

Cite this: *RSC Adv.*, 2018, 8, 4766

A dual-emission nano-rod MOF equipped with carbon dots for visual detection of doxycycline and sensitive sensing of MnO_4^- †

Xin Fu, Rui Lv, Jian Su, Hui Li, Boyi Yang, Wen Gu* and Xin Liu *

Herein, ethanediamine-modified carbon dots (CDs) were encapsulated into luminescent MOF(Eu), which was designed for a dual-emission hybrid material (CDs@MOF(Eu)) with diverse fluorescence applications. This material exhibited high selectivity and sensitivity towards doxycycline. With an increasing concentration of doxycycline, the blue light emission of CDs could be quenched, whereas the red light emission of MOF(Eu) was enhanced. In view of this result, more convenient “test paper” was used first as a new tool for doxycycline detection, the colour of which turned from blue-purple to red as observed by the naked eyes under 365 nm UV-irradiation. This hybrid material also was a probe for sensing MnO_4^- with a low limit of detection and good anti-interference performance. We propose that CDs can improve detection sensitivity compared with the original MOF(Eu). The possible sensing mechanism was discussed in detail. Importantly, the feasibility of this composite for sensing doxycycline in a simulated biological system and sensing MnO_4^- in tap water was investigated.

Received 8th November 2017

Accepted 11th January 2018

DOI: 10.1039/c7ra12252g

rsc.li/rsc-advances

Introduction

At present, the abuse of antibiotics is becoming more common. Doxycycline is a tetracycline compound that can be used to treat infections of the respiratory, urinary, and biliary tracts caused by sensitive bacteria.¹ Doxycycline can also prevent falciparum malaria and leptospirosis infection, and is often used as a veterinary drug in animal breeding. However, the residue of doxycycline in water-based environments and everyday foods has several side effects, such as hepatotoxicity, yellow teeth, gastrointestinal symptoms and mentation changes, especially in children; in addition, doxycycline use can lead to antibiotic resistance.^{2,3} Numerous test methods have been developed to detect tetracycline antibiotics: electrochemical analysis,^{4,5} UV absorbance spectroscopy (UV-vis),^{6,7} liquid chromatography,^{8–10} flow-injection analysis,^{11,12} and fluorescence spectrophotometry.^{13,14} Fluorescence spectrophotometry stands out among these methods because it is sensitive, fast and cost-effective. Currently, “test paper” based on fluorescence spectrophotometry has become popular in molecular detection because it is convenient, inexpensive and easy to carry out. However, few studies on antibiotic detection using test paper have been

undertaken. Several materials have been used for doxycycline detection based on fluorescence spectrophotometry, especially quantum dots.¹⁵ During recent years, quantum-dot materials have been used in various applications that have advantages such as nanometre size, simple synthesis method, low toxicity and high quantum yield.^{16–18} Carbon dots (CDs) are a novel form of quantum dots. CDs have been applied to detect many metal ions,^{19–22} pollutants,^{23–25} biomolecules²⁶ and antibiotics²⁷ with high fluorescence selectivity and sensitivity.

Luminescent lanthanide metal organic frameworks (Ln-MOFs) are a class of crystalline materials that have attracted much attention in recent years. Ln-MOFs have large specific surface areas, adjustable pore sizes, unsaturated metal coordination sites, and good fluorescence of rare earth metal ions. Thus, several applications have been based on the advantages stated above, such as fluorescence probes for detection.^{28–30} If CDs are incorporated with MOFs, the formed hybrid materials would have enhanced fluorescence properties. If the MOFs have no fluorescence, dual-emission fluorescence could be obtained by doping rare earth ions, whereas MOFs could act as a “box” for loading CDs and rare earth ions. For instance, Dong *et al.* used Eu-MOFs encapsulating blue emission N,S-CDs to form a dual-emission material for visual detection of water in organic solvents. This material showed red light emission in organic solvents and blue light emission in water.³¹ Xu *et al.* synthesized MOF-253 encapsulating CDs and Eu^{3+} . The prepared fluorescent probe can detect Hg^{2+} in aqueous solution with a limit of detection (LOD) of 13 ppb. Hg^{2+} only quenched the fluorescence of CDs; the red emission of Eu^{3+} was not affected. Also, the

