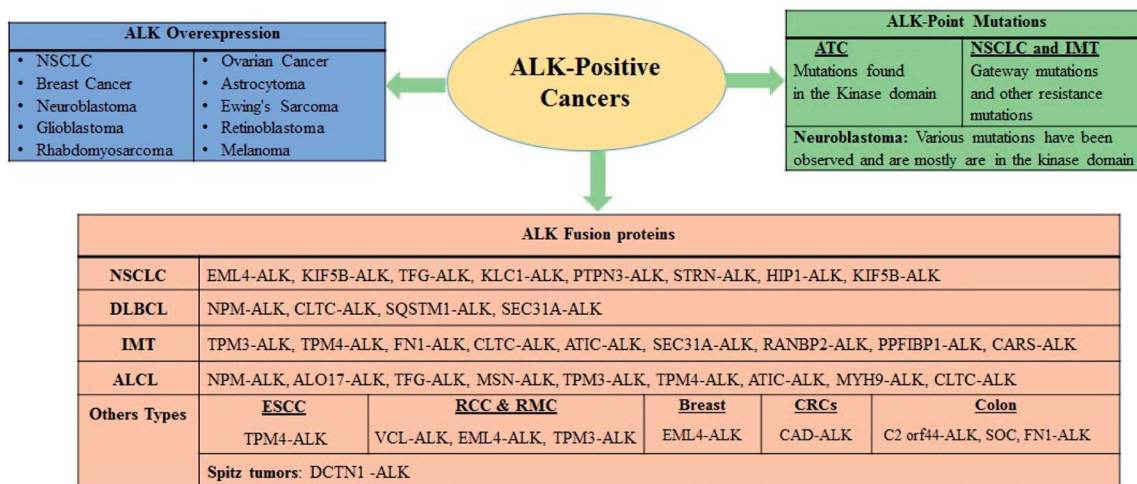


Fig. 1 An overview of ALK in cancer and schematic summary of ALK overexpression, point mutations, and fusions in ALK positive cancer.

scalable (pilot plant) synthetic pathways of clinically emerging DAAPalouges. In addition, the significance of DAAPalouges to treat the patients with ALK⁺-NSCLC in clinical settings has been detailed.

2 ALK in non-small cell lung cancer

ALK is a receptor tyrosine kinase (RTK) and a key member of the insulin receptor protein-tyrosine kinase superfamily. It was first discovered in 1994 in the anaplastic large-cell lymphoma (ALCL) as a non-Hodgkin lymphoma subtype.^{19–21} ALK is a novel orphan RTK which is considered to play an essential role in usual growth and in function of nervous system. The reports have revealed that ALK is usually expressed during the growth of nervous tissues and also down-modulates in the adult.^{22–24} ALK is closely correlated to leukocyte tyrosine kinase (LTK)^{25–29} and

has been found to implicate in numerous solid and hematologic cancers. Considering oncogenic manifestations, the ALK rearrangements were first identified as a fusion to a portion of the nucleophosmin (NPM) gene in 60% of ALCL, a small subset of non-Hodgkin lymphomas.^{20,30} It has been revealed that in lymphoma cells containing t(2;5), the NPM gene plays a critical role as an active promoter to initiate or facilitate the expression of the ALK catalytic domain. It has also been envisioned that the truncated ALK is deregulated and phosphorylates the intracellular molecules (substrate) to prompt further malignant transformations.^{31–33} Thus new NPM-ALK fusion protein was discovered while screening the patients associated with ALCL malignancy where the N-terminal portion of NPM was found to fuse with the kinase domain of ALK as a consequence of t(2;5)(p23;q35) translocation.^{34,35}

Since the identification of novel potential mutation driver observed in 5–10% NSCLC cases several other ALK fusion genes have also been identified recently. The fusion of the echinoderm microtubule-associated protein-like 4 gene (EML4) with the ALK gene resulted in the generation of a fusion protein (EML4-ALK) and in NSCLC it is considered to be the most significant discovery being an oncogenic factor in solid tumors.³⁶ EML4-ALK fused protein was identified in 2007 in the Japanese patients with NSCLC.^{37–40} Surprisingly, since the preliminary reports regarding the discovery of the NPM-ALK fusion protein in clinical settings pertaining to the patients investigated with ALCL, a plethora of ALK fusion proteins have been described in NSCLC,^{37,41} inflammatory myofibroblastic tumors (IMTs),^{42,43} squamous cell carcinoma (SCC) of the oesophagus,^{44,45} renal cell carcinoma,⁴⁶ and diffuse large B-cell lymphomas (DLBCLs)⁴⁷ (Fig. 1). Furthermore, oncogenic progression *via* overexpression and point mutations has been investigated in various cancers.^{48–54}

Physiologically, the function of ALK in mammals is still under investigation and unclear. However, the significance and key role of full-length ALK-RTK in the development or onset of numerous diseases is well investigated and consequently



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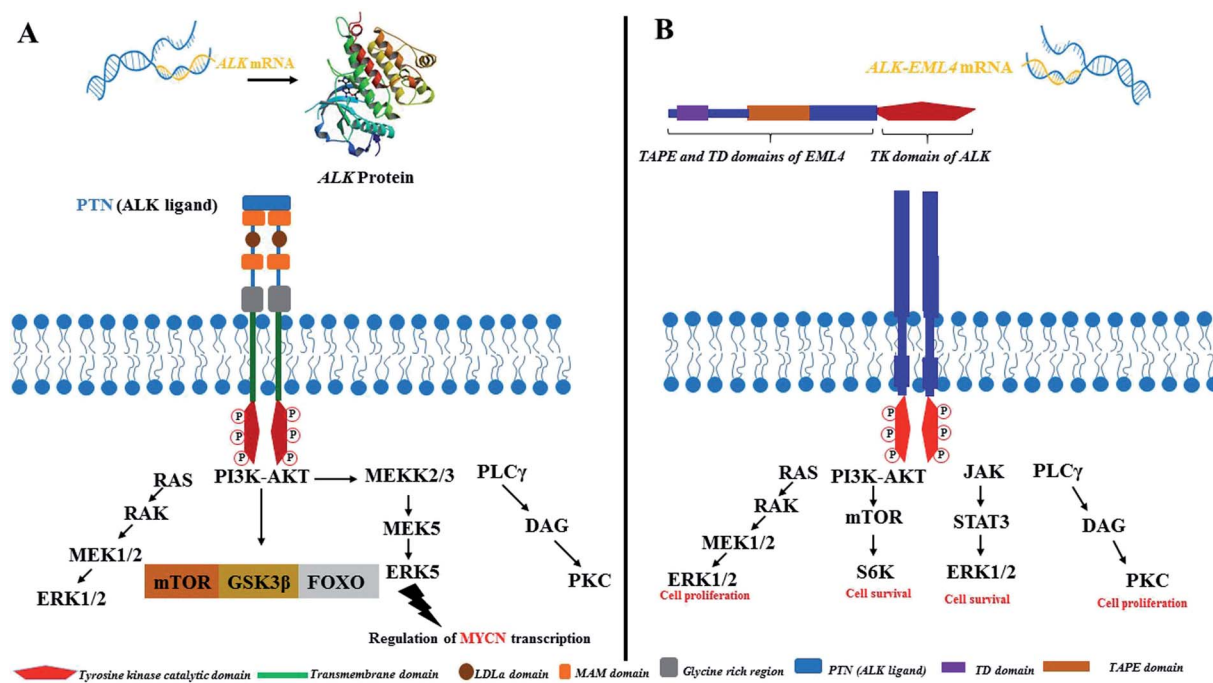


Fig. 2 The anaplastic lymphoma kinase receptor tyrosine kinase. (A) Schematic representation of pleiotrophin (PTN) bound ligand activated anaplastic lymphoma kinase (ALK) receptor signalling. (B) Constitutively active ALK receptor signalling due to EML4 fusion. The fusion arises due to paracentric inversion between EML4 and ALK genes located in the short arm of chromosome 2. EML4-ALK translocation is thought to lead to the constitutive activation of oncogenic signaling through multiple pathways such as PI3K/Akt, JAK/STAT, and RAS/RAF/MEK/ERK. Aberrant activation of PI3K-Akt initiate mTOR, FOXO and GSK3 β signalling which lead to survival and proliferation of tumour cells. PI3K/Akt activation also lead to MEKK2/3/MEK5/ERK5 pathway which subsequently promote the expression of the oncogene MYCN. The STAT3 signaling pathway has shown a key role in NPM-ALK mediated alteration however the significance of STAT3 activation in EML4-ALK⁺-NSCLC is unclear. ALK has also been shown to phosphorylate PLC γ to mediate its mitogenicity. (mTOR; mechanistic target of rapamycin, S6K; ribosomal S6 kinase, TD; trimerisation domain, TAPE; tandem atypical propeller domain, MAM; meprin, A5 protein, and protein tyrosine phosphatase Mu domain).

known to be expressed in several types of tumors including NSCLC (Fig. 2). Numerous signal transduction pathways have been detected which are activated by the involvement of ALK. Initially, fusion of ALK ligand to extracellular domain occurs during the signal transduction process followed by dimerization and trans-phosphorylation which in turn prompts cellular process involved in oncogenesis. ALK signaling *via* several downstream pathways have been reported. Ras/Raf/MEK/ERK1/2 pathways, PI3K/Akt pathway, JAK/STAT pathway, and PLC- γ pathway have been reported to be the principle pathways activated by ALK signal transduction process. Cell survival is mediated by the JAK/STAT and PI3K/Akt pathways while PLC- γ and Ras/ERK1/2 pathways play a critical role in cell proliferation.⁵⁵ To date, ample number of ALK inhibitors with promising oncolytic activity against ALK⁺-NSCLC tumors have been investigated and developed in the recent decade.

3 ALK inhibitors in NSCLC

ALK is one of the highly competitive drug targets in oncologic drug discovery and development campaigns. ALK-fused genes such as NPM-ALK, EML4-ALK, and KIF5-ALK are putative oncogenes which are accountable for several cancers such as anaplastic large-cell lymphomas (ALCL), inflammatory

myofibroblastic tumors (IMT), diffuse large B cell lymphoma (DLBCL) and a variety of solid tumor types.^{39,56-58} Genetic abnormalities in ALK have been investigated as key drivers of oncogene addicted lung adenocarcinoma and have boosted the researchers to develop novel and targeted therapies. Nevertheless, growing interest to develop synthetic ALK inhibitors has led to a bigger number of research reports related to new drug candidates for cancer therapy. Some of the exciting entities for ALK inhibition approved by FDA with substantial impact in the field of medical oncology are listed in Fig. 3.

Crizotinib (PF2341066, Xalkori), the Pfizer's first-generation dual c-Met/ALK inhibitor⁵⁹ was given approval by FDA in 2011 as first line treatment for ALK⁺-NSCLC.⁶⁰ About 1% of ROS1 rearrangements have been reported in NSCLC patients. FDA in 2016, approved the crizotinib for the management of ROS1-positive metastatic NSCLC.⁶¹ Although crizotinib demonstrated substantial objective response rate (ORR = 74%, $N = 343$, phase III) and remarkable progression-free survival (PFS = 10.9 months). Unfortunately, resistance to crizotinib has been reported as one of the critical issues to date.⁶²⁻⁶⁶ The clinical investigations have revealed that the most common identified secondary mutations are L1196M (gatekeeper mutation), G1202R, L1152R, G1269A, 1151T-ins, C1156Y, S1206Y, and F1174L.⁶⁷⁻⁷² To overcome the limitations exhibited by crizotinib



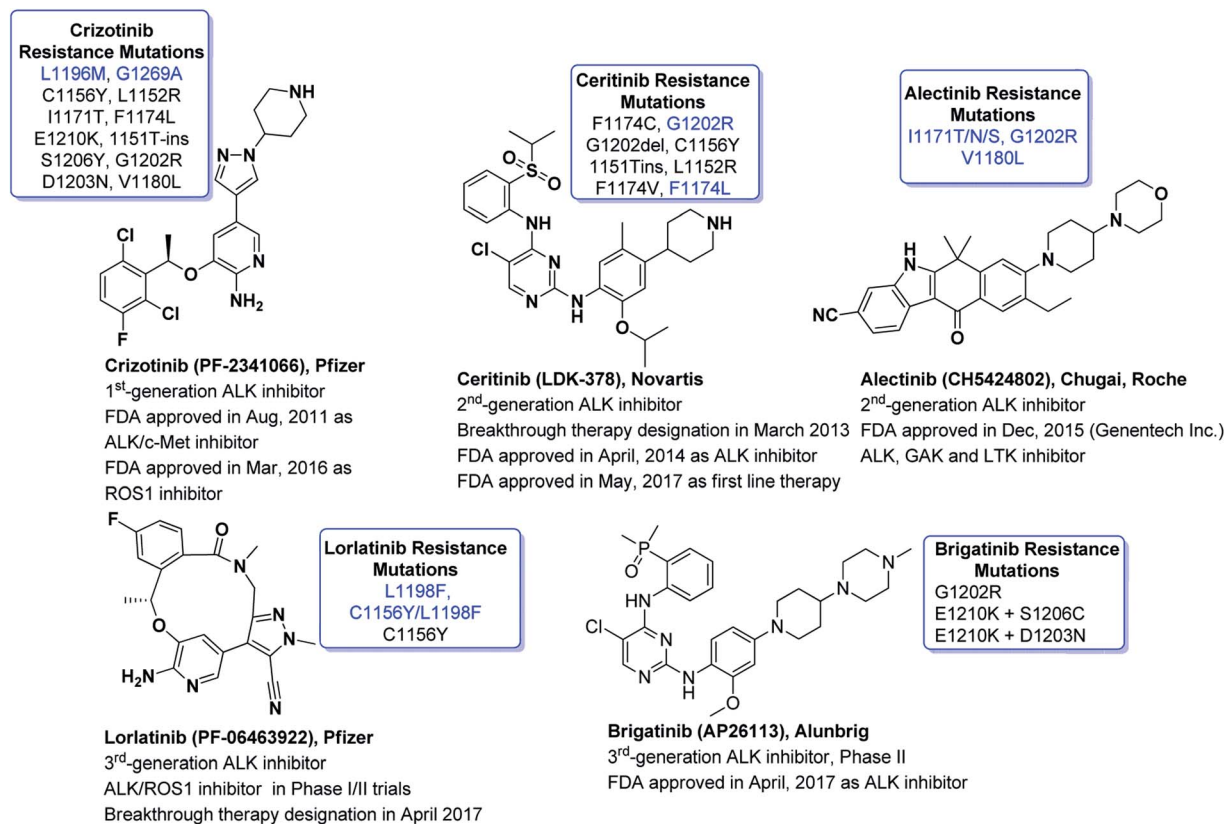


Fig. 3 FDA approved ALK inhibitors with substantial impact in the field of medical oncology with reported point mutations to drug resistance. (Only most critical mutations are shown. The mutations written in blue are most frequent and critical.)

(Xalkori, Pfizer), extensive drug discovery campaigns have led to the innovation of numerous next generation ALK inhibitors to combat disease resistance including alectinib, ceritinib and lorlatinib. It is noteworthy, once crizotinib therapy is failed and then lorlatinib is reported to overcome entire secondary point mutations. Reportedly, point mutations such as L1196M,

G1269A, S1206Y and I1171T can be overcome by ceritinib whereas L1196M, G1269A, F1174L and C1156Y can be overcome by alectinib. Although crizotinib (Xalkori, Pfizer), alectinib (CH5424802, AF-001JP, Alecensa)⁷³ and lorlatinib (PF-06463922, 3rd-generation ALK/ROS1 macrocyclic inhibitor, Pfizer)⁷⁴ are not

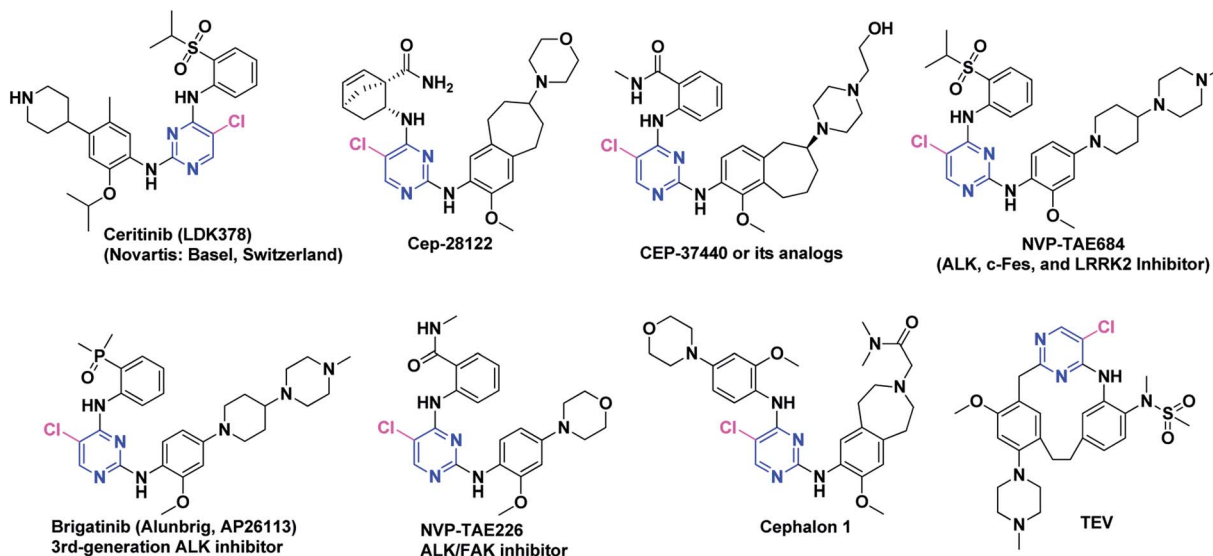


Fig. 4 Novel, potent and selective DAAPalogue as ALK inhibitors.





Table 1 Key features of DAAPalogues as ALK inhibitor advancing to clinical study

	Crizotinib ^a	Ceritinib ^b	Alectinib ^c	Lorlatinib ^d	Brigatinib ^e (Alumbrig)	CEP-28122 ^f	CEP-37440 ^g	NVP-TAE684 ^h	CEP-1
Mol. wt	450	558	483	406	584	539	580	614	596
c log P	4.2	6.5	5.2	2.7	5.9	4.7	4.7	3.9	5.2
TPSA	77	105	72	110	82	—	—	—	—
HBD	3	3	1	2	2	2	2	3	—
Phase	I, II, III	I, II, III	I/II	I/II	I, II, III	Pre-clinical (discontinued)	I	Pre-clinical (discontinued)	(Disc)
FDA approval status	<i>b</i>	<i>c</i>	<i>d</i>	<i>e</i>	<i>f</i>	<i>g</i>	<i>h</i>	<i>i</i>	<i>j</i>
Targets other than ALK, [(IC ₅₀ nM)]	ROS1, c-Met	IGF-1R, InsR, ROS1	RET, LTK, GAK	ROS1	ROS (1.9 nM) IGF1R (24.9 nM), FLT3 (2.1 nM)	FLT4 (IC ₅₀ = 46 nM)	RSK2, SK3, RSK4, FAK	ALK, c-Fes, LRRK2	
ALK ^(enzymatic) (nM)	24	—	—	—	0.37	2.0	3.1	NA	4
IC ₅₀ (nM)	3	0.15	1.9	2	0.62	40	22	3	30
ALK ^(cellular) , (nM)	56	41	2	1.3	5	—	—	16 ⁱ 44 ^k	
EML4-ALK ^{wt}									
IC ₅₀ (nM)	549 (R)	64 (S)	90 (S)	21 (S)	3 (S)	NA	NA	NA	NA
EML4-ALK ^{L1196M}	328	444	207	77	56	ND	ND	ND	ND
EML4-ALK ^{G1202R}	144	164	2	1.6	3	ND	ND	ND	ND
EML4-ALK ^{C1156Y}	81	101	3	0.2	12	ND	ND	ND	ND
EML4-ALK ^{F1174L}	645	2747	169	9.0	2300	ND	ND	ND	ND
EML4-ALK ^{L1152R}	857	668	72	38	142	ND	ND	ND	ND
EML4-ALK ^{L1151Tms}	512	57	9	1.5	2	ND	ND	ND	ND
EML4-ALK ^{G1269A}	65	33	2	4.2	12	ND	ND	ND	ND
EML4-ALK ^{S1206Y}	65, 114 and 115	80, 81, 103, 113 and 115	89 and 115	74, 75, 77, 115 and 116	115 and 117–119	115 and 120	98 and 121–123	115	115
Ref									

Topological polar surface area (TPSA), hydrogen bond donors (HBD), breakthrough therapy designation (BTD), cellular system = (EML4-ALK), discontinued (Disc), the letter S symbolizes mutations which are “sensitive” (clinical and/or preclinical data) to a given drug entity, and “R” symbolizes resistance, data not available (NA), not determined (ND).^a In 2011, FDA granted accelerated approval to crizotinib to treat patients with ALK-positive cancer but traditional approval was given in 2013. However on March 11, 2016, crizotinib got FDA approval as ROS1 inhibitor as well.^b In April 2014, ceritinib obtained accelerated FDA approval for patients with ALK-positive metastatic NSCLC but on May 26, 2017, FDA granted regular approval to ceritinib.^c In June 2013, FDA granted alectinib BTD based on preliminary evidence of clinical activity but got FDA approval in 2014. On Dec 11, 2015, he U.S. FDA granted accelerated approval to alectinib (alectinib) capsules for the treatment of patients with advanced (metastatic) ALK-positive NSCLC.^d In 2015, lorlatinib (Pfizer) received FDA orphan drug status for therapy NSCLC patients. But in April, 2017, BTD was granted by FDA to lorlatinib for therapy of patients with ALK⁺-NSCLC who previously received one or more ALK inhibitors.^e In 2016, FDA granted BTD to brigatinib. In April, 2017, the U.S. FDA granted accelerated approval to brigatinib (ALUNBRIG) for the treatment of patients confirmed with metastatic ALK⁺-NSCLC who progressed on or were intolerant to crizotinib.^f Further optimized to CEP-28122.^g Further optimized to CEP-37440.^h Further optimized to CEP-28122.ⁱ Further optimized to give other CEP analogs (Scheme 5).^j The shown potency was against the two EML4-ALK⁺-NSCLC cancer cell lines such as NCI-H2228 and NCI-H3122.^k The shown potency was against the two EML4-ALK⁺-NSCLC cancer cell lines such as NCI-H2228 and NCI-H3122.

2,4-diarylaminopyrimidines (DAAPalogues) yet they show outstanding potential in clinical settings.

Crizotinib (acyclic) and lorlatinib (macrocyclic) are among the FDA approved ALK inhibitors (Fig. 3) which comprised of similar central core such as aminopyridine. Crizotinib (approved by FDA in 2016) is an ALK/ROS1/c-MET tyrosine kinase inhibitor that has revealed a high activity in ALK⁺-NSCLC. Lorlatinib is a 3rd-generation chemotype that was given breakthrough therapy designation (BTD) by FDA in 2017 and has been used for the treatment of patients with ALK-positive metastatic NSCLC which were treated earlier with one or more ALK inhibitors.^{75–77} We recently documented the strategic development of lorlatinib from crizotinib.⁷⁸ It is noteworthy that both crizotinib and lorlatinib have aminopyridine core and are non-DAAPalogues. Notably, ceritinib (Zykadia®) is the first FDA approved DAAPalogue for ALK/ROS1 inhibition^{79–81} which has been described comprehensively elsewhere.^{82–87} Interestingly, ceritinib being a member of DAAPalogues showed potency to overcome the gatekeeper mutation of L1196M which is still considered a resistance to crizotinib (non-DAAPalogue agent). In crizotinib, the existence of the polar-NH₂ (aromatic amine) substituent at C2-position of pyridine core is considered not to provide favorable interactions with a large lipophilic residue at the gatekeeper position, such as methionine. On the contrary, in ceritinib the chloro (Cl) substituent at C5-position of pyrimidine core could interact more favorably with a methionine gatekeeper and has revealed a remarkable interaction to gatekeeper mutation. On the other hand, alectinib is another FDA approved, highly selective, orally active, potent, and CNS penetrant ALK inhibitor which has shown a promising therapeutic window for the patients treated with crizotinib harboring ALK secondary mutations.^{88–93} Alectinib is non-DAAPalogue drug which comprised of benzo[*b*]carbazolone structural motif^{94–97} that is not ubiquitous template in second or even third generation TKIs. In medicinal chemistry campaign limited structure–activity relationship (SAR) investigation has been reported for benzo[*b*]carbazolone core possibly due to difficulty in chemical synthesis.^{97–101} However, recently reported JH-VIII-157-02 has been potent against the G1202R mutant and comprised of benzo[*b*]carbazolone core.¹⁰² In the designing and development of next-generation ALK-TKIs, the 2,4-diarylaminopyrimidine (DAAP) structural moiety has been incorporated extensively. The literature survey has revealed that majority of the next-generation TKIs being evaluated clinically or currently at preclinical phases have been derived from DAAP scaffold and referred as DAAPalogues. Such ALK-TKIs were developed to potently inhibit all acquired ALK mutants reported previously for first and second generation ALK-TKIs. In recent decade, rapid progress of pharmacological strategies targeting ALK has shown marvelous achievements in clinical settings. Continued research efforts of researchers have made it possible to advance several DAAPalogues to clinical trials. To date, numerous ALK inhibitors have been discovered and diversified chemical classes have been evaluated at pre-clinical or clinical phases and most of them are DAAPalogues.^{60,83,103–107} Some novel, potent and highly selective DAAPalogues for ALK inhibition are shown in Fig. 4.

The DAAP skeleton as a conventional kinase inhibitor core is well recognized and such DAAPalogues has demonstrated high potency against many kinase targets, particularly ALK, c-Met and ROS1. To this end, the drug designing strategy stemmed from our literature survey has demonstrated that ceritinib (LDK-378, 2nd-generation ALK/ROS1 inhibitor Zykadia®, Novartis), brigatinib (Alunbrig™, AP26113), NVP-TAE684, Cephalon 2812, Cephalon 1, and CEP-37440 comprised of DAAP-motif are clinically novel ALK-TKIs.^{87,108–113} It is significant to note that on DAAP template chloro (Cl) substituent have been installed as an optimal substituent at C-5 position of pyrimidine which in most of the DAAPalogues interacts with ALK1196 gatekeeper residue. The designing strategies, biological characteristics, therapeutic exploration, and clinical data of DAAPalogues as ALK inhibitors is well-detailed in literature. In the following sections, we have summarized hit-to-drug evolution strategies, activity spectrum, milestones related to medicinal chemistry discovery efforts, scalable synthetic pathway, and development of some potent ALK-TKIs comprised of DAAP template which are either progressing as investigational or preclinical candidates. Some key features of DAAPalogues as ALK inhibitor advancing to clinical study are shown in Table 1.

4 Brigatinib (Alunbrig™, AP26113)- a second-generation DAAPalogue

Brigatinib (Alunbrig™, AP26113), developed by ARIAD Pharmaceuticals as antineoplastic ALK inhibitor is a 2nd-generation ALK inhibitor with DAAP structural motif. It is a potent DAAPalogue with imperious *in vitro* potential against ALK (IC₅₀ = 0.6 nM L⁻¹), ROS1 (IC₅₀ = 1.9 nM L⁻¹), FLT3 (IC₅₀ = 2.1 nM L⁻¹) and EGFR (IC₅₀ = 1.9 nM L⁻¹). Being a potent DAAPalogue, brigatinib received an accelerated FDA approval for therapy of patients suffering from ALK⁺-NSCLC who either progressed on or were intolerant to crizotinib therapy. The occurrence of resistance to crizotinib posed a therapeutic task and has led to the advancement of 2nd-generation ALK-TKIs such as brigatinib. Brigatinib have exhibited activity against treatment-resistant ALK mutants (Table 1). Brigatinib showed favorable clinical potential and satisfactory safety profile in the patients treated with crizotinib and crizotinib-naïve ALK-rearranged NSCLC patients.^{117,124} The structure–activity relationship reported by Katayama *et al.*, have revealed that brigatinib also demonstrated potency against the triple-mutant EGFR (IC₅₀ < 100 nM). Brigatinib suppresses EGFR phosphorylation and its downstream signaling pathway in cells which express several types of EGFR mutations such as EGFR^{C797S/del19} (IC₅₀ = 28.4 nM), IGF1R (IC₅₀ = 32 nM), and EGFR^{del19} (IC₅₀ = 36.8 nM).^{125,126} This feature has shown that brigatinib will be a promising candidate to overcome the acquired resistance of 3rd-generation EGFR-TKIs if it is used as combination therapy with anti-EGFR antibody. In the following section, description related to hit-to-drug evolution design and the milestones achieved during the development of brigatinib are summarized. These advancements ultimately led the brigatinib to its first global approval for the therapy of patients with ALK⁺-NSCLC who either progressed on or



remained intolerant to crizotinib.¹²⁶ The synthetic strategy of brigatinib (AlunbrigTM) from one of the potent ALK inhibitor (AP23464, IC₅₀ = 0.45 nM) discovered in early discovery efforts is described first.

4.1 Synthetic strategy of brigatinib (AlunbrigTM, AP26113) from (1)

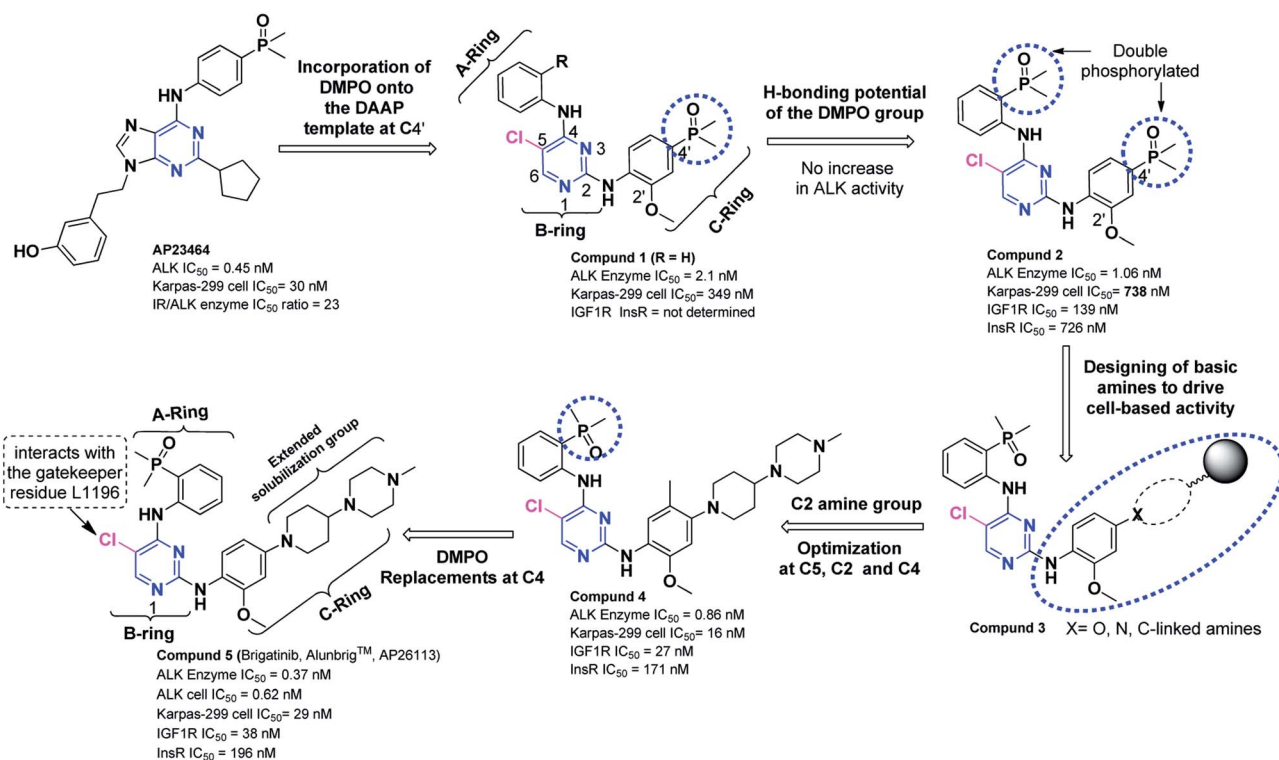
Although phosphorus is an abundant element in human body but rarely employed in drug discovery strategies. Poor cell penetration, biological instability or low oral bioavailability have narrowed down the clinical utilization of mono-phosphonic or bisphosphonic acid-based drugs, phosphonates, phosphinates, and phosphate-containing prodrugs in drug discovery.

Ridaforolimus (mTOR inhibitor), Src-kinase inhibitors (anti-resorptive agents) have been reported in recent decade having phosphine oxide (P=O) functionality. The P=O bond has been investigated as a H-bond acceptor in designing of kinase inhibitors. To this end, analogous efforts related to the discovery of AP23464 (IC₅₀ = 0.45 nM), a potent ATP-based oncogenic protein kinase inhibitor,^{127–129} further stimulated the discovery of potent DAAPalogue (1) where dimethylphosphine oxide (DMPO) onto the 2-anilinopyrimidine template was surveyed.¹¹¹ Aforementioned investigations provoked to incorporate DMPO motif in drug designing which led to the discovery of several DAAPalouges (small-molecules) as ALK inhibitor from a known SRC inhibitor (AP23464) (Scheme 1). The DMPO installation at C-4' enhanced the ALK activity by approximately 7-fold compared to the unsubstituted derivatives in a small focused library closely related to (1). The H-bonding ability of

the DMPO functional group with the Lys-NH (K1150) on the protein was explored near the hinge region by synthesizing double-phosphorylated analog (2) which did not exhibit substantial increase in ALK activity (ALK IC₅₀ = 1.06 nM) but demonstrated remarkably improved selectivity over both IGF1R and InsR (IGF1R IC₅₀ = 139 nM, InsR IC₅₀ = 726 nM).¹¹¹

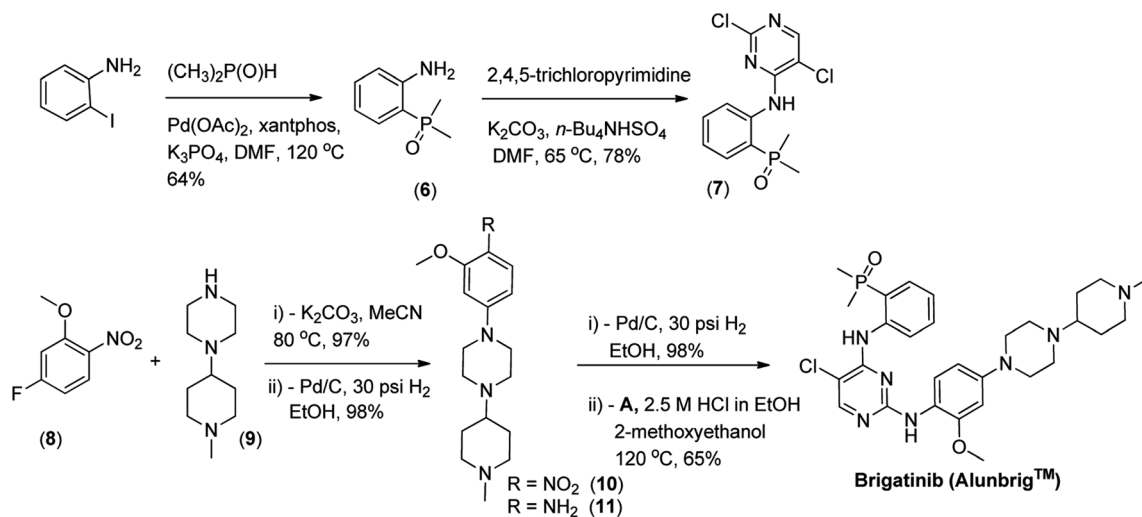
In subsequent investigations, the influence on ALK activity and cellular potency with variety of basic amines containing solubilizing groups as C-ring was surveyed by keeping DMPO motif at *ortho*-position on A-ring. To this end, solubilizing group screening investigation efforts led to the identification of numerous equipotent amines similar to (3). Among them DAAP (5) was most promising analog with promising potency and selectivity profile (ALK IC₅₀ = 0.37 nM, IGF1R IC₅₀ = 24.9 nM, InsR IC₅₀ = 196 nM, Karpas-299 (ALK⁺) = 29 nM, U937 (ALK⁻) = 3194 nM). Subsequent lead optimization explored at other positions on the inhibitor core by maintaining the C-ring resulted in (4) bearing a methyl (Me) group at C-5' position and it was equipotent to (5) in cellular assays but exhibited moderate enzymatic potency loss. 2'-Methoxy (*ortho*-MeO) at C-ring and 5-chloro (Cl) on pyrimidine core was optimal in terms of potency and selectivity. Finally to confirm the critical role DMPO functionality at C-4 of pyrimidine core, a systematic modification of DMPO was carried out with numerous H-bond acceptors (nitrile, amides, *t*-amides, sulfonamides, methylsulfone, isopropylsulfone acetyl *etc.*). The DMPO at C-4 revealed as an crucial structural unit indispensable to attain the appropriate balance between potency and selectivity of DAAPalouge (5).