College of Chemistry, Key Laboratory of Advanced Energy Materials Chemistry (MOE), Tianjin Key Laboratory of Metal and Molecule Based Material Chemistry, Collaborative Innovation Center of Chemical Science and Engineering, Nankai University, Tianjin 300071, P. R. China. E-mail: liuxin64@nankai.edu.cn; guwen68@nankai.edu.cn

† Electronic supplementary information (ESI) available. See DOI: 10.1039/c7ra12252g



colour of this probe changed significantly under UV irradiation.³²

Potassium permanganate (KMnO₄) is a strong oxidant and often serves as an antiseptic and disinfectant. KMnO₄ is also applied for treating fish diseases and polluted water.³³ However, it is toxic, corrosive and harmful to human health, and can lead to respiratory infections and skin irritation.³⁴ Hence, the precise detection of MnO₄⁻ has become a major component of water-pollution detection for environmental evaluation.

In this work, we report a bifunctional hybrid material CDs@MOF(Eu) that can detect doxycycline and MnO₄⁻. This composite was synthesized through a simple method that induced blue light emission CDs into MOF(Eu). The ratio of two fluorescence peaks ($F_{616\text{ nm}}/F_{436\text{ nm}}$) exhibited excellent sensitivity and selectivity for doxycycline. Also, detection was possible with the naked eyes using test paper under UV irradiation of 365 nm. Compared with other materials, this hybrid material based on the fluorescence ratio of two peaks could reduce interference from the environment and minimize the drift value of the instrument, thereby making the results reliable.³⁵ Also, the two emissions may result in colour changes in the presence of doxycycline, which makes visual detection easier than that for single-emission materials. Besides, the composite could also be applied to MnO₄⁻ detection for instruments with a low LOD; CDs accelerated the speed of the detection and made the LOD lower compared with that of the original MOF(Eu). More importantly, this sensor also exhibited very promising applications for sensing doxycycline in simulated biological systems and for detecting MnO₄⁻ in tap water.

Experimental

Materials

Citric acid, ethylenediamine, 1,3,5-benzenetricarboxylic acid (H₃BTC), europium nitrate hexahydrate (Eu(NO₃)₃·6H₂O), tris(hydroxymethyl)aminomethane (Tris), DMF, ethanol, HCl, chlorotetracycline, doxycycline, isoniazid, oxytetracycline, neomycin, kanamycin, metronidazole, chloramphenicol, erythromycin, tetracycline, phenylalanine (Phe), valine (Val), cysteine (Cys), glutathione (GSH), aspartic acid (Asp), glycine (Gly), KNO₃, NaNO₃, Zn(NO₃)₂·6H₂O, Ca(NO₃)₂, KMnO₄, CH₃-COOK, K₂CO₃, KClO₃, K₂C₂O₄, K₂SO₃, KHCO₃, NaNO₂ and NaH₂PO₄·H₂O were of analytical grade and used without further purification. Ultra-pure grade water (18.2 MΩ) was used as the solvent in all experiments.

Apparatus

Transmission electron microscopy (TEM) was undertaken using an FEI Tecnai G2 S-Twin transmission electron microscope. Scanning electron microscopy (SEM) was done using a JEOL JSM-7500F field emission scanning electron microscope. Energy-dispersive X-ray spectroscopy (EDS) was done on a Merlin Compact scanning electron microscope. X-ray diffraction (XRD) patterns were measured on D8 Focus diffractometer with Cu-Kα radiation ($\lambda = 0.15405\text{ nm}$). Fourier transform infrared (FTIR) spectroscopy was undertaken on a Bruker Tensor 27 FTIR

spectrometer using KBr pellets. All fluorescence spectra were recorded on a FLS 920P Edinburgh instrument. N₂ adsorption-desorption was measured on a Micromeritics ASAP 2020 HD88 apparatus. UV-vis spectra were recorded on a Shimadzu UV-3600 spectrometer. X-ray photoelectron spectroscopy (XPS) was carried out on an Axis Ultra DLD X-ray photoelectron spectrometer.

Synthesis of CDs

CDs were synthesized according to a simple method. In brief, citric acid (1.0507 g) was added into 10 mL ultra-pure water and dissolved completely. Then, 335 μL ethylene diamine was injected quickly into the solution, the mixture transferred into a 25 mL Teflon™-lined autoclave, and maintained for 5 h at 200 °C. Finally, a dark-brown solution was obtained, filtered using a membrane, and placed in darkness for further use.

Synthesis of CDs@MOF(Eu)

The synthetic method of MOF(Eu) has been described.³⁶ First, a water solution of CDs was prepared with a proportion of $V_{\text{CDs}} : V_{\text{water}} = 1 : 10$ (the amount of CDs in each reaction kettle was $6.56 \times 10^{-4}\text{ mol}$). Typically, Eu(NO₃)₃·6H₂O (0.00331 g) and H₃BTC (0.0156 g) were added into a mixed solution containing 3 mL DMF and 3 mL ethanol. Then, 2 mL prepared CDs in water solution was injected into the mixture and sonicated for 5 min. The mixture was transferred to a 25 mL Teflon-lined autoclave and maintained for 24 h at 80 °C. A light-brown solid were obtained by centrifugation for 5 min at 8000 rpm. The precipitate was washed thrice with DMF and water. Then, the sample was immersed in methanol and CH₂Cl₂ for 12 h, respectively. Samples were obtained by centrifugation and then dried 12 h at 50 °C.

Detection of fluorescence and test paper

Initially, the concentration of tested molecule solutions was 10⁻² M. The sample solutions to be tested were prepared in 0.05 M Tris-HCl buffered solution (pH 7.3). Then, 1 mg CDs@MOF(Eu) was dispersed into 5 mL buffered solution and sonicated for 5 min to obtain a well-dispersed suspension which was used for luminescence experiments. For sensing doxycycline in real samples, Tris-buffered saline (0.1 M Tris-HCl-buffered solution containing 0.015 mol NaCl, pH 7.4) was chosen as a simulated biological system. All selectivity and anti-interference experiments were done by adding 100 μL analyte into the sample solution, and shaking thoroughly before detection. The test paper was made from filter paper and cut into rectangles. When detecting doxycycline, the test paper was soaked into sample solutions containing different concentrations of doxycycline for 2 min. Then, the test paper was clipped with tweezers and left to dry naturally in air. Photographs of test papers were taken in a dark box under UV irradiation at 365 nm.

Results and discussion

Characterization

The morphology of the composites was confirmed by TEM and SEM. Fig. 1a and b shows the images of CDs, which were