Final DAAPalouge (5) (brigatinib) emerged as most the advanced clinical candidate loaded with phosphine oxide (P=



Scheme 1 The strategic development of brigatinib (AlunbrigTM, AP26113) (5) from a SRC inhibitor (AP23464).





Scheme 2 Medicinal chemistry discovery pathway of brigatinib.

O) warhead and to date it is being investigated in a global phase II registration trial in patients detected with advanced malignancies and has showed activity in crizotinib-resistant patients. Brigatinib revealed almost 10-fold higher potency and selectivity compared to crizotinib.¹³⁰ Brigatinib has advanced to phase I/II clinical trials which for this unique inhibitor were initiated¹³¹ in September 2011.

Brigatinib, a pyrimidine-based molecule, is a potent and orally bioavailable 2nd-generation DAAPalogue (an ALK inhibitor) which occupies the ATP-binding site of ALK preferably in U-shaped conformation.^{111,132} Brigatinib incorporates many chemical characteristics that enhance its affinity towards ALK inhibition that were lacked in crizotinib. These features include *ortho*-methoxy (OMe) group on C-ring, an extended solubilization group, an optimal chloro (Cl) substituent at C-5 and a distinctive *ortho*-DMPO functionality on the C-4 aniline (Scheme 1). Brigatinib demonstrated potent inhibition with a kinase activity of ALK ($IC_{50} = 0.62 \text{ nmol L}^{-1}$), IGF-1R ($IC_{50} = 38 \text{ nmol L}^{-1}$), insulin receptor kinase ($IC_{50} = 262 \text{ nmol L}^{-1}$) and for other five tested mutant variants (C1156Y $IC_{50} = 0.6$, F1174L $IC_{50} = 1.4$, L1196M $IC_{50} = 1.7$, G1202R $IC_{50} = 4.9$, R1275Q $IC_{50} = 6.6 \text{ nmol L}^{-1}$). Brigatinib also revealed a higher selectivity and only inhibited eleven additional mutants or native kinases (out of 289 screened) with $IC_{50} < 10 \text{ nmol L}^{-1}$. These included ROS1 ($IC_{50} = 1.9 \text{ nmol L}^{-1}$), FLT3 ($IC_{50} = 2.1 \text{ nmol L}^{-1}$), and mutant variants of FLT3 (D835Y) and EGFR (L858R; IC_{50} , 1.5–2.1 nmol L^{-1}). Brigatinib exhibited more modest activity against EGFR with a T790M resistance mutation (L858R/T790M), native EGFR, IGF1R, and INSR with (IC_{50} , 29–160 nmol L^{-1}) but did not inhibit c-MET ($IC_{50} > 1000 \text{ nmol L}^{-1}$). This DAAPalogue has exhibited dual inhibition of ALK and EGFR T790M. Due to this inhibitory mechanism brigatinib could be a promising therapeutic option for patients progressing on crizotinib for the activation of the EGFR bypass track. *In vitro* and in cellular assays against select kinases, the brigatinib has also showed a consistent data for ALK ($IC_{50} = 0.62 \text{ nmol L}^{-1}$ in kinase assay, $IC_{50} = 14 \text{ nmol L}^{-1}$ in EML4-ALK as a cellular system) and ROS1

($IC_{50} = 1.9 \text{ nmol L}^{-1}$ in kinase assay, $IC_{50} = 18 \text{ nmol L}^{-1}$ in CD74-ROS1 cellular system).¹¹⁷

4.2 Medicinal chemistry discovery pathway of brigatinib

Pd-catalyzed phosphorous arylation (P-arylation) methodology was employed to synthesize phosphorus-containing aniline (6) directly from iodoaniline and dimethylphosphine oxide without competitive *N*-arylation though reactive anilino NH_2 substituent was there and such kind of transformations were reported recently.^{133,134} To this end, S_N -Ar reaction of phosphorus-containing aniline (6) with 2,4,5-trichloropyrimidine using base mediated conditions delivered a known intermediate (7) in the following step. In another pathway, S_N -Ar displacement of fluoro (F) group of 2-nitro-5-fluoroanisole (8) with secondary amine (9) using potassium carbonate as basic media yielded nitro-intermediate (10), followed by common hydrogenation provided aniline (11) in high yield. Finally, subsequent substitution at C-2 of (7) was achieved *via* an acid-promoted S_N -Ar reaction in good yield to provide brigatinib. A summarized drug discovery synthetic pathway of brigatinib recently reported by Huang *et al.*, is depicted in Scheme 2.

4.3 Therapeutic trial of brigatinib (Alunbrig™)

4.3.1 Phase I/II trials.

In a preliminary dose-finding study, the patients were given 30–300 mg daily of brigatinib (in the phase I) and in the phase II patient received 90, 180, or 90 mg per day for 7 days followed by 180 mg day^{-1} where 70 patients were crizotinib-resistant and 8 patients were crizotinib-naïve. The most common post-treatment-adverse events were nausea (50%), diarrhea (40%), headache (30%), and cough (30%). Interestingly, in 8% of patients early-onset pulmonary events occurred with dyspnea, pneumonia, and hypoxia, usually within seven days after starting the treatment. This peculiar pulmonary toxicity was considered to drug related and not a class-related event, observed with a higher starting dose of the drug.^{126,135}



The promising clinical activity of brigatinib in the patients pre-treated with crizotinib and crizotinib-naïve ALK rearranged NSCLC has been reported. During the advancement in phase I/II trial, single-arm study, an open label and multicenter study, the brigatinib has revealed significant clinical activity against brain metastases (NCT01449461). In order to establish the recommended dose in phase II trials, dose escalation evaluation in phase I revealed total doses of 30–300 mg per day. Phase II evaluation demonstrated three once daily dosage regimens; 90 mg, 180 mg, and 180 mg with a 7 days lead in at 90 mg. A cohort of 137 patients ($n = 79$, 58% were ALK-rearranged NSCLC) enrolled from academic hospitals or cancer centres in the USA and Spain in phase I/II during the period from September 2011 to July 2014 and all were treated. In phase II clinical study, among the total patients established with ALK-rearranged NSCLC ($n = 79$), 52 patients (66%) showed confirmed objective response. In the patients pre-treated with crizotinib ($n = 71$), a population of patients ($n = 51$) showed an objective response of 72% and objective response of 100% has been reported for crizotinib naïve patients ($n = 8$) (NCT01449461).

In phase I/II clinical findings, brigatinib demonstrated an objective response rate (ORR = 70%, $n = 70$, the patient were on pretested with crizotinib), and median progression free survival (PFS = 13.5, $n = 8$, previously on crizotinib). The one-year overall survival rate (OS = 80%, patients previously on crizotinib) and (OS = 100%, in crizotinib-naïve).¹³⁶

In a phase II single arm trial (ALTA) study, patients ($n = 222$) resistant to crizotinib received brigatinib, the ORR (45%) at 90 mg day⁻¹ and ORR (55%) at 180 mg day⁻¹ was observed in two cohorts. In first dose-finding trial (in ALTA investigation), within the first 7 days pulmonary toxicity occurred in the 6% of patients (3% were grade 3–4) and it was correlated to the higher dose of the drug (180 mg day⁻¹).¹³⁷ In phase I/II trial and ALTA investigations in patients ($n = 32$), the relationship between brigatinib efficacy and ALK mutational status has also been demonstrated.¹³⁸ In the samples obtained at baseline (before treatment with brigatinib), the secondary ALK mutations were detected in the 30% of patients, and the ORR was >60% both in patients with or without secondary ALK mutations which demonstrated the activity of brigatinib regardless of the presence of a secondary ALK mutations conditioning resistance to crizotinib. Although two specific mutations (F1174L and G1202R) have been detected after long-term treatment with brigatinib (in two out of the five post-brigatinib samples), however the responses were also observed when these mutations were present at baseline.¹³⁹

Brigatinib is currently under evaluation in both first-line setting *versus* crizotinib in a phase III trial (NCT02737501) and in phase II study (NCT02706626) after unfruitful outcome of other second-generation ALK inhibitors (ceritinib and alectinib). This last study aims to explore the possibility of brigatinib to overcome the resistance to ALK inhibitors mediated by secondary ALK mutations such as G1202R and I117N/S/T.^{140,141} The clinical activity of brigatinib may further inspire application of phosphorus oxide group as a unique functionality more broadly in the field of medicinal chemistry. In patients with

measurable CNS disease, 53% (8 out of 15) demonstrated an intracranial objective response with an intracranial disease control rate of 87%, and median duration of response was 18.9 months. Therapy emergent side effects were fatigue, nausea, diarrhea, cough and hypertension. ALTA, a phase II trial of brigatinib in patients with ALK⁺-NSCLC who progressed on crizotinib, has completed recruitment of patients (NCT02094573). In this trial preliminary data include 67% patient with CNS disease and 74% with prior treatment who were treated at 180 mg daily. The reported PFS was 12.9 months (95% CI: 11.1–NR), ORR 54% (97.5% CI: 43–65) and CNS response of 67% provides a favorable comparison with previously reported outcomes with ceritinib and alectinib.^{141,142}

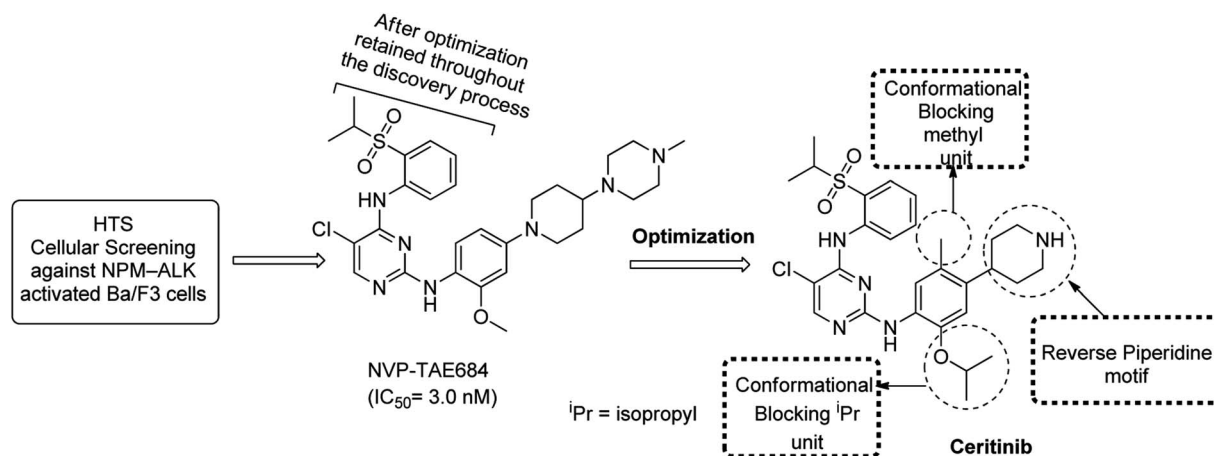
4.3.2 Brigatinib safety profile. In ongoing phase I/II clinical study pertaining to brigatinib, adverse effects (AEs) such as diarrhea (40%), headache (33%), nausea (52%), cough (32%), and fatigue (42%) were most common. The most common grade ≥ 3 AEs were increased lipase (9%), dyspnea (7%), fatigue (4%) and increased amylase (4%). The most common serious AEs were dyspnea (7%) and pneumonia (6%). Seven patients out of 79 (9%) discontinued the study due to AEs. Overall, 9% of patients showed pulmonary events within seven days of therapy commencement. The rate was lower (4%) in patients who started on 90 mg daily (QD) than those who started immediately on the full 180 mg QD dose (14%). No reports of pulmonary events after dose escalation in patients who received 90 mg QD for about 7 days followed by 180 mg QD were observed.^{124,126,141–143}

4.3.3 Ongoing clinical trials of brigatinib. Phase III ALTA-1L trial (NCT02737501) commenced in April 2016 which were designed to draw a comparison between brigatinib and crizotinib in the patients confirmed with advanced NSCLC. This phase III multicenter open-label study of brigatinib is expected to be completed primarily in April 2019. On the other side, phase II study funded by ARIAD started in March 2017 is underway and recruiting patients. This phase II trial is planned to evaluate drug safety and effectiveness in patients with advanced NSCLC who either had first-line cancer treatment but got worse, even after, or while taking 2nd-generation ALK inhibitors. This trial is expected to be completed in June 2020 (NCT02706626).

5 Discovery and clinical development of NVP-TAE 684 DAAPalouge

This DAAPalouge was discovered by Novartis and had 2,4-diaminopyrimidine (DAAP) core decorated with chloro (Cl) at C-5 position and exhibits anticancer chemotherapeutic activity. It was characterized as a highly potent and specific inhibitor of NPM-ALK. The DAAPalouge (NVP-TAE684) is an inhibitor of ALK and exhibited Ba/F₃-NPM-ALK IC₅₀ = 3.0 nM, Ba/F₃-Tel-InsR IC₅₀ = 44 nM, Ba/F₃-WT IC₅₀ = 1336 nM and Karpas-299 IC₅₀ = 2.2 nM. Leucine-rich repeat kinase 2 (LRRK2) is another highly attractive therapeutic target and its association with Parkinson's disease has been reported. NVP-TAE684 is also a highly potent inhibitor of LRRK2 and has demonstrated promising LRRK2 kinase activity (LRRK2^{wt} IC₅₀ = 7.8 nM,





Scheme 3 Summarized layout of NVP-TAE684 DAAPlounge and evolution of ceritinib.

G2019S mutant $IC_{50} = 6.1$ nM). NVP-TAE684 remarkably inhibited Ser935 and Ser910 phosphorylation of both LRRK2^{wt} and G2019S mutant at a concentration ranging from 0.1 to 0.3 μ M in cells, and in mouse spleen and kidney, but not in brain, at an oral doses of 10 mg kg^{-1} .¹⁴⁴ In patients who demonstrated anti-VEGF refractory cancers, c-Fes plays central role in angiogenic cellular responses of endothelial cell and has been reported another potential target. Beside this bone marrow-derived monocytic lineage regulates angiogenesis and c-Fes has been responsible to express in these cells. Investigations pertaining to c-Fes has made TAE-684 a potent inhibitor of c-Fes.^{145–147} NVP-TAE684 induces apoptosis and inhibited growth of diffuse large B-cell lymphoma cells in animal models and induces tumor regression. NVP-TAE684 also induces cell cycle arrest and apoptosis in animal models of anaplastic large-cell lymphoma, preventing lymphomagenesis and inducing regression.

NVP-TAE684 was discovered through high throughput screening (HTS) of a kinase-directed small-molecule library aimed to explore potent entity selectively cytotoxic to BaF₃ NPM-ALK but should not be toxic to non-transformed parental BaF₃ cells.⁵⁶ An inhibition for proliferation of BaF₃ NPM-ALK cells ($IC_{50} = 3$ nM) and for parental BaF₃ cells ($IC_{50} > 1$ μ M) was observed which facilitated the initiation of discovery process of NVP-TAE684. Interestingly, NVP-TAE684 inhibited the human Karpas-299 ALCL cell line ($IC_{50} = 3$ nM) and showed an improvement on crizotinib. Additionally, NVP-TAE684 exhibited from 100–1000 times higher IC_{50} values for non-ALK kinases. The assay was comprised of a panel of thirty five (35) BaF₃ cells which were transformed with several tyrosine kinases constitutively activated by fusion to the TEL oncogene. This specificity was associated as a consequence of the bulky hinge region residue at the 1198 position of the non-ALK kinases (*e.g.* Tyr or Phe compared to Lys existed in ALK).

In another investigation, screening of more than six hundred cancer cell lines was performed for sensitivity to obtain selective ALK inhibitor. As an outcome of the aforesaid findings, NVP-TAE684 was reported to be superior ALK-TKI over Xalkori (crizotinib) in Karpas-299 lymphoma and in the SU-DHL-1 cell

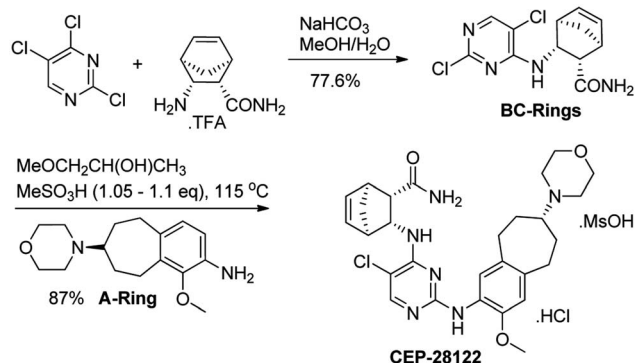
lines.¹⁶ NVP-TAE684 has also emerged as selective for oncogenic ALK containing tumors.¹⁴⁸ The evolution of NVP-TAE684 from the HTS cellular screening is summarized in Scheme 3.