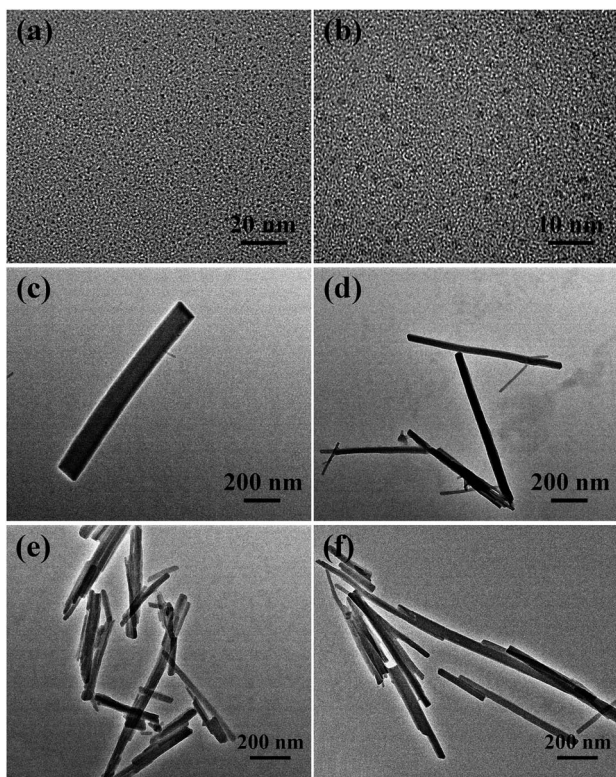


Fig. 1 Transmission electron micrographs of carbon dots (a and b), CDs@MOF(Eu) (c and d) and MOF(Eu) (e and f).

spherical and of size ≈ 2 nm. The composite and MOF(Eu) were all rod-like and of the same size (Fig. 1c–f, S1a and b†). The microstructure morphology of the composite is shown in Fig. S2.† The surface of the rod was smooth, which suggested that the added CDs were not attached to the surface of the MOFs. By virtue of their structure, the strong peaks of MOF(Eu) were not destroyed in the XRD pattern after combination with CDs (Fig. 2a). It was difficult to observe CDs in the TEM images of the composite (Fig. 2b) but a typical type-I isotherm of MOF was obtained. Also, a sharp decrease of the Brunauer–Emmett–Teller surface area from $102.8699 \text{ m}^2 \text{ g}^{-1}$ to $10.9979 \text{ m}^2 \text{ g}^{-1}$ demonstrated that CDs were encapsulated into MOF(Eu). FTIR spectra were also used to confirm the main groups in the composite (Fig. S3†). A broad peak at 3396 cm^{-1} was assigned to an O–H stretching vibration, and a sharp peak at 1614 cm^{-1} was from the

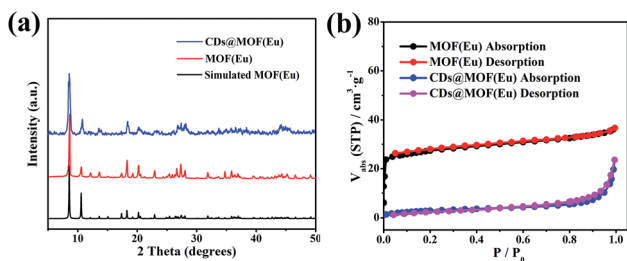


Fig. 2 (a) Comparison of the XRD patterns of MOF(Eu) and CDs@MOF(Eu) composite. (b) N₂ absorption–desorption isotherms of MOF(Eu) and CDs@MOF(Eu).

bending vibration of N–H. The band at 1114 cm^{-1} was attributed to C–N stretching vibrations, and the bands at 771 cm^{-1} and 726 cm^{-1} were from an N–H wagging vibration. These results indicated the functional groups in the composite and induced CDs did not destroy the structure of the complex.

Detection and sensing mechanism of doxycycline

The property of sensing antibiotics was observed. As shown in Fig. 3a, the fluorescence intensity at 616 nm increased with the addition of doxycycline, conversely, the fluorescence intensity at 436 nm decreased. Meanwhile, the value of $\ln(F_{616 \text{ nm}}/F_{436 \text{ nm}})$ had a good linear relationship with doxycycline concentration with an adjusted R^2 of 0.99401 (Fig. 3b). The LOD ($3\sigma/s$) was calculated to be $0.36 \mu\text{M}$, which was better than the other methods stated in the literature (Table S1†). Fig. 3c shows the subtle colour changes of the test paper under a 365 nm UV lamp upon doxycycline addition; the colour changed from blue-purple to pink-red as the concentration of doxycycline increased (0 – $200 \mu\text{M}$). Different categories of antibiotics were selected to investigate selectivity under identical conditions. $\ln(F_{616 \text{ nm}}/F_{436 \text{ nm}})$ showed no obvious change for other antibiotics, which demonstrated that doxycycline could be identified readily from other antibiotics (Fig. 4a and b). With respect to anti-interference ability, the $\ln(F_{616 \text{ nm}}/F_{436 \text{ nm}})$ of the composite showed no obvious change in the presence of other interfering substances (Fig. 4c). Three other tetracycline antibiotics were also studied (Fig. S4†); although they had similar structures, the value of $\ln(F_{616 \text{ nm}}/F_{436 \text{ nm}})$ of doxycycline was more obvious.

Electronic absorption spectra (UV-vis) were used to analyse the possible sensing mechanism of doxycycline. As depicted in Fig. 4d, the overlaps between the absorption peak of doxycycline and emission peak of the CDs indicated that energy transfer might occur, resulting in the fluorescence quenching of CDs on the basis of the Förster resonance energy transfer mechanism. Inspired by the studies carried out by Jiang *et al.*,^{37–39} doxycycline was an ideal ligand for Eu^{3+} that could form a doxycycline– Eu^{3+} complex and enhance the fluorescence of Eu^{3+} . Hence, the integration of doxycycline and Eu^{3+} may cause the fluorescence enhancement of the emission peak at 616 nm. Accordingly, the spectrum showed a fall and rise simultaneously in the presence of doxycycline.

MnO₄[−] detection and possible sensing mechanism

The effects of different anions (MnO_4^- , NO_3^- , CH_3COO^- , CO_3^{2-} , ClO_3^- , $\text{C}_2\text{O}_4^{2-}$, SO_3^{2-} , HCO_3^- , NO_2^- , H_2PO_2^-) on the composites were studied. The fluorescence intensity at 616 nm was quenched upon addition of MnO_4^- (Fig. 5a). An experiment using original MOF(Eu) without CDs was also investigated. As shown in Fig. S5,† the fluorescence of original MOF(Eu) showed no obvious change as the MnO_4^- concentration increased. Conversely, the 616 nm fluorescence of the composite was quenched by $\approx 50\%$ at the same concentration of MnO_4^- and the corresponding quenching constant was calculated: it was $3.641 \times 10^4 \text{ L mol}^{-1}$ (Fig. 5b). This observation suggested that CDs probably accelerated this process, and lowered the LoD (from $12.43 \mu\text{M}$ to $0.68 \mu\text{M}$). This sensor had good selectivity towards MnO_4^- and the