Although the DAAPlounge TAE-684 is a potent ALK inhibitor but later findings revealed that NVP-TAE684 upon metabolic oxidation could produce large number of reactive adducts. Metabolic oxidation was reported to be a potential threat for significant toxicological liabilities. Approximately 20% of NVP-TAE684 was found to be converted into reactive products upon incubation in liver microsomes as determined by semi-quantitative LC-MS analysis. The reactive species were determined using glutathione (GSH).¹⁴⁹ No relationship was established between hepatotoxic drugs and the development of GSH adducts.¹⁵⁰ However, it was hypothesized that reactive metabolites could have a critical role in idiosyncratic¹⁵¹ and/or in other toxicities.¹⁵² To address the further developmental deficiencies of NVP-TAE684 (as a first generation ALK inhibitor), modeling information, designing strategy, and focused investigation to achieve enhanced kinase selectivity of NVP-TAE684 was further surveyed extensively. The installation of iso-propoxy (i-Pr) instead of methoxy (MeO) revealed as well tolerated moiety to address selectivity issues. Further optimization of NVP-TAE684 and potential idiosyncratic toxicity issues led to the eventual discovery of ceritinib which got accelerated FDA approval for the patients with ALK-positive metastatic NSCLC in April 2014. However, on May 26, 2017, FDA declared regular approval to ceritinib for the patients suffering from ALK-positive metastatic NSCLC.¹⁵³

6 Discovery and clinical development of Cephalon DAAPalouges

In the following sections, medicinal chemistry efforts, early discovery description and clinical developments of novel Cephalon DAAPalouges such Cephalon 1, CEP-28122 and CEP-37440 is discussed. Based on reported investigation regarding the involvement of ALK gene rearrangements as an oncogenic driver of ALK-positive cancers, the inhibition of ALK with highly

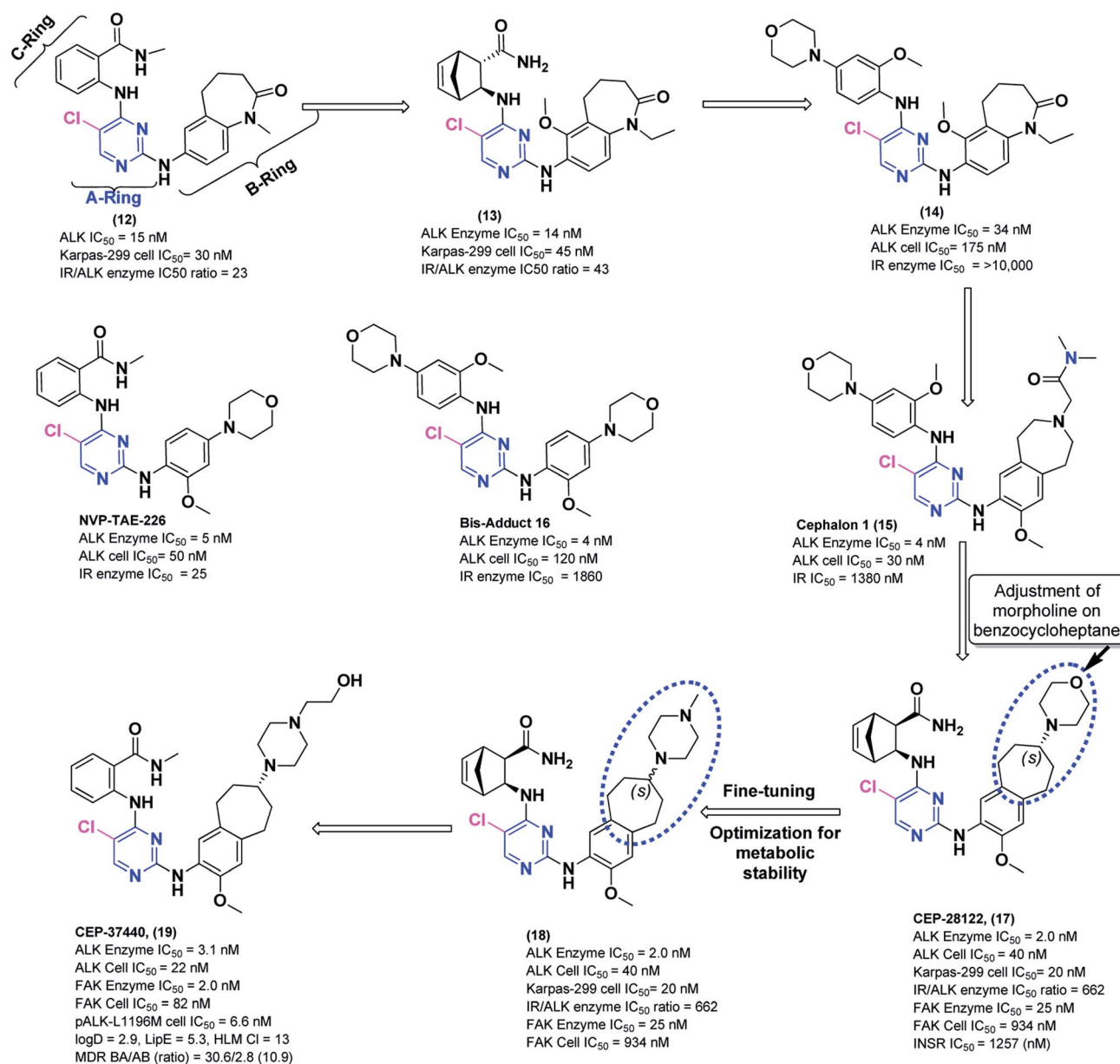




Scheme 4 Pilot plant and scalable synthesis of CEP-28122 (17).

selective small molecular entities has emerged as an attractive area of research and this has led the multiple pharmaceutical companies to discover novel ALK inhibitors. To this end, plethora of unique chemical scaffolds and interesting small-molecules for ALK inhibition were explored. Surprisingly, at that time the DAAPlogues were not well investigated rather tetrahydropyrido[2,3-*b*]pyrazine,¹⁵⁴ indolocarbazole,¹⁵⁵ pyridone,¹⁵⁶ 1*H*-pyrrolo[2,3-*d*]pyrimidine,¹⁵⁷ dianilinopyrimidine,¹⁰⁸ and aminopyridine¹⁵⁸ were among the well probed pharmacophores.¹⁵⁹⁻¹⁶¹

Cephalon 1 (15) (Scheme 5) is a DAAPalogue derivative developed by Cephalon. It is a potent, selective, and orally bioavailable ALK inhibitor but this analog exhibited potential hERG liability. The 2,3,4,5-tetrahydro-1*H*-benzo[*d*]azepine on B-ring motif in Cephalon 1 (15) provided a suitable site in



Scheme 5 The strategic development of CEP-37440 from (12).



subsequent efforts for functional motif exploitation to balance out potency with promising physicochemical properties.¹⁶² This approach resulted in another potent diarylaminopyrimidine ALK inhibitor (DAAPalouge) known as CEP-28122. Interestingly, CEP-28122 was another DAAPalouge developed by Cephalon. The synthesis of CEP-28122 (17) was conducted at the department of chemistry at Cephalon, Inc., and its synthetic routes have been reported elsewhere.¹⁶³ CEP-28122 is a potent, selective, and orally bioavailable ALK inhibitor. CEP-28122 (17) demonstrated ALK kinase activity ($IC_{50} = 1.9$ nM) in an enzyme-based TRF assay. It has shown concentration-dependent growth inhibition/cytotoxicity of ALK⁺-ALCL, NSCLC and neuroblastoma cells. CEP-28122 (17) also exhibited dose-dependent inhibition of ALK tyrosine phosphorylation in tumor xenografts in mice.¹⁶⁴ The strategic development of Cephalon 1 (15) and CEP-28122 (17) is shown (Scheme 5). Medicinal chemistry route as an initial strategy for the synthesis of CEP-28122 (17) exploited the coupling of three key structural motifs such as a morpholine substituted on benzocycloheptane fragment as B-ring, a diaminopyrimidine central core as A-ring, and a bicyclic amino amide portion as C-ring.¹⁶⁵ However, to meet preclinical and clinical supplies, a unique eight-step (longest linear), convergent, and scalable manufacturing route discovered later. The development of scalable new route to CEP-28122 (17) was reported in 2012 which employed several distinctive synthetic approaches such as a selective nitration using *para*-blocking group methodology, a one-pot amination-transfer hydrogenation to provide four reductions approximately in simultaneous fashion, resolution based on an enzymatic reaction, and the leveraging of an *in situ* generated mixed mesylate hydrochloride salt of CEP-28122.¹⁶⁵ The final pilot plant synthetic pathway to CEP-28122 is shown in Scheme 4.

Interestingly, subsequent medicinal chemistry efforts lead to the discovery of CEP-37440 which is another emerging DAAPalouge.¹²¹ CEP-37440 highly kinase selective, potent ATP-competitive, and orally active ALK/FAK inhibitor (ALK_(enzymatic) $IC_{50} = 3.1$ nM, ALK_(cellular) $IC_{50} = 22$ nM) and (FAK1_(enzymatic) $IC_{50} = 2.3$ nM, FAK1_(cellular) $IC_{50} = 82$ nM). CEP-37440 is structurally related to CEP-28122 (an ALK inhibitor). Recently reported investigations related to CEP-37440 has led to enhanced metabolic stability, imperious pharmacokinetic features, and adequate *in vitro* activity in clinically described resistance mutations.¹¹² In the following section, medicinal chemistry efforts related to early discovery of CEP-37440 (12) and clinical developments are discussed.

6.1 Synthetic strategy of emerging DAAPalouge CEP-37440 from 12

In a quest to discover potent ALK inhibitors, 7-amino-1,3,4,5-tetrahydrobenzo-[*b*]azepin-2-one (12) (Scheme 5) was identified as a novel DAAPalouge (a small molecule) which exhibited potent activity (ALK $IC_{50} = 15$ nM, IR $IC_{50} = 23$ nM, Karpas-299 cell $IC_{50} = 30$ nM) but the margin against IR was merely 1.5-fold. Benzamide functional group was a key potency determinant for ALK in (12). Due to the involvement of insulin receptor (IR) in the regulation of glucose transport, glycogen, and fat

biosynthesis, selectivity towards IR were of particular interest. Based on promising SAR, potent enzymatic/cellular ALK inhibition, and modulation of IR activity, improvement in modest oral bioavailability and optimization of IR selectivity was surveyed on azepinone core (B-ring) (Scheme 5). Ethyl substitution on nitrogen atom of azepinone for improved pharmacokinetic properties (PK) parameters was envisioned as essential group. For reasonable selectivity, *ortho*-MeO group either at 6-position or 8-position of the benzazepinone was proposed to be superior. Furthermore, focused modification on A- and C-rings, a bicyclo[2.2.1]hept-5-ene ring system was identified as an alternative C-ring partner for improved IR selectivity. Further optimization of potency, off-target selectivity, and PK parameters led to the discovery of a novel and potent bridged bicyclic derivative (13) (Scheme 5). Gratifyingly, (13) demonstrated impressive activity in an ALK assay (enzymatic or cellular) measuring oncogenic NPM-ALK autophosphorylation (ALK $IC_{50} = 14$ nM, IR $IC_{50} = 597$ nM, Karpas-299 cell $IC_{50} = 45$ nM) with modestly improved IR selectivity (IR/ALK = 43). Moreover, oral administration of lead inhibitor (13) to mice induced with ALK positive ALCL SCID xenografts resulted in near-complete/complete regressions.^{166,167} Although (13) presented a balanced profile of ALK potency and exhibited global kinase selectivity but further efforts were needed to improve IR selectivity (enzymatic IR/ALK IC_{50} ratio of >100). Despite of the promising features, (13) still lacked the capability of dose escalation in preclinical toxicological models which precluded its further improvement.

Cephalon researcher during the course of their endeavors synthesized and profiled a diversity of analogs closely related to NVP-TAE226 (ref. 168) but bis-adduct (16) (Scheme 5) was discovered accidentally.¹⁶⁹ This bis-adduct (16) demonstrated ALK inhibition equivalent to NVP-TAE226 and tremendous escalation in IR selectivity but lower (2.4-fold) cellular inhibition. The serendipitous findings were capitalized and a morpholine-phenyl moiety was incorporated as C-ring fragment which led to emergence of novel compound (14) with benzazepinone as B-ring. Compound (14) exhibited drop-off in both ALK enzymatic activity and cellular potency was observed but improvement in potency while maintaining this high degree of IR selectivity was sought. Rather than benzazepinone moiety, a structural motif such as 2,3,4,5-tetrahydro-1*H*-benzo[*d*]azepine was perceived to be a synthetically tractable functionality with potential handles to modulate both potency and physicochemical properties and this was incorporated into the pharmacophore. Extensive SAR investigation and substitutions at benzazepine nitrogen of (15) and subtle variations to the nature of groups impacted on both enzyme and cell activity prominently. Numerous analogs were designed wherein basicity of benzazepine nitrogen emerged to be critical contributing factor. Gratifyingly, novel amide (15) was identified as orally bioavailable, most potent and highly selective ALK inhibitor (ALK enzyme $IC_{50} = 4$ nM, ALK cell $IC_{50} = 30$ nM, IR enzyme $IC_{50} = 1380$ nM, IR/ALK IC_{50} ratio of 345) and displayed promising *in vivo* efficacy in ALCL tumor models.¹⁶⁹ Candidate (15) demonstrated tumor regressions at 30 mg kg⁻¹ and 55 mg kg⁻¹ b.i.d., po doses, at which (15) was found to be tolerated fairly. Neither





Table 2 Comparison of PK and key features of intermediate DAAPalouges in discovery of CEP-37440 (19)

Comp.	ALK enzyme IC ₅₀ (nM)	ALK cell IC ₅₀ (nM)	INSR enzyme IC ₅₀ (nM)	INSR cell IC ₅₀ (nM)	IR/ALK IC ₅₀	FAK enzyme IC ₅₀ (nM)	FAK cell IC ₅₀ (nM)	Karpas-299 cell IC ₅₀ (nM)	hERG IC ₅₀ (µM)	CYP IC ₅₀ (µM)	Liver microsome						F (%) rat	Oral AUC _{0-∞} (ng h mL ⁻¹)			
											LM (t _{1/2} , min)										
											M	R	Mo	H	H	H					
12	15				23			30													
13	14				43			45													
14	34	175	>10 000																		
16	4		1860																		
15	4	30	1380		345			190											30	1019	
17	1.9	42	662	>10 000	662	30	944	20	>10	>10		8	16	17	11	11	2.5			1203	
18	9	200	508			6.5			7.9	>10	>10	40	39	7	11						5526
19	3.1	22	65	2000		2.0	80		>10	>5.6		40	40	21	40	42					8360

evident toxicity nor body weight loss were observed. Although compound (15) exhibited potent *in vitro* ALK activity but potential hERG liability was main concern (IC₅₀ = 3.9 µM for hERG, patch clamp assay) and against the most important CYP isoforms (15) demonstrated minimal inhibition for CYP1A2 and 2D6.¹⁷⁰

In subsequent investigations, the basic nitrogen atom was shifted outside the 7-membered ring in (17) to further control the potency and selectivity variances that were associated with the analogs of azepine (15). The new B-ring was amino-benzocycloheptene motif with (*S*)-configuration at the morpholino carbon juncture. To generate potent ALK inhibition, enhanced IR selectivity, and dose proportional escalation in preclinical toxicology models, a dedicated medicinal chemistry efforts were therefore initiated to evaluate the amendments on aminobenzocycloheptene ring system. With this strategy Cephalon researchers identified (17) wherein at C-2 position of azepine or azepinone fragments were replaced with amino-benzocycloheptene motif as B-ring. Furthermore, earlier established bicyclo[2.2.1]hept-5-ene ring moiety was incorporated at C-4 position of pyrimidine as C-ring to afford the fully assembled HPLC separable (*S*)-enantiomer DAAPalouge scaffold. Compound (17) revealed as exceedingly potent and highly selective DAAPalouge (an ALK inhibitor). The PK of compound (17) was investigated in numerous species as shown in Table 2.

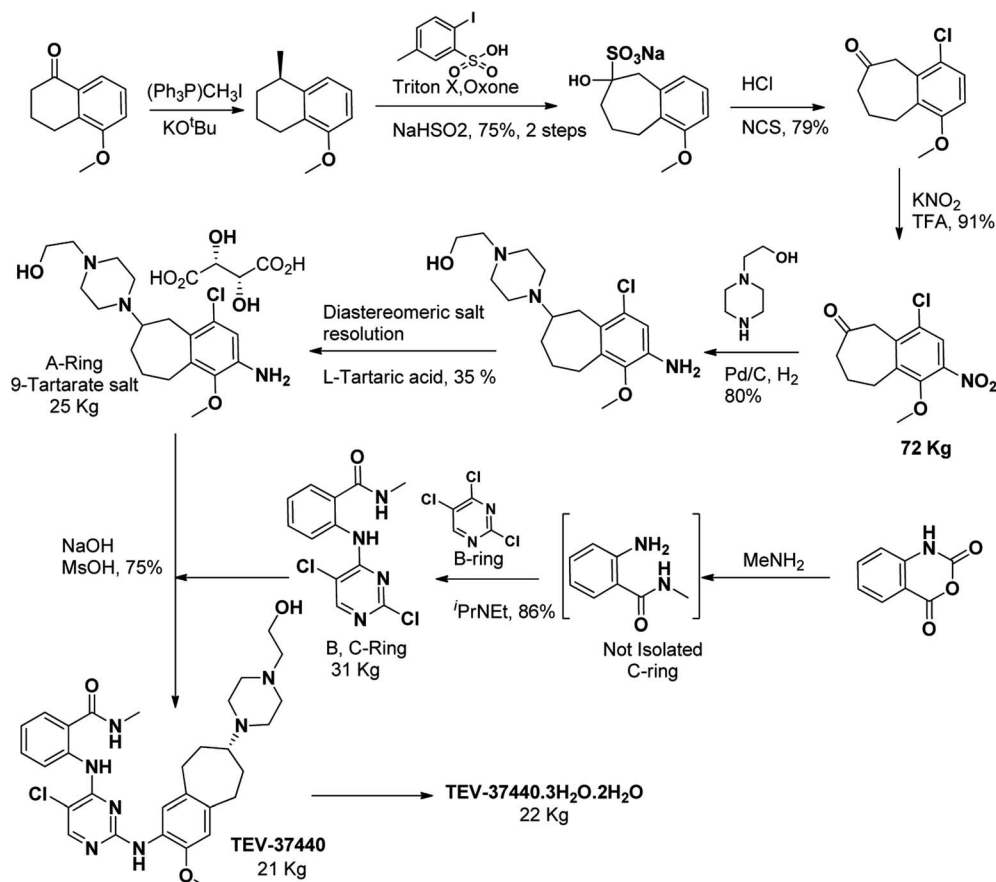
6.2 Pilot plant synthesis of CEP-37440 DAAPalouge

To provide early-phase clinical supplies of CEP-37440, Allwein *et al.*, reviewed the previously described medicinal chemistry route of CEP-28122 and presented a scalable route to TEV-CEP-37440 (ref. 171) to obtain 22 kilograms of (19) as trihydrochloride dihydrate. The pilot plant synthesis involve the use of several steps such as a unique ring expansion approach, *para*-blocking group strategy followed by selective nitration, amination-hydrogenation in a single-pot, resolution of diastereomeric salt, a step to avoid a dangerous intermediates, and a large-scale production of trihydrochloride dihydrate salt were the key highlighted features. The optimized pilot plant synthesis provided TEV-37440 as trihydrochloride dihydrate in >99.5% purity and in 10.7% overall yield (Scheme 6).¹⁷²

6.3 Therapeutic trial of CEP-37440 (19)

An oral dose escalation investigation for (17) was conducted in rats using variable concentrations of 10, 30, 55, and 100 mg kg⁻¹. Compound (17) showed a proportional increase in both AUC (~15-fold increase) and C_{max} (~9-fold increase) at 10 and 100 mg kg⁻¹ doses. Interestingly, using dose escalation from 10 to 100 mg kg⁻¹ (~10-fold increment), previously discovered benzazepinone (13) displayed C_{max} (only ~2-fold increase) and AUC (only ~2-fold increase). To this end, based on dose escalation comparative data, an enhanced PK exposure was noticed for (17) and further efforts for preclinical advancement of (13) were terminated in favor of (17).