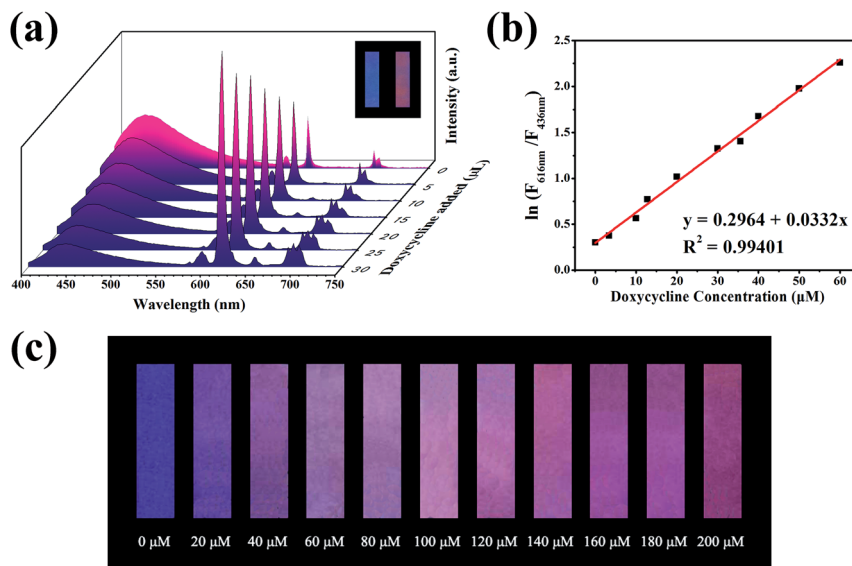


Fig. 3 (a) Fluorescence spectra of composites with different concentrations of doxycycline under 298 nm excitation. (b) The linear relationship of $\ln(F_{616\text{ nm}}/F_{436\text{ nm}})$ and concentration of doxycycline. (c) The colour change of "test paper" as the concentration of doxycycline increased under 365 nm UV irradiation.

quenching process did not suffer interference by other anions (Fig. 5c and d).

The colour of KMnO_4 lightened after detection, indicating that some reactions may have occurred. Characterization methods (SEM, XPS, FT-IR spectroscopy, XRD, UV-vis spectroscopy) were employed to study the possible sensing mechanism. On account of the contrast experiment of MOF(Eu) mentioned above, we suspected that CDs may react with MnO_4^- , resulting

in acceleration of the quenching progress. The UV spectra of the CDs@MOF(Eu)- KMnO_4 hybrid system changed in comparison with previous spectra (Fig. S6[†]), which suggested that a ground state complex could have been generated.⁴⁰ SEM images and the corresponding element mapping images are given in Fig. S7.[†] It can be seen clearly that the manganese element is distributed uniformly on the surface of the composite, thereby showing the formation of manganese

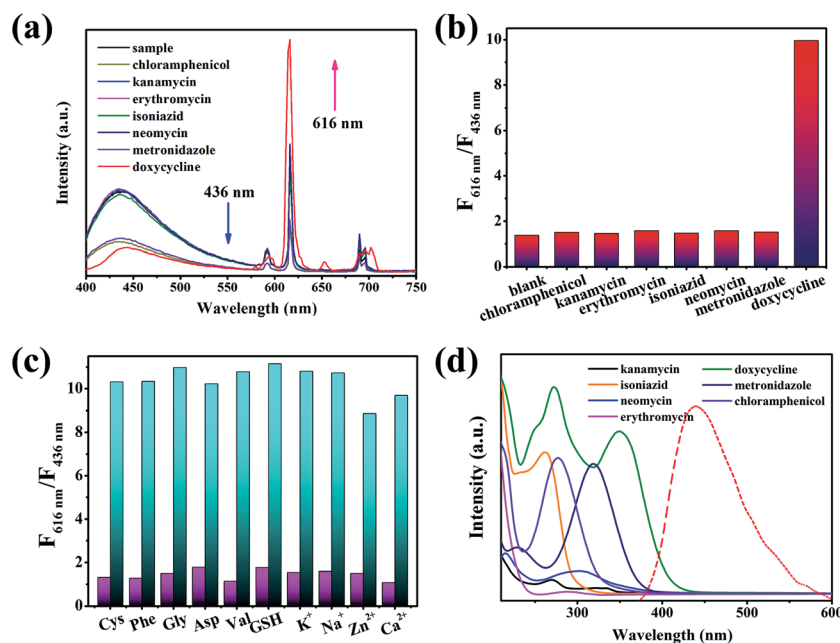


Fig. 4 (a) Fluorescence spectra of composites added with different antibiotics. (b) Selectivity of doxycycline based on the ratio of $F_{616\text{ nm}}$ and $F_{436\text{ nm}}$. (c) Value of $F_{616\text{ nm}}/F_{436\text{ nm}}$ in the presence of other interfering substances (excited at 298 nm). (d) Absorption spectrum of different antibiotics and emission spectra of carbon dots.