A single dose of novel CEP-28122 (17) given orally at 30 mg kg⁻¹ revealed more than 90% inhibition of NPM-ALK phosphorylation in the tested NPM-ALK positive ALCL Karpas-299



Scheme 6 Pilot plant synthesis of CEP-37440 DAApalouge to provide clinical supplies.¹⁷¹

tumor xenograft in SCID mouse. Antitumor efficacy of (17) was evaluated in a Karpas-299 tumor xenograft model in SCID mouse and dose-dependent antitumor activity was measured at both oral doses of 10 mg kg^{-1} and 30 mg kg^{-1} , b.i.d. CEP-28122 (17) demonstrated complete tumor regressions at the 30 mg kg^{-1} oral b.i.d. dose. Administration of CEP-28122 (17) was well tolerated and at both dosing regimens neither evident toxicity nor a significant body weight loss was observed.¹¹²

Dose-dependent antitumor activity potential was also assessed in NPM-ALK-positive ALCL and EML4-ALK-positive NSCLC tumor xenografts in mice wherein CEP-28122 (17) was administered orally. It showed complete or near to complete tumor regression by following a therapy at a dose of 30 mg kg^{-1} b.i.d. or higher. Furthermore, CEP-28122 (17) induced growth inhibition and cytotoxicity of neuroblastoma cell lines harboring ALK activating mutants (F1174L in NB-1643 cells and R1275Q in SH-SY5Y cells, respectively). However, CEP-28122 (17) did not induce growth inhibition and cytotoxicity in those which contained ALK wild type (WT) without gene amplification (SKNAS cells). This indicated that (17) was active against the two ALK activating mutants usually found in human neuroblastoma. From the combination of entire favorable biological and pharmaceutical properties, the CEP-28122 (17) was discovered as a preclinical candidate.¹¹² The PK characteristics of novel

analogs (13–19) discovered during the strategic development of CEP-37440 from (12) are presented in Table 3.

Since the discovery of CEP-28122 DAApalouge (17), the landscape of small molecule inhibitor had changed to broader extent and four distinct molecular entities had been approved by FDA to treat EML4-ALK positive NSCLC. Although (17) demonstrated overall moderate *in vivo* clearance potential [CD-1 mouse, $\text{Cl} = 29 \text{ mL min}^{-1} \text{ kg}^{-1}$; rat, $\text{Cl} = 17 \text{ mL min}^{-1} \text{ kg}^{-1}$; monkey, $\text{Cl} = 28 \text{ mL min}^{-1} \text{ kg}^{-1}$], and the *in vitro* liver microsomal stability values [liver microsome $t_{1/2}$ (min): CD-1 mouse, 13; rat, 7; human, 10] which prompted the further possibility for improvement. Innovative chemical architecture which could constrain the small-molecule ligands into the putative active conformation was executed which resulted in generation of architecturally distinctive template to overcome associated pharmaceutical liabilities. Recently focal adhesion kinase (FAK) has been investigated as a cytoplasmic protein tyrosine kinase and its overexpression and activation has been characterized in several advanced-stage solid cancers. FAK activation and overexpression are suggestive of numerous solid tumors, mainly those have susceptibility for bone metastasis especially NSCLC, ovarian, breast, prostate, and HNSCC carcinomas.^{173–175} Due to the growing interest in focal adhesion kinase (FAK) inhibitors and their clinical development,¹⁷⁶ extensive SAR was sought on (17) which resulted in superior kinase selectivity profile,



Table 3 Pharmacokinetic parameters of novel analogs (13–19) in CD-1 mouse, Sprague-Dawley (SD) rats, and cynomolgus monkeys

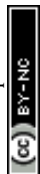
PK parameters	Compound																			
	(13)					(15)					CEP-28122 (17)					CEP-37440 (19)				
	SCID mouse	Rat	Dog	S-D rat	CD-1 mouse	CD-1 mouse	Rat	Dog	Mo	CD-1 mouse	SD rat	Mo	CD-1 mouse	SD rat	Mo	CD-1 mouse	SD rat	Mo		
iv																				
Dose (mg kg ⁻¹)	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	
t _{1/2} (h)	0.3	0.8	1.1	0.80 ± 0.05	0.7	0.7	0.9 ± 0.2	2.4 ± 0.2	1.2 ± 0.05	0.9 ± 0.2	0.9 ± 0.2	2.4 ± 0.2	1.2 ± 0.05	3.0	2 ± 0.4	5.4 ± 0.6	2 ± 0.4	5.4 ± 0.6	30.0 ± 0.5	
Cl (mL min ⁻¹ kg ⁻¹)	37	13	22	16 ± 1.5	20	29	17 ± 2	40 ± 1	28 ± 4	17 ± 2	17 ± 2	40 ± 1	28 ± 4	10	4 ± 0.2	30.0 ± 0.5	4 ± 0.2	30.0 ± 0.5	—	
% Cl/hepatic blood flow	41	20	58	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	
V _d (L kg ⁻¹)	0.9	0.9	2.1	1.1 ± 0.1	1.2	1.8	1.3 ± 0.2	8.2 ± 0.5	2.9 ± 0.3	1.3 ± 0.2	1.3 ± 0.2	8.2 ± 0.5	2.9 ± 0.3	2.7	0.8 ± 0.2	13.2 ± 1.9	0.8 ± 0.2	13.2 ± 1.9	—	
AUC _{0-∞} (ng h mL ⁻¹)	446	1276	773	1019 ± 91	830	584	989 ± 127	409 ± 16	615 ± 89	989 ± 127	989 ± 127	409 ± 16	615 ± 89	1612	4005 ± 237	554 ± 11	4005 ± 237	554 ± 11	—	
Po																				
Dose (mg kg ⁻¹)	10.0	5.0	10.0	5.0	10.0	10.0	10.0	10.0	10.0	10.0	10.0	10.0	10.0	10.0	10.0	10.0	10.0	10.0	10.0	
C _{max} (ng mL ⁻¹)	0.25	0.8	1.1	3.06 ± 53	1192	880	599 ± 3	427	170 ± 51	599 ± 3	599 ± 3	427	170 ± 51	1533	1340 ± 107	239 ± 6	1340 ± 107	239 ± 6	—	
t _{max} (h)	2.4	2.2	1.6	n.d	3.1	0.25	1.3 ± 0.3	0.8	4.7 ± 1.3	1.3 ± 0.3	1.3 ± 0.3	0.8	4.7 ± 1.3	2	3.3 ± 0.7	6	3.3 ± 0.7	6	—	
t _{1/2} (h)	1406	4088	2482	1395 ± 281	3895	2993	6.6 ± 1.6	3.3	1.2 ± 0.05	6.6 ± 1.6	6.6 ± 1.6	3.3	1.2 ± 0.05	—	—	—	—	—	—	
AUC _{0-∞} (ng h mL ⁻¹)	32	64	31	30 ± 6	64	51	4833 ± 797	2126	954 ± 209	4833 ± 797	4833 ± 797	2126	954 ± 209	16 429	8360 ± 540	2757 ± 114	8360 ± 540	2757 ± 114	—	
F (%)	—	—	—	—	—	—	50 ± 8	52	16 ± 3	50 ± 8	50 ± 8	52	16 ± 3	102	42 ± 3	50 ± 3	42 ± 3	50 ± 3	—	

favorable brain penetration, and promising activity in ALK⁺-NSCLC tumor xenografts. Compound (18) also exhibited imperious biochemical and pharmacodynamic activity both *in vitro* and *in vivo*, and enhanced antitumor effectiveness in FAK and FAK-dependent tumors.¹¹²

In further exploration, nature of substituents (morpholine, *N*-methyl piperazine and methoxyethylamine) on benzocycloheptane ring system and position refinement of morpholine itself was surveyed. Compared to (19) (ALK IC₅₀ = 3.1 nM, cell IC₅₀ = 22 nM), both diastereomers of (18) were almost 4-fold lesser potent in enzymatic assays, and 10-fold lesser potent in cells based assays but maintained plausible hERG selectivity (7.9–9.1 μM) and CYP inhibition (>10 μM). Furthermore, an enhancement in liver microsomal stability in mouse and rat was observed but human and monkey liver microsomal stability was lesser than anticipated. Pharmacokinetics of (18) in rat demonstrated high iv AUCs but lower Cl and bioavailability (*F* = 11%). Encouraging results compelled to improve ALK potency by maintaining the additional FAK effectiveness (potency) and human liver microsomal metabolic stability as next envisioned agenda. Subsequent investigation led to the optimization of substituent at piperazine and C-2 position of pyrimidine. Previously reported bicyclo[2.2.1]heptene motif at C-4 of pyrimidine was main metabolic hotspot in (17). To this end, to expedite discovery efforts the racemic mixtures of aromatic sulfones, sulfonamides, *N*-alkylsulfonamides, imidazoles, pyrazoles, and cyclic sultams were surveyed as novel H-bond acceptors on C-4 position of pyrimidine which resulted in *N*-methylbenzamide as optimal C-ring partner. On the other hand, variation of *N*-methyl substituents on the piperazine ring to hydroxyethyl functionality was identified as a noteworthy group on B-ring.

Racemic mixture of (19) revealed as a novel lead candidate with promising ALK enzymatic activity (IC₅₀ = 4 nM), cellular potency (IC₅₀ = 40 nM), and acceptable dual ALK and FAK kinome selectivity. Candidate (19) proved adequate *in vitro* liver microsomal stability (rat and human t_{1/2} > 40 min) across the tested species, adequate oral bioavailability (47%), and issues of hERG activity (IC₅₀ ≥ 10 μM) were also circumvented relative other analogs. Furthermore, (*R*) and (*S*)-enantiomers of B-ring were installed and individual enantiomers of (19) were synthesized and profiled. Findings revealed both of the enantiomers fairly comparable across off-target selectivity, on-target potency, and *in vitro* ADME/toxicity parameters but in spite of comparable *in vitro* metabolic stability of both enantiomers, (*S*)-enantiomer (19) showed 2-3-fold enhancement in oral bioavailability and considerable higher area under the curve (AUC). The structural basis for this observed potency was recognized using molecular docking. For clinically-derived resistance mutations against small-molecule, the ALK inhibitors compound (19) was tested for ALK mutants and showed equipotent potencies (ALK_(L1151Tins) K_d = 2.6 nM, ALK_(C1156Y) K_d = 1.6 nM, ALK_(F1174L) K_d = 1.1 nM, and ALK_(L1196M) K_d = 3.3 nM) to that of FDA approved first and 2nd-generation ALK inhibitors.¹¹²

Compound (19) exhibited promising *in vitro* ADME properties and appropriate oral bioavailability in three investigated



species. CEP-37440 (19) showed promising brain exposure which may influence physiologically related escape pathways, for example brain metastases. Moreover, dose-dependent anti-tumor efficacy has also been reported in several animal models of anaplastic lymphoma kinase- positive and FAK-driven cancers. CEP-37440 (19) being a dual FAK and ALK active DAAPalouge is of distinctive interest in terms of the recent observations in inflammatory breast carcinoma where the subsets are comprised of ALK or FAK oncogenic units. CEP-37440 (19) is currently undergoing human clinical trials. The effect of CEP-37440 (19) using human IBC cells and preclinical models of IBC has demonstrated that CEP-37440 inhibits the proliferation of some IBC cells by reducing the level of phospho-FAK1 (Tyr 397) but none of the cells showed expression of ALK. The investigations using IBC xenografts models revealed that compound (19) also efficiently reduces the growth of the primary tumor xenografts and inhibits the development of brain metastases in mice.¹⁷⁷ In preclinical profiling, CEP-37440 (19) has demonstrated advantageous metabolic stability, pharmacokinetic and proven the CNS brain penetration.¹²¹

7 Conclusion and future prospectives

The trend and rapidity pertaining to drug discovery of anti-cancer agents has substantially changed the understanding of pathogenesis and cancer therapy. The extensive inclusion of computer-aided approaches has proven important involvement and has eliminated the requirement of expensive and time-consuming tasks frequently used in experimental drug discovery processes.^{178–180} To this end, widespread use of the techniques such as HTS, virtual screening (VS), pharmacophore modeling, and three-dimensional quantitative structure–activity relationships (3D QSAR) have made the lead compound discovery process much faster. Although the HTS is costly technique yet it has shown remarkable utilization in drug discovery campaigns since 1980. However, VS approach applied in the private sector and academia is a cost-saving technology, and has emerged as a versatile tool in the hit to lead identification and eventually becoming an integral part of drug discovery war, whereas the pharmacophore modeling has also been effectively used in recent years to identify a common pharmacophore hypothesis. In addition, the 3D QSAR model unfolds the statistical correlation between structure of designed analog and predictive functions. The 3D QSAR models are also playing significant role to accomplish greater success rate of experimentation, cost reduction of development, and short cycle of research in drug discovery.^{181–183}

Due to inclusion of above said technologies in drug discovery campaigns, the current decade (2007–2017) has dragged prodigious progress globally in NSCLC pertaining to drug development and in clinical settings.^{183–187} Novel targeted anti-neoplastic agents loaded with anticancer warhead to demonstrate imperious efficacy have been synthesized and introduced for clinical use. Second generation DAAPalouges including alectinib are known to be effective in greater than 50% NSCLC patients who were refractory to crizotinib. Unfortunately, the emergence of resistance to 2nd-generation ALK-TKIs is a major

concern to date. Among the identified multiple resistance mechanisms, the G1202R mutation of ALK is emerging as resistance to all currently available ALK-TKIs including DAA-Palouges. Lorlatinib (non-DAAPalouge) is a first macrocyclic 3rd-generation ALK-TKI which have demonstrated effectiveness against the G1202R mutation of ALK but again compound mutation such as C1156Y/L1198F in the ALK kinase domain has been revealed as a major concern. Brigatinib is a DAAPalouge which has shown the potential to overcome the resistance mechanism to first and second generation ALK inhibitors including the G1202R mutation of ALK in xenograft models.¹¹⁷ However, many other newly discovered molecular entities have demonstrated effectiveness against the G1202R mutation.^{188–190} Furthermore, the current median overall survival of patients with ALK-rearranged lung cancer is above 4 years only. This over survival is achieved through the treatment protocols of using sequential ALK-TKIs in addition to other FDA approved chemotherapeutics. Finally, despite the discovery the development of several potent ALK TKIs including DAAPalouges have led to the substantial increase of drugs but still it is believed that currently available therapies are palliative rather curative. Thus further discoveries are indispensable to develop superior therapeutic strategies.

Conflicts of interest

The authors also declare no conflicts of interest.

Abbreviations

ADME	Absorption, distribution, metabolism, and excretion
ALCL	Anaplastic large cell lymphoma
ALK	Anaplastic lymphoma kinase
ATP	Adenosine triphosphate
BTD	Breakthrough therapy designation
c-MET	Mesenchymal to epithelial transition
CNS	Central nervous system
DAAP	2,4-Diarylaminopyrimidine
DAAPalouges	2,4-Diarylaminopyrimidine analogues
DLBCL	Diffuse large B-cell lymphomas
DMPO	Dimethylphosphine oxide
EGFR	Endothelial growth factor receptor
EML4	Echinoderm microtubule-associated protein-like 4 gene
FDA	Food and Drug Administration
GSH	Glutathione
HBD	Hydrogen bond donors
HGF/SF	Hepatocyte growth factor/scatter factor
HTS	Throughput screening
IGF1R	Insulin-like growth factor 1 receptor
IMT	Inflammatory myofibroblastic tumours
INSR	Insulin receptor
IR	Insulin receptor
JAK	Janus activated kinase
LRRK2	Leucine-rich repeat kinase 2
LTK	Leukocyte tyrosine kinase



MTT	Molecular targeted therapy
MW	Molecular weight
S _N -Ar	Nucleophilic aromatic substitution
NPM	Nucleophosmin
NSCLC	Non-small-cell lung cancer
ORR	Objective response rate
P-arylation	Phosphorous arylation
PFS	Progression-free survival
PI3K	Phosphatidylinositol 3-kinase
PK	Pharmacokinetic properties
PLC	Phospholipase C
PTN	Pleotrophin
pY	Phosphotyrosine
ROS1	ROS Proto-Oncogene 1
SAR	Structure–activity relationship
SCC	Squamous cell carcinoma
SCLC	Small-cell-lung cancer
TKI	Tyrosine kinase inhibitor
TPSA	Topological polar surface area
VEGFR	Vascular endothelial growth factor receptor
VS	Virtual screening
3D-QSAR	Three-dimensional quantitative structure–activity relationships

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References

- 1 S. Rebecca, N. Deepa and J. Ahmedin, *Ca-Cancer J. Clin.*, 2013, **63**, 11–30.
- 2 R. Siegel, J. Ma, Z. Zou and A. Jemal, *Ca-Cancer J. Clin.*, 2014, **64**, 9–29.
- 3 R. L. Siegel, K. D. Miller and A. Jemal, *Ca-Cancer J. Clin.*, 2015, **65**, 5–29.
- 4 R. L. Siegel, K. D. Miller and A. Jemal, *Ca-Cancer J. Clin.*, 2016, **66**, 7–30.
- 5 R. L. Siegel, K. D. Miller and A. Jemal, *Ca-Cancer J. Clin.*, 2017, **67**, 7–30.
- 6 A. D. Lopez, N. E. Collishaw and T. Piha, *Tob. Control*, 1994, **3**, 242.
- 7 L. Collins, C. Haines, R. Perkel and R. Enck, *Am. Fam. Physician*, 2007, **75**, 56–63.
- 8 P. National Center for Chronic Disease and S. Health Promotion Office on and Health, in *The Health Consequences of Smoking-50 Years of Progress: A Report of the Surgeon General*, Centers for Disease Control and Prevention (US), Atlanta (GA), 2014.
- 9 R. Courtney, *Drug Alcohol Rev.*, 2015, **34**, 694–695.
- 10 S. McGuire, *Adv. Nutr.*, 2016, **7**, 418–419.
- 11 J. Subramanian and R. Govindan, *J. Clin. Oncol.*, 2007, **25**, 561–570.
- 12 W. Travis, *Clin. Chest Med.*, 2011, **32**, 669–692.
- 13 M. Latif, A. Saeed and S. Kim, *Arch. Pharmacol Res.*, 2013, **36**, 1051–1054.
- 14 R. Clay, F. Maldonado and E. S. Edell, *Interventions in Pulmonary Medicine*, ed. J. P. Díaz-Jimenez and A. N. Rodriguez, Springer International Publishing, Cham, 2nd edn, 2018, pp. 313–322, DOI: 10.1007/978-3-319-58036-4_19.
- 15 G. Scagliotti, *Ann. Oncol.*, 2007, **18**(suppl. 10), 41.
- 16 U. McDermott, A. Iafrate, N. Gray, T. Shioda, M. Classon, S. Maheswaran, W. Zhou, H. Choi, S. Smith, L. Dowell, L. Ulkus, G. Kuhlmann, P. Greninger, J. Christensen, D. Haber and J. Settleman, *Cancer Res.*, 2008, **68**, 3389–3395.
- 17 P. Ong and D. Ost, *Interventions in Pulmonary Medicine*, ed. J. P. Díaz-Jimenez and A. N. Rodriguez, Springer International Publishing, Cham, 2nd edn, 2018, pp. 323–332, DOI: 10.1007/978-3-319-58036-4_20.
- 18 S. O. Dolly, D. C. Collins, R. Sundar, S. Popat and T. A. Yap, *Drugs*, 2017, **77**, 813–827.
- 19 S. W. Morris, M. N. Kirstein, M. B. Valentine, K. G. Dittmer, D. N. Shapiro, D. L. Saltman and A. T. Look, *Science*, 1994, **263**, 1281.
- 20 M. Shiota, J. Fujimoto, T. Semba, H. Satoh, T. Yamamoto and S. Mori, *Oncogene*, 1994, **9**, 1567–1574.
- 21 S. W. Morris, M. N. Kirstein, M. B. Valentine, K. Dittmer, D. N. Shapiro, A. T. Look and D. L. Saltman, *Science*, 1995, **267**, 316–317.
- 22 Y. S. Li, P. G. Milner, A. K. Chauhan, M. A. Watson, R. M. Hoffman, C. M. Kodner, J. Milbrandt and T. F. Deuel, *Science*, 1990, **250**, 1690.
- 23 T. Iwahara, J. Fujimoto, D. Wen, R. Cupples, N. Bucay, T. Arakawa, S. Mori, B. Ratzkin and T. Yamamoto, *Oncogene*, 1997, **14**, 439.
- 24 S. W. Morris, C. Naeve, P. Mathew, P. L. James, M. N. Kirstein, X. Cui and D. P. Witte, *Oncogene*, 1997, **14**, 2175–2188.
- 25 Y. Ben-Neriah and A. R. Bauskin, *Nature*, 1988, **333**, 672–676.
- 26 A. Bernards and S. M. de la Monte, *EMBO J.*, 1990, **9**, 2279–2287.
- 27 Y. Maru, H. Hirai and F. Takaku, *Oncog. Res.*, 1990, **5**, 199–204.
- 28 H. Kozutsumi, H. Toyoshima, K. Hagiwara, Y. Yazaki and H. Hirai, *Oncogene*, 1994, **9**, 2991–2998.
- 29 S. W. Morris, C. Naeve, P. Mathew, P. L. James, M. N. Kirstein, X. Cui and D. P. Witte, *Oncogene*, 1997, **14**, 2175–2188.
- 30 K. Pulford, L. Lamant, S. W. Morris, L. H. Butler, K. M. Wood, D. Stroud, G. Delsol and D. Y. Mason, *Blood*, 1997, **89**, 1394–1404.
- 31 R. Chiarle, C. Voena, C. Ambrogio, R. Piva and G. Inghirami, *Nat. Rev. Cancer*, 2008, **8**, 11–23.
- 32 R. H. Palmer, E. Vernersson, C. Grabbe and B. Hallberg, *Biochem. J.*, 2009, **420**, 345–361.



- 33 A. Barreca, E. Lasorsa, L. Riera, R. Machiorlatti, R. Piva, M. Ponzoni, I. Kwee, F. Bertoni, P. P. Piccaluga, S. A. Pileri and G. Inghirami, *J. Mol. Endocrinol.*, 2011, **47**, R11–R23.
- 34 S. Morris, M. Kirstein, M. Valentine, K. Dittmer, D. Shapiro, D. Saltman and A. Look, *Science*, 1994, **263**, 1281–1284.
- 35 B. Hallberg and R. Palmer, *Nat. Rev. Cancer*, 2013, **13**, 685–700.
- 36 H. Mano, *Cancer Sci.*, 2008, **99**, 2349–2355.
- 37 M. Soda, Y. Choi, M. Enomoto, S. Takada, Y. Yamashita, S. Ishikawa, S.-i. Fujiwara, H. Watanabe, K. Kurashina, H. Hatanaka, M. Bando, S. Ohno, Y. Ishikawa, H. Aburatani, T. Niki, Y. Sohara, Y. Sugiyama and H. Mano, *Nature*, 2007, **448**, 561–566.
- 38 A. T. Shaw, B. Y. Yeap, M. Mino-Kenudson, S. R. Digumarthy, D. B. Costa, R. S. Heist, B. Solomon, H. Stubbs, S. Admane, U. McDermott, J. Settleman, S. Kobayashi, E. J. Mark, S. J. Rodig, L. R. Chirieac, E. L. Kwak, T. J. Lynch and A. J. Iafrate, *J. Clin. Oncol.*, 2009, **27**, 4247–4253.
- 39 T. Sasaki, S. Rodig, L. Chirieac and P. Jänne, *Eur. J. Cancer*, 2010, **46**, 1773–1780.
- 40 J. Tanizaki, I. Okamoto, K. Takezawa, K. Sakai, K. Azuma, K. Kuwata, H. Yamaguchi, E. Hatashita, K. Nishio, P. A. Janne and K. Nakagawa, *Br. J. Cancer*, 2012, **106**, 763–767.
- 41 K. Rikova, A. Guo, Q. Zeng, A. Possemato, J. Yu, H. Haack, J. Nardone, K. Lee, C. Reeves, Y. Li, Y. Hu, Z. Tan, M. Stokes, L. Sullivan, J. Mitchell, R. Wetzell, J. Macneill, J. M. Ren, J. Yuan, C. E. Bakalarski, J. Villen, J. M. Kornhauser, B. Smith, D. Li, X. Zhou, S. P. Gygi, T. L. Gu, R. D. Polakiewicz, J. Rush and M. J. Comb, *Cell*, 2007, **131**, 1190–1203.
- 42 C. Griffin, A. Hawkins, C. Dvorak, C. Henkle, T. Ellingham and E. Perlman, *Cancer Res.*, 1999, **59**, 2776–2780.
- 43 B. Hallberg and R. H. Palmer, *Nat. Rev. Cancer*, 2013, **13**, 685–700.
- 44 F.-R. Jazii, Z. Najafi, R. Malekzadeh, T.-P. Conrads, A.-A. Ziaee, C. Abnet, M. Yazdznbod, A.-A. Karkhane and G.-H. Salekdeh, *World J. Gastroenterol.*, 2006, **12**, 7104–7112.
- 45 X.-L. Du, H. Hu, D.-C. Lin, S.-H. Xia, X.-M. Shen, Y. Zhang, M.-L. Luo, Y.-B. Feng, Y. Cai, X. Xu, Y.-L. Han, Q.-M. Zhan and M.-R. Wang, *J. Mol. Med.*, 2007, **85**, 863–875.
- 46 L. Debelenko, S. Raimondi, N. Daw, B. Shivakumar, D. Huang, M. Nelson and J. Bridge, *Mod. Pathol.*, 2011, **24**, 430–442.
- 47 J. Cook, L. Dehner, M. Collins, Z. Ma, S. Morris, C. Coffin and D. Hill, *Am. J. Surg. Pathol.*, 2001, **25**, 1364–1371.
- 48 Y. Chen, J. Takita, Y. Choi, M. Kato, M. Ohira, M. Sanada, L. Wang, M. Soda, A. Kikuchi, T. Igarashi, A. Nakagawara, Y. Hayashi, H. Mano and S. Ogawa, *Nature*, 2008, **455**, 971–974.
- 49 H. Carén, F. Abel, P. Kogner and T. Martinsson, *Biochem. J.*, 2008, **416**, 153–159.
- 50 R. George, T. Sanda, M. Hanna, S. Fröhling, W. Luther, J. Zhang, Y. Ahn, W. Zhou, W. London, P. McGrady, L. Xue, S. Zozulya, V. Gregor, T. Webb, N. Gray, D. Gilliland, L. Diller, H. Greulich, S. Morris, M. Meyerson and A. Look, *Nature*, 2008, **455**, 975–978.
- 51 I. Janoueix-Lerosey, D. Lequin, L. Brugières, A. Ribeiro, L. de Pontual, V. Combaret, V. Raynal, A. Puisieux, G. Schleiermacher, G. Pierron, D. Valteau-Couanet, T. Frebourg, J. Michon, S. Lyonnet, J. Amiel and O. Delattre, *Nature*, 2008, **455**, 967–970.
- 52 Y. Mossé, M. Laudenslager, L. Longo, K. Cole, A. Wood, E. Attiyeh, M. Laquaglia, R. Sennett, J. Lynch, P. Perri, G. Laureys, F. Speleman, C. Kim, C. Hou, H. Hakonarson, A. Torkamani, N. Schork, G. Brodeur, G. Tonini, E. Rappaport, M. Devoto and J. Maris, *Nature*, 2008, **455**, 930–935.
- 53 R. Palmer, E. Vernersson, C. Grabbe and B. Hallberg, *Biochem. J.*, 2009, **420**, 345–361.
- 54 S. Ogawa, J. Takita, M. Sanada and Y. Hayashi, *Cancer Sci.*, 2011, **102**, 302–308.
- 55 R. Roskoski Jr, *Pharmacol. Res.*, 2013, **68**, 68–94.
- 56 A. Galkin, J. Melnick, S. Kim, T. Hood, N. Li, L. Li, G. Xia, R. Steensma, G. Chopiuk, J. Jiang, Y. Wan, P. Ding, Y. Liu, F. Sun, P. Schultz, N. Gray and M. Warmuth, *Proc. Natl. Acad. Sci. U. S. A.*, 2007, **104**, 270–275.
- 57 J. P. Koivunen, C. Mermel, K. Zejnullahu, C. Murphy, E. Lifshits, A. J. Holmes, H. G. Choi, J. Kim, D. Chiang, R. Thomas, J. Lee, W. G. Richards, D. J. Sugarbaker, C. Ducko, N. Lindeman, J. P. Marcoux, J. A. Engelman, N. S. Gray, C. Lee, M. Meyerson and P. A. Jänne, *Clin. Cancer Res.*, 2008, **14**, 4275–4283.
- 58 P. Minoo and H. Y. Wang, *Int. J. Clin. Exp. Pathol.*, 2012, **5**, 397–410.
- 59 J. J. Cui, M. Tran-Dube, H. Shen, M. Nambu, P. P. Kung, M. Pairish, L. Jia, J. Meng, L. Funk, I. Botrous, M. McTigue, N. Grodsky, K. Ryan, E. Padrique, G. Alton, S. Timofeevski, S. Yamazaki, Q. Li, H. Zou, J. Christensen, B. Mroczkowski, S. Bender, R. S. Kania and M. P. Edwards, *J. Med. Chem.*, 2011, **54**, 6342–6363.
- 60 A. T. Shaw, U. Yasothan and P. Kirkpatrick, *Nat. Rev. Drug Discovery*, 2011, **10**, 897–898.
- 61 FDA News Release “FDA expands use of Xalkori to treat rare form of advanced non-small cell lung cancer”, <https://www.fda.gov/newsevents/newsroom/pressannouncements/ucm490329.htm>, (accessed on April 2018).
- 62 J. F. Gainor and A. T. Shaw, *J. Clin. Oncol.*, 2013, **31**, 3987–3996.
- 63 A. T. Shaw and J. A. Engelman, *J. Clin. Oncol.*, 2013, **31**, 1105–1111.
- 64 L. Friboulet, N. Li, R. Katayama, C. C. Lee, J. F. Gainor, A. S. Crystal, P. Y. Michellys, M. M. Awad, N. Yanagitani, S. Kim, A. C. Piferdekamper, J. Li, S. Kasibhatla, F. Sun, X. Sun, S. Hua, P. McNamara, S. Mahmood, E. L. Lockerman, N. Fujita, M. Nishio, J. L. Harris, A. T. Shaw and J. A. Engelman, *Cancer Discovery*, 2014, **4**, 662–673.
- 65 A. T. Shaw, S. H. Ou, Y. J. Bang, D. R. Camidge, B. J. Solomon, R. Salgia, G. J. Riely, M. Varella-Garcia, G. I. Shapiro, D. B. Costa, R. C. Doebele, L. P. Le, Z. Zheng, W. Tan, P. Stephenson, S. M. Shreeve,



- L. M. Tye, J. G. Christensen, K. D. Wilner, J. W. Clark and A. J. Iafrate, *N. Engl. J. Med.*, 2014, **371**, 1963–1971.
- 66 M. Caccese, R. Ferrara, S. Pilotto, L. Carbognin, G. Grizzi, A. Calio, M. Brunelli, F. Cuppone, S. Petraglia, A. Scarpa, G. Tortora and E. Bria, *Expert Opin. Pharmacother.*, 2016, **17**, 2253–2266.
- 67 Y. L. Choi, M. Soda, Y. Yamashita, T. Ueno, J. Takashima, T. Nakajima, Y. Yatabe, K. Takeuchi, T. Hamada, H. Haruta, Y. Ishikawa, H. Kimura, T. Mitsudomi, Y. Tanio and H. Mano, *N. Engl. J. Med.*, 2010, **363**, 1734–1739.
- 68 T. Sasaki, K. Okuda, W. Zheng, J. Butrynski, M. Capelletti, L. Wang, N. S. Gray, K. Wilner, J. G. Christensen, G. Demetri, G. I. Shapiro, S. J. Rodig, M. J. Eck and P. A. Janne, *Cancer Res.*, 2010, **70**, 10038–10043.
- 69 T. Sasaki, J. Koivunen, A. Ogino, M. Yanagita, S. Nikiforow, W. Zheng, C. Lathan, J. P. Marcoux, J. Du, K. Okuda, M. Capelletti, T. Shimamura, D. Ercan, M. Stumpfova, Y. Xiao, S. Weremowicz, M. Butaney, S. Heon, K. Wilner, J. G. Christensen, M. J. Eck, K. K. Wong, N. Lindeman, N. S. Gray, S. J. Rodig and P. A. Janne, *Cancer Res.*, 2011, **71**, 6051–6060.
- 70 R. C. Doebele, A. B. Pilling, D. L. Aisner, T. G. Kutateladze, A. T. Le, A. J. Weickhardt, K. L. Kondo, D. J. Linderman, L. E. Heasley, W. A. Franklin, M. Varela-Garcia and D. R. Camidge, *Clin. Cancer Res.*, 2012, **18**, 1472–1482.
- 71 C. M. Lovly and W. Pao, *Sci. Transl. Med.*, 2012, **4**, 120–122.
- 72 J. F. Gainor and A. T. Shaw, *J. Clin. Oncol.*, 2013, **31**, 3987–3996.
- 73 K. Kinoshita, K. Asoh, N. Furuichi, T. Ito, H. Kawada, S. Hara, J. Ohwada, T. Miyagi, T. Kobayashi, K. Takanashi, T. Tsukaguchi, H. Sakamoto, T. Tsukuda and N. Oikawa, *Bioorg. Med. Chem.*, 2012, **20**, 1271–1280.
- 74 T. W. Johnson, P. F. Richardson, S. Bailey, A. Brooun, B. J. Burke, M. R. Collins, J. J. Cui, J. G. Deal, Y.-L. Deng, D. Dinh, L. D. Engstrom, M. He, J. Hoffman, R. L. Hoffman, Q. Huang, R. S. Kania, J. C. Kath, H. Lam, J. L. Lam, P. T. Le, L. Lingardo, W. Liu, M. McTigue, C. L. Palmer, N. W. Sach, T. Smeal, G. L. Smith, A. E. Stewart, S. Timofeevski, H. Zhu, J. Zhu, H. Y. Zou and M. P. Edwards, *J. Med. Chem.*, 2014, **57**, 4720–4744.
- 75 B. J. Solomon, T. M. Bauer, E. Felip, B. Besse, L. P. James, J. S. Clancy, K. J. Klamerus, J.-F. Martini, A. Abbattista and A. T. Shaw, *J. Clin. Oncol.*, 2016, **34**, 9009.
- 76 E. Felip, T. Bauer, B. Solomon, B. Besse, L. James, J. Clancy, K. Klamerus, J. F. Martini, A. Abbattista and A. Shaw, *J. Thorac. Oncol.*, 2016, **12**, S383–S384.
- 77 S. Basit, Z. Ashraf, K. Lee and M. Latif, *Eur. J. Med. Chem.*, 2017, **134**, 348–356.
- 78 S. Basit, Z. Ashraf, K. Lee and M. Latif, *Eur. J. Med. Chem.*, 2017, **134**, 348–356.
- 79 V. Subbiah, D. S. Hong and F. Meric-Bernstam, *Proc. Natl. Acad. Sci. U. S. A.*, 2016, **113**, E1419–E1420.
- 80 I. Dagogo-Jack and A. T. Shaw, *J. Clin. Oncol.*, 2017, **35**, 2595–2597.
- 81 S. M. Lim, H. R. Kim, J.-S. Lee, K. H. Lee, Y.-G. Lee, Y. J. Min, E. K. Cho, S. S. Lee, B.-S. Kim, M. Y. Choi, H. S. Shim, J.-H. Chung, Y. L. Choi, M. J. Lee, M. Kim, J.-H. Kim, S. M. Ali, M.-J. Ahn and B. C. Cho, *J. Clin. Oncol.*, 2017, **35**, 2613–2618.
- 82 S. Dhillon and M. Clark, *Drugs*, 2014, **74**, 1285–1291.
- 83 S. Khozin, G. M. Blumenthal, L. Zhang, S. Tang, M. Brower, E. Fox, W. Helms, R. Leong, P. Song, Y. Pan, Q. Liu, P. Zhao, H. Zhao, D. Lu, Z. Tang, A. Al Hakim, K. Boyd, P. Keegan, R. Justice and R. Pazdur, *Clin. Cancer Res.*, 2015, **21**, 2436.
- 84 M. R. Cooper, H. Chim, H. Chan and C. Durand, *Ann. Pharmacother.*, 2015, **49**, 107–112.
- 85 I. B. Muller, A. J. De Langen, R. J. Honeywell, E. Giovannetti and G. J. Peters, *Expert Rev. Anticancer Ther.*, 2016, **16**, 147–157.
- 86 T. S. K. Mok, L. Crino, E. Felip, R. Salgia, T. De Pas, D. S. W. Tan and L. Q. M. Chow, *Cancer Treat. Rev.*, 2017, **55**, 181–189.
- 87 A. T. Shaw, T. M. Kim, L. Crino, C. Gridelli, K. Kiura, G. Liu, S. Novello, A. Bearz, O. Gautschi, T. Mok, M. Nishio, G. Scagliotti, D. R. Spigel, S. Deudon, C. Zheng, S. Pantano, P. Urban, C. Massacesi, K. Viraswami-Appanna and E. Felip, *Lancet Oncol.*, 2017, **18**, 874–886.
- 88 T. Kodama, T. Tsukaguchi, M. Yoshida, O. Kondoh and H. Sakamoto, *Cancer Lett.*, 2014, **351**, 215–221.
- 89 Y. Ohe, M. Nishio, K. Kiura, T. Seto, K. Nakagawa, M. Maemondo, A. Inoue, T. Hida, H. Yoshioka, M. Harada, N. Nogami, H. Murakami, K. Takeuchi, T. Shimada, H. Kuriki, T. Tanaka and T. Tamura, *J. Clin. Oncol.*, 2015, **33**, 8061.
- 90 L. Gandhi, S. I. Ou, A. T. Shaw, F. Barlesi, A. C. Dingemans, D. W. Kim, D. R. Camidge, B. G. M. Hughes, J. C. Yang, J. de Castro, L. Crino, H. Lena, P. Do, S. Golding, W. Bordogna, A. Zeaiter, A. Kotb and S. Gadgeel, *Eur. J. Cancer*, 2017, **82**, 27–33.
- 91 T. Tamura, T. Seto, K. Nakagawa, M. Maemondo, A. Inoue, T. Hida, H. Yoshioka, M. Harada, Y. Ohe, N. Nogami, H. Murakami, K. Takeuchi, T. Asakawa, K. Kikuchi, T. Tanaka and M. Nishio, *Int. J. Radiat. Oncol., Biol., Phys.*, 2014, **90**, S6.
- 92 S. Peters, D. R. Camidge, A. T. Shaw, S. Gadgeel, J. S. Ahn, D. W. Kim, S. I. Ou, M. Perol, R. Dziadziuszko, R. Rosell, A. Zeaiter, E. Mitry, S. Golding, B. Balas, J. Noe, P. N. Morcos and T. Mok, *N. Engl. J. Med.*, 2017, **377**, 829–838.
- 93 T. Seto, K. Kiura, M. Nishio, K. Nakagawa, M. Maemondo, A. Inoue, T. Hida, N. Yamamoto, H. Yoshioka, M. Harada, Y. Ohe, N. Nogami, K. Takeuchi, T. Shimada, T. Tanaka and T. Tamura, *Lancet Oncol.*, 2013, **14**, 590–598.
- 94 H. Sakamoto, T. Tsukaguchi, S. Hiroshima, T. Kodama, T. Kobayashi, T. A. Fukami, N. Oikawa, T. Tsukuda, N. Ishii and Y. Aoki, *Cancer Cell*, 2011, **19**, 679–690.
- 95 K. Kinoshita, T. Kobayashi, K. Asoh, N. Furuichi, T. Ito, H. Kawada, S. Hara, J. Ohwada, K. Hattori, T. Miyagi, W.-S. Hong, M.-J. Park, K. Takanashi, T. Tsukaguchi, H. Sakamoto, T. Tsukuda and N. Oikawa, *J. Med. Chem.*, 2011, **54**, 6286–6294.
- 96 K. Kinoshita, Y. Ono, T. Emura, K. Asoh, N. Furuichi, T. Ito, H. Kawada, S. Tanaka, K. Morikami, T. Tsukaguchi,



- H. Sakamoto, T. Tsukuda and N. Oikawa, *Bioorg. Med. Chem. Lett.*, 2011, **21**, 3788–3793.
- 97 X. Jiang, J. Zhou, J. Ai, Z. Song, X. Peng, L. Xing, Y. Xi, J. Guo, Q. Yao, J. Ding, M. Geng and A. Zhang, *Eur. J. Med. Chem.*, 2015, **105**, 39–56.
- 98 X. Jiang, J. Zhou, J. Ai, Z. Song, X. Peng, L. Xing, Y. Xi, J. Guo, Q. Yao, J. Ding, M. Geng and A. Zhang, *Eur. J. Med. Chem.*, 2015, **105**, 39–56.
- 99 Z. Song, Z. Xia, Y. Ji, L. Xing, Y. Gao, J. Ai, M. Geng and A. Zhang, *Eur. J. Med. Chem.*, 2016, **118**, 244–249.
- 100 M. Han, C. Wang, Y. Ji, Z. Song, L. Xing, Y. Su, X. Wang, A. Zhang, J. Ai and M. Geng, *Bioorg. Med. Chem. Lett.*, 2016, **26**, 5399–5402.
- 101 S. R. Peddi, S. K. Sivan and V. Manga, *J. Recept. Signal Transduction*, 2016, **36**, 488–504.
- 102 J. M. Hatcher, M. Bahcall, H. G. Choi, Y. Gao, T. Sim, R. George, P. A. Janne and N. S. Gray, *J. Med. Chem.*, 2015, **58**, 9296–9308.
- 103 S. Dhillon and M. Clark, *Drugs*, 2014, **74**, 1285–1291.
- 104 T. W. Johnson, P. F. Richardson, S. Bailey, A. Brooun, B. J. Burke, M. R. Collins, J. J. Cui, J. G. Deal, Y. L. Deng, D. Dinh, L. D. Engstrom, M. He, J. Hoffman, R. L. Hoffman, Q. Huang, R. S. Kania, J. C. Kath, H. Lam, J. L. Lam, P. T. Le, L. Lingardo, W. Liu, M. McTigue, C. L. Palmer, N. W. Sach, T. Smeal, G. L. Smith, A. E. Stewart, S. Timofeevski, H. Zhu, J. Zhu, H. Y. Zou and M. P. Edwards, *J. Med. Chem.*, 2014, **57**, 4720–4744.
- 105 S. Khozin, G. M. Blumenthal, L. Zhang, S. Tang, M. Brower, E. Fox, W. Helms, R. Leong, P. Song, Y. Pan, Q. Liu, P. Zhao, H. Zhao, D. Lu, Z. Tang, A. Al Hakim, K. Boyd, P. Keegan, R. Justice and R. Pazdur, *Clin. Cancer Res.*, 2015, **21**, 2436–2439.
- 106 E. Larkins, G. M. Blumenthal, H. Chen, K. He, R. Agarwal, G. Gieser, O. Stephens, E. Zahalka, K. Ringgold, W. Helms, S. Shord, J. Yu, H. Zhao, G. Davis, A. E. McKee, P. Keegan and R. Pazdur, *Clin. Cancer Res.*, 2016, **22**, 5171–5176.
- 107 <https://clinicaltrials.gov/ct2/show/NCT01970865>, <https://clinicaltrials.gov/show/NCT02584634>, <https://clinicaltrials.gov/ct2/show/NCT02927340>.
- 108 A. V. Galkin, J. S. Melnick, S. Kim, T. L. Hood, N. Li, L. Li, G. Xia, R. Steensma, G. Chopiuk, J. Jiang, Y. Wan, P. Ding, Y. Liu, F. Sun, P. G. Schultz, N. S. Gray and M. Warmuth, *Proc. Natl. Acad. Sci. U. S. A.*, 2007, **104**, 270–275.
- 109 C. H. Park, H. Choe, I.-Y. Jang, S. Y. Kwon, M. Latif, H. K. Lee, H. J. Lee, E. H. Yang, J. I. Yun, C. H. Chae, S. Y. Cho, S. U. Choi, J. D. Ha, H. Jung, H. R. Kim, P. Kim, C. O. Lee, C.-S. Yun and K. Lee, *Bioorg. Med. Chem. Lett.*, 2013, **23**, 6192–6196.
- 110 T. H. Marsilje, W. Pei, B. Chen, W. Lu, T. Uno, Y. Jin, T. Jiang, S. Kim, N. Li, M. Warmuth, Y. Sarkisova, F. Sun, A. Steffy, A. C. Pferdekamper, A. G. Li, S. B. Joseph, Y. Kim, B. Liu, T. Tunland, X. Cui, N. S. Gray, R. Steensma, Y. Wan, J. Jiang, G. Chopiuk, J. Li, W. P. Gordon, W. Richmond, K. Johnson, J. Chang, T. Groessl, Y. Q. He, A. Phimister, A. Aycinena, C. C. Lee, B. Bursulaya, D. S. Karanewsky, H. M. Seidel, J. L. Harris and P. Y. Michellys, *J. Med. Chem.*, 2013, **56**, 5675–5690.
- 111 W.-S. Huang, S. Liu, D. Zou, M. Thomas, Y. Wang, T. Zhou, J. Romero, A. Kohlmann, F. Li, J. Qi, L. Cai, T. A. Dwight, Y. Xu, R. Xu, R. Dodd, A. Toms, L. Parillon, X. Lu, R. Anjum, S. Zhang, F. Wang, J. Keats, S. D. Wardwell, Y. Ning, Q. Xu, L. E. Moran, Q. K. Mohemmad, H. G. Jang, T. Clackson, N. I. Narasimhan, V. M. Rivera, X. Zhu, D. Dalgarno and W. C. Shakespeare, *J. Med. Chem.*, 2016, **59**, 4948–4964.
- 112 G. R. Ott, M. Cheng, K. S. Learn, J. Wagner, D. E. Gingrich, J. G. Lisko, M. Curry, E. F. Mesaros, A. K. Ghose, M. R. Quail, W. Wan, L. Lu, P. Dobrzanski, M. S. Albom, T. S. Angeles, K. Wells-Knecht, Z. Huang, L. D. Aimone, E. Bruckheimer, N. Anderson, J. Friedman, S. V. Fernandez, M. A. Ator, B. A. Ruggeri and B. D. Dorsey, *J. Med. Chem.*, 2016, **59**, 7478–7496.
- 113 J. C. Soria, D. S. W. Tan, R. Chiari, Y. L. Wu, L. Paz-Ares, J. Wolf, S. L. Geater, S. Orlov, D. Cortinovis, C. J. Yu, M. Hochmair, A. B. Cortot, C. M. Tsai, D. Moro-Sibilot, R. G. Campelo, T. McCulloch, P. Sen, M. Dugan, S. Pantano, F. Branle, C. Massacesi and G. de Castro Jr, *Lancet*, 2017, **389**, 917–929.
- 114 J. Mazieres, G. Zalcman, L. Crino, P. Biondani, F. Barlesi, T. Filleron, A. M. Dingemans, H. Lena, I. Monnet, S. I. Rothschild, F. Cappuzzo, B. Besse, L. Thiberville, D. Rouviere, R. Dziadziuszko, E. F. Smit, J. Wolf, C. Spirig, N. Pecuchet, F. Leenders, J. M. Heuckmann, J. Diebold, J. D. Milia, R. K. Thomas and O. Gautschi, *J. Clin. Oncol.*, 2015, **33**, 992–999.
- 115 T. P. Heffron, *J. Med. Chem.*, 2016, **59**, 10030–10066.
- 116 H. Y. Zou, Q. Li, L. D. Engstrom, M. West, V. Appleman, K. A. Wong, M. McTigue, Y.-L. Deng, W. Liu, A. Brooun, S. Timofeevski, S. R. P. McDonnell, P. Jiang, M. D. Falk, P. B. Lappin, T. Affolter, T. Nichols, W. Hu, J. Lam, T. W. Johnson, T. Smeal, A. Charest and V. R. Fantin, *Proc. Natl. Acad. Sci. U. S. A.*, 2015, **112**, 3493–3498.
- 117 S. Zhang, R. Anjum, R. Squillace, S. Nadworny, T. Zhou, J. Keats, Y. Ning, S. D. Wardwell, D. Miller, Y. Song, L. Eichinger, L. Moran, W. S. Huang, S. Liu, D. Zou, Y. Wang, Q. Mohemmad, H. G. Jang, E. Ye, N. Narasimhan, F. Wang, J. Miret, X. Zhu, T. Clackson, D. Dalgarno, W. C. Shakespeare and V. M. Rivera, *Clin. Cancer Res.*, 2016, **22**, 5527–5538.
- 118 D.-W. Kim, M. Tiseo, M.-J. Ahn, *et al.*, Brigatinib (BRG) in patients (pts) with crizotinib (CRZ)-refractory ALK+ non-small cell lung cancer (NSCLC): First report of efficacy and safety from a pivotal randomized phase (ph) 2 trial (ALTA), *J. Clin. Oncol.*, 2016, **34**, 9007.
- 119 <https://clinicaltrials.gov/show/NCT02737501>, (accessed on May 13, 2017).
- 120 M. Cheng, M. R. Quail, D. E. Gingrich, G. R. Ott, L. Lu, W. Wan, M. S. Albom, T. S. Angeles, L. D. Aimone, F. Cristofani, R. Machiorlatti, C. Abele, M. A. Ator, B. D. Dorsey, G. Inghirami and B. A. Ruggeri, *Mol. Cancer Ther.*, 2012, **11**, 670–679.



- 121 M. J. Jacobs, G. R. Ott and L. Courvoisier, *WO* 2013134353 A1, 2013.
- 122 G. R. Ott, M. Cheng, K. S. Learn, J. Wagner, D. E. Gingrich, J. G. Lisko, M. Curry, E. F. Mesaros, A. K. Ghose, M. R. Quail, W. Wan, L. Lu, P. Dobrzanski, M. S. Albom, T. S. Angeles, K. Wells-Knecht, Z. Huang, L. D. Aimone, E. Bruckheimer, N. Anderson, J. Friedman, S. V. Fernandez, M. A. Ator, B. A. Ruggeri and B. D. Dorsey, *J. Med. Chem.*, 2016, **59**, 7478–7496.
- 123 Information from the National Library of Medicine, “To determine the maximum tolerated dose of oral CEP-37440 in patients with advanced or metastatic solid tumors”, <https://clinicaltrials.gov/ct2/show/NCT01922752>, (accessed on March 2018).
- 124 S. N. Gettinger, L. A. Bazhenova, C. J. Langer, R. Salgia, K. A. Gold, R. Rosell, A. T. Shaw, G. J. Weiss, M. Tugnait, N. I. Narasimhan, D. J. Dorer, D. Kerstein, V. M. Rivera, T. Clackson, F. G. Haluska and D. R. Camidge, *Lancet Oncol.*, 2016, **17**, 1683–1696.
- 125 K. Uchibori, N. Inase, M. Araki, M. Kamada, S. Sato, Y. Okuno, N. Fujita and R. Katayama, *Nat. Commun.*, 2017, **8**, 14768.
- 126 A. Markham, *Drugs*, 2017, **77**, 1131–1135.
- 127 T. O'Hare, R. Pollock, E. P. Stoffregen, J. A. Keats, O. M. Abdullah, E. M. Moseson, V. M. Rivera, H. Tang, C. A. Metcalf 3rd, R. S. Bohacek, Y. Wang, R. Sundaramoorthi, W. C. Shakespeare, D. Dalgarno, T. Clackson, T. K. Sawyer, M. W. Deininger and B. J. Druker, *Blood*, 2004, **104**, 2532–2539.
- 128 J. Bain, L. Plater, M. Elliott, N. Shpiro, C. J. Hastie, H. McLaughlan, I. Klevernic, J. Simon, C. Arthur, D. R. Alessi and P. Cohen, *Biochem. J.*, 2007, **408**, 297–315.
- 129 A. S. Corbin, S. Demehri, I. J. Griswold, Y. Wang, C. A. Metcalf 3rd, R. Sundaramoorthi, W. C. Shakespeare, J. Snodgrass, S. Wardwell, D. Dalgarno, J. Iulicci, T. K. Sawyer, M. C. Heinrich, B. J. Druker and M. W. Deininger, *Blood*, 2005, **106**, 227–234.
- 130 V. Rivera, R. Anjum, F. Wang, S. Zhang, J. Keats, Y. Ning, S. Wardwell, L. Moran, E. Ye and D. Chun, *Cancer Res.*, 2010, **70**, 3623.
- 131 S. Zhang, F. Wang, J. Keats, Y. Ning, S. D. Wardwell, L. Moran, Q. K. Mohemmad, E. Ye, R. Anjum, Y. Wang, X. Zhu, J. J. Miret, D. Dalgarno, N. I. Narasimhan, T. Clackson, W. C. Shakespeare and V. M. Rivera, *101st AACR Annual Meeting*, Washington, DC, USA, 2010, pp. 17–21.
- 132 Q. Huang, T. W. Johnson, S. Bailey, A. Brooun, K. D. Bunker, B. J. Burke, M. R. Collins, A. S. Cook, J. J. Cui, K. N. Dack, J. G. Deal, Y. L. Deng, D. Dinh, L. D. Engstrom, M. He, J. Hoffman, R. L. Hoffman, P. S. Johnson, R. S. Kania, H. Lam, J. L. Lam, P. T. Le, Q. Li, L. Lingardo, W. Liu, M. W. Lu, M. McTigue, C. L. Palmer, P. F. Richardson, N. W. Sach, H. Shen, T. Smeal, G. L. Smith, A. E. Stewart, S. Timofeevski, K. Tsaparikos, H. Wang, H. Zhu, J. Zhu, H. Y. Zou and M. P. Edwards, *J. Med. Chem.*, 2014, **57**, 1170–1187.
- 133 Y.-C. Kim, S. G. Brown, T. K. Harden, J. L. Boyer, G. Dubyak, B. F. King, G. Burnstock and K. A. Jacobson, *J. Med. Chem.*, 2001, **44**, 340–349.
- 134 J. Xuan, T.-T. Zeng, J.-R. Chen, L.-Q. Lu and W.-J. Xiao, *Chem.-Eur. J.*, 2015, **21**, 4962–4965.
- 135 The U.S. Food and Drug Administration, “Drug Safety and Availability”, https://www.accessdata.fda.gov/drugsatfda_docs/label/2017/208772lbl.pdf, accessed on Dec 12, 2017.
- 136 R. Rosell, S. N. Gettinger, L. A. Bazhenova, C. J. Langer, R. Salgia, A. T. Shaw, N. I. Narasimhan, D. J. Dorer, D. Kerstein and D. R. Camidge, *J. Thorac. Oncol.*, 2016, **11**, S114.
- 137 D.-W. Kim, M. Tiseo and M.-J. Ahn, *et al.*, Brigatinib (BRG) in patients (pts) with crizotinib (CRZ)-refractory ALK+ non-small cell lung cancer (NSCLC): first report of efficacy and safety from a pivotal randomized phase (ph) 2 trial (ALTA), *ASCO Meet Abstr.*, 2016, vol. 34(suppl. 15), p. 9060.
- 138 S. N. Gettinger, S. Zhang, J. G. Hodgson, *et al.*, Activity of brigatinib (BRG) in crizotinib (CRZ) resistant patients (pts) according to ALK mutation status, *ASCO Meet Abstr.*, 2016 May 31, vol. 34(suppl. 15), p. 9060.
- 139 S. N. Gettinger, S. Zhang and J. G. Hodgson, *et al.*, Activity of brigatinib (BRG) in crizotinib (CRZ) resistant patients (pts) according to ALK mutation status, *ASCO Meet Abstr.*, 2016 May 31, vol. 34(suppl. 15), p. 9060.
- 140 S. H. Ou, S. J. Klempner, J. R. Greenbowe, M. Azada, A. B. Schrock, S. M. Ali, J. S. Ross, P. J. Stephens and V. A. Miller, *J. Thorac. Oncol.*, 2014, **9**, 1821–1825.
- 141 J. K. Sabari, F. C. Santini, A. M. Schram, I. Bergagnini, R. Chen, C. Mrad, W. V. Lai, K. C. Arbour and A. Drilon, *OncoTargets Ther.*, 2017, **10**, 1983.
- 142 D.-W. Kim, M. Tiseo, M.-J. Ahn, K. L. Reckamp, K. H. Hansen, S.-W. Kim, R. M. Huber, H. J. West, H. J. M. Groen, M. J. Hochmair, N. B. Leigh, S. N. Gettinger, C. J. Langer, L. G. Paz-Ares, E. F. Smit, E. S. Kim, W. G. Reichmann, D. Kerstein, F. G. Haluska and D. R. Camidge, *J. Clin. Oncol.*, 2016, **34**, 9007.
- 143 C. J. Langer, S. N. Gettinger, L. Bazhenova, R. Salgia, K. A. Gold, R. Rosell, A. T. Shaw, G. J. Weiss, D. J. Dorer, V. M. Rivera, F. G. Haluska, D. Kerstein and D. R. Camidge, *J. Clin. Oncol.*, 2016, **34**, 9057.
- 144 J. Zhang, X. Deng, H. G. Choi, D. R. Alessi and N. S. Gray, *Bioorg. Med. Chem. Lett.*, 2012, **22**, 1864–1869.
- 145 S. Kanda and Y. Miyata, *Frontiers in Bioscience*, 2011, **16**, 1024–1035.
- 146 S. Hellwig, C. V. Miduturu, S. Kanda, J. Zhang, P. Filippakopoulos, E. Salah, X. Deng, H. G. Choi, W. Zhou, W. Hur, S. Knapp, N. S. Gray and T. E. Smithgall, *Chem. Biol.*, 2012, **19**, 529–540.
- 147 S. Hellwig, N. S. Gray and T. E. Smithgall, *Mol. Cancer Ther.*, 2014, **10**, A234.
- 148 U. McDermott, S. Sharma, L. Dowell, P. Greninger, C. Montagut, J. Lamb, H. Archibald, R. Raudales, A. Tam, D. Lee, S. Rothenberg, J. Supko, R. Sordella, L. Ulkus, A. Iafrate, S. Maheswaran, C. Njauw, H. Tsao, L. Drew, J. Hanke, X.-J. Ma, M. Erlander, N. Gray, D. Haber and



- J. Settleman, *Proc. Natl. Acad. Sci. U. S. A.*, 2007, **104**, 19936–19941.
- 149 D. P. Walker, F. C. Bi, A. S. Kalgutkar, J. N. Bauman, S. X. Zhao, J. R. Soglia, G. E. Aspnes, D. W. Kung, J. Klug-McLeod, M. P. Zawistoski, M. A. McGlynn, R. Oliver, M. Dunn, J. C. Li, D. T. Richter, B. A. Cooper, J. C. Kath, C. A. Hulford, C. L. Autry, M. J. Luzzio, E. J. Ung, W. G. Roberts, P. C. Bonnette, L. Buckbinder, A. Mistry, M. C. Griffor, S. Han and A. Guzman-Perez, *Bioorg. Med. Chem. Lett.*, 2008, **18**, 6071–6077.
- 150 R. S. Obach, A. S. Kalgutkar, J. R. Soglia and S. X. Zhao, *Chem. Res. Toxicol.*, 2008, **21**, 1814–1822.
- 151 H. Orhan and N. P. Vermeulen, *Curr. Drug Metab.*, 2011, **12**, 383–394.
- 152 T. A. Baillie, *Chem. Res. Toxicol.*, 2008, **21**, 129–137.
- 153 FDA approved drugs, “FDA broadens ceritinib indication to previously untreated ALK-positive metastatic NSCLC”, <https://www.fda.gov/Drugs/InformationOnDrugs/ApprovedDrugs/ucm560873.htm>, accessed on March, 2018.
- 154 K. L. Milkiewicz, L. R. Weinberg, M. S. Albom, T. S. Angeles, M. Cheng, A. K. Ghose, R. C. Roemmele, J. P. Theroff, T. L. Underiner, C. A. Zifcsak and B. D. Dorsey, *Bioorg. Med. Chem.*, 2010, **18**, 4351–4362.
- 155 W. Wan, M. S. Albom, L. Lu, M. R. Quail, N. C. Becknell, L. R. Weinberg, D. R. Reddy, B. P. Holskin, T. S. Angeles, T. L. Underiner, S. L. Meyer, R. L. Hudkins, B. D. Dorsey, M. A. Ator, B. A. Ruggeri and M. Cheng, *Blood*, 2006, **107**, 1617–1623.
- 156 R. Li, L. Xue, T. Zhu, Q. Jiang, X. Cui, Z. Yan, D. McGee, J. Wang, V. R. Gantla, J. C. Pickens, D. McGrath, A. Chucholowski, S. W. Morris and T. R. Webb, *J. Med. Chem.*, 2006, **49**, 1006–1015.
- 157 P. Sabbatini, S. Korenchuk, J. L. Rowand, A. Groy, Q. Liu, D. Leperi, C. Atkins, M. Dumble, J. Yang, K. Anderson, R. G. Kruger, R. R. Gontarek, K. R. Maksimchuk, S. Suravajjala, R. R. Lapierre, J. B. Shotwell, J. W. Wilson, S. D. Chamberlain, S. K. Rabindran and R. Kumar, *Mol. Cancer Ther.*, 2009, **8**, 2811–2820.
- 158 J. G. Christensen, H. Y. Zou, M. E. Arango, Q. Li, J. H. Lee, S. R. McDonnell, S. Yamazaki, G. R. Alton, B. Mroczkowski and G. Los, *Mol. Cancer Ther.*, 2007, **6**, 3314–3322.
- 159 R. Li and S. W. Morris, *Med. Res. Rev.*, 2008, **28**, 372–412.
- 160 C. C. Lee, Y. Jia, N. Li, X. Sun, K. Ng, E. Ambing, M. Y. Gao, S. Hua, C. Chen, S. Kim, P. Y. Michellys, S. A. Lesley, J. L. Harris and G. Spraggon, *Biochem. J.*, 2010, **430**, 425–437.
- 161 R. T. Bossi, M. B. Saccardo, E. Ardini, M. Menichincheri, L. Rusconi, P. Magnaghi, P. Orsini, N. Avanzi, A. L. Borgia, M. Nesi, T. Bandiera, G. Fogliatto and J. A. Bertrand, *Biochemistry*, 2010, **49**, 6813–6825.
- 162 D. E. Gingrich, J. G. Lisko, M. A. Curry, M. Cheng, M. Quail, L. Lu, W. Wan, M. S. Albom, T. S. Angeles, L. D. Aimone, R. C. Haltiwanger, K. Wells-Knecht, G. R. Ott, A. K. Ghose, M. A. Ator, B. Ruggeri and B. D. Dorsey, *J. Med. Chem.*, 2012, **55**, 4580–4593.
- 163 D. E. Gingrich, J. G. Lisko, M. A. Curry, M. Cheng, M. Quail, L. Lu, W. Wan, M. S. Albom, T. S. Angeles, L. D. Aimone, R. C. Haltiwanger, K. Wells-Knecht, G. R. Ott, A. K. Ghose, M. A. Ator, B. Ruggeri and B. D. Dorsey, *J. Med. Chem.*, 2012, **55**, 4580–4593.
- 164 M. Cheng, M. Quail, D. Gingrich, G. Ott, L. Lu, W. Wan, M. Albom, T. Angeles, L. Aimone, F. Cristofani, R. Machiorlatti, C. Abele, M. Ator, B. Dorsey, G. Inghirami and B. Ruggeri, *Mol. Cancer Ther.*, 2012, **11**, 670–679.
- 165 S. P. Allwein, R. C. Roemmele, J. J. Haley, D. R. Mowrey, D. E. Petrillo, J. J. Reif, D. E. Gingrich and R. P. Bakale, *Org. Process Res. Dev.*, 2012, **16**, 148–155.
- 166 G. R. Ott, R. Tripathy, M. Cheng, R. McHugh, A. V. Anzalone, T. L. Underiner, M. A. Curry, M. R. Quail, L. Lu, W. Wan, T. S. Angeles, M. S. Albom, L. D. Aimone, M. A. Ator, B. A. Ruggeri and B. D. Dorsey, *ACS Med. Chem. Lett.*, 2010, **1**, 493–498.
- 167 M. Cheng and G. R. Ott, *Anti-Cancer Agents Med. Chem.*, 2010, **10**, 236–249.
- 168 T. J. Liu, T. LaFortune, T. Honda, O. Ohmori, S. Hatakeyama, T. Meyer, D. Jackson, J. de Groot and W. K. Yung, *Mol. Cancer Ther.*, 2007, **6**, 1357–1367.
- 169 E. F. Mesaros, J. P. Burke, J. D. Parrish, B. J. Dugan, A. V. Anzalone, T. S. Angeles, M. S. Albom, L. D. Aimone, M. R. Quail, W. Wan, L. Lu, Z. Huang, M. A. Ator, B. A. Ruggeri, M. Cheng, G. R. Ott and B. D. Dorsey, *Bioorg. Med. Chem. Lett.*, 2011, **21**, 463–466.
- 170 E. F. Mesaros, J. P. Burke, J. D. Parrish, B. J. Dugan, A. V. Anzalone, T. S. Angeles, M. S. Albom, L. D. Aimone, M. R. Quail, W. Wan, L. Lu, Z. Huang, M. A. Ator, B. A. Ruggeri, M. Cheng, G. R. Ott and B. D. Dorsey, *Bioorg. Med. Chem. Lett.*, 2011, **21**, 463–466.
- 171 S. P. Allwein, D. R. Mowrey, D. E. Petrillo, J. J. Reif, V. C. Purohit, K. L. Milkiewicz, R. P. Bakale, M. A. Christie, M. A. Olsen, C. J. Neville and G. J. Gilmartin, *Org. Process Res. Dev.*, 2017, **21**, 740–747.
- 172 S. P. Allwein, R. P. Bakale, D. R. Mowrey, D. E. Petrillo and S. Kluwer, PCT Int. Appl., WO 2016105529 A1 20160630, 2016.
- 173 J. Zhao and J. L. Guan, *Cancer Metastasis Rev.*, 2009, **28**, 35–49.
- 174 J. Schwock, N. Dhani and D. W. Hedley, *Expert Opin. Ther. Targets*, 2010, **14**, 77–94.
- 175 F. J. Sulzmaier, C. Jean and D. D. Schlaepfer, *Nat. Rev. Cancer*, 2014, **14**, 598–610.
- 176 B. Y. Lee, P. Timpson, L. G. Horvath and R. J. Daly, *Pharmacol. Ther.*, 2015, **146**, 132–149.
- 177 I. Salem, M. Alsalahi, I. Chervoneva, L. D. Aburto, S. Addya, G. R. Ott, B. A. Ruggeri, M. Cristofanilli and S. V. Fernandez, *Breast Cancer Res.*, 2016, **18**, 37.
- 178 G. Sliwoski, S. Kothiwale, J. Meiler and E. W. Lowe, *Pharmacol. Rev.*, 2014, **66**, 334.
- 179 T. Usha, D. Shanmugarajan, A. K. Goyal, C. S. Kumar and S. K. Middha, *Curr. Top. Med. Chem.*, 2017, **17**, 3296–3307.
- 180 M. Zheng, J. Zhao, C. Cui, Z. Fu, X. Li, X. Liu, X. Ding, X. Tan, F. Li, X. Luo, K. Chen and H. Jiang, *Med. Res. Rev.*, 2018, **38**, 914–950, DOI: 10.1002/med.21483.
- 181 N. James, V. Shanthi and K. Ramanathan, *Appl. Biochem. Biotechnol.*, 2017, DOI: 10.1007/s12010-017-2650-x.



- 182 N. James and K. Ramanathan, *Cell Biochem. Biophys.*, 2017, 1–27, DOI: 10.1007/s12013-017-0800-y.
- 183 W. Fulong, L. Qianqian, W. Zhonghua, L. Dandan, P. Peng, Y. Yan, C. Siheng and W. Fanhong, *Lett. Drug Des. Discovery*, 2017, **14**, 270–286.
- 184 H. Z. Xie, H. Lan, Y. L. Pan, J. Zou, Z. R. Wang, L. L. Li, Q. Huang, H. Zhang and S. Y. Yang, *Chem. Biol. Drug Des.*, 2013, **81**, 175–184.
- 185 S. J. Macalino, V. Gosu, S. Hong and S. Choi, *Arch. Pharmacol. Res.*, 2015, **38**, 1686–1701.
- 186 W. Lu, R. Zhang, H. Jiang, H. Zhang and C. Luo, *Front. Chem.*, 2018, **6**, 57.
- 187 N. James and K. Ramanathan, *Appl. Biochem. Biotechnol.*, 2018, 1–24, DOI: 10.1007/s12010-018-2724-4.
- 188 C. H. Kang, J. I. Yun, K. Lee, C. O. Lee, H. K. Lee, C. S. Yun, J. Y. Hwang, S. Y. Cho, H. Jung, P. Kim, J. D. Ha, J. H. Jeon, S. U. Choi, H. G. Jeong, H. R. Kim and C. H. Park, *Biochem. Biophys. Res. Commun.*, 2015, **464**, 762–767.
- 189 G. R. Mathi, C. H. Kang, H. K. Lee, R. Achary, H. Y. Lee, J. Y. Lee, J. D. Ha, S. Ahn, C. H. Park, C. O. Lee, J. Y. Hwang, C. S. Yun, H. J. Jung, S. Y. Cho, H. R. Kim and P. Kim, *Eur. J. Med. Chem.*, 2017, **126**, 536–549.
- 190 Y. Wang, S. Chen, G. Hu, J. Wang, W. Gou, D. Zuo, Y. Gu, P. Gong and X. Zhai, *Eur. J. Med. Chem.*, 2018, **143**, 123–136.