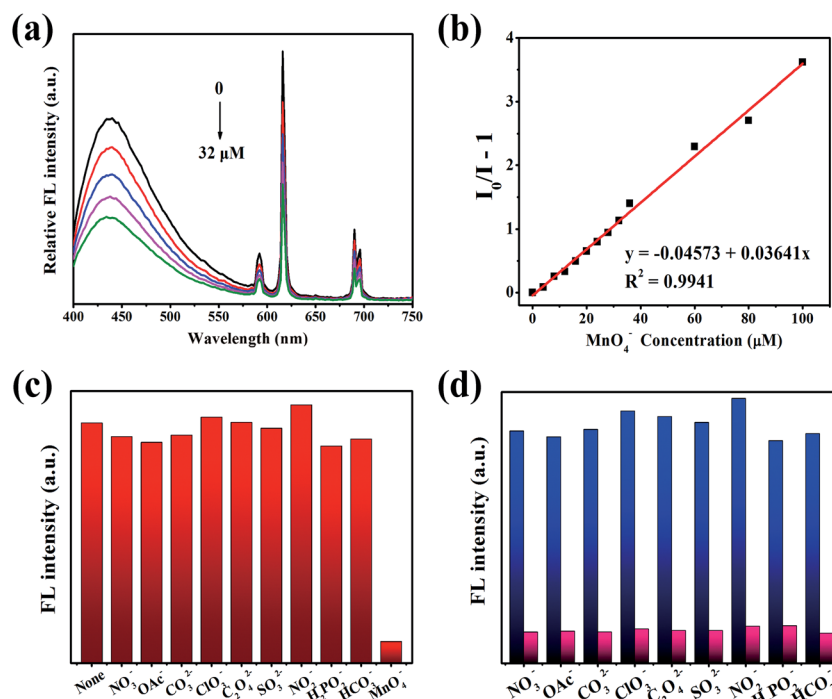


Fig. 5 (a) Emission spectra of CDs@MOF(Eu) with added MnO₄⁻ (0–32 μM). (b) The linear relationship between MnO₄⁻ concentration and I₀/I - 1 based on the 616 nm emission peak. (c) The selectivity of MnO₄⁻ with that of other anions. (d) Fluorescence intensity at 616 nm of composites with added MnO₄⁻ (0.2 mM) in the interference of other anions (0.2 mM).

compounds. XPS was done to judge the valence of the manganese element in compounds. As shown in Fig. S8a,[†] the peaks of C1s (282 eV), O1s (529 eV), N1s (397 eV), Eu4d (134 eV) and Mn2p (639 eV) appeared in the broad spectrum, demonstrating the main elements and contents in the CDs@MOF(Eu)-KMnO₄ hybrid system. The content of manganese was relatively small, so it was not obviously displayed in the broad spectrum. In the C1s spectrum, the three peaks at 284.5, 286, and 288.2 eV were assigned to C-C or C=C, C-N, N-C=O peaks (Fig. S8b[†]). The two peaks at 531.2 and 532.5 eV in the O1s spectrum corresponded to O=C and O-C peaks (Fig. S8c[†]). The two peaks at 399.5 and 401.4 eV in the N1s spectrum belonged to C-N-C and N-H peaks (Fig. S8d[†]). In the Mn2p spectrum (Fig. S8e[†]), the two peaks at 641.4 and 653.1 eV were attributed to Mn2p_{3/2} and Mn2p_{1/2}, respectively. Their spin energy separation was 11.7 eV, illustrating that MnO₂ could exist in the hybrid system, a result that was in good agreement with the literature.^{41,42} Hence, we proposed two possible explanations for the sensing mechanism. First, the quenching of 616 nm fluorescence could be explained in terms of energy. As shown in Fig. S9,[†] an overlap was present between the excitation of the composite and the absorption of MnO₄⁻. The competition in energy of excitation and adsorption between MOF(Eu) and MnO₄⁻ could lead to fluorescence quenching,⁴³ and CDs may accelerate the progress of energy competition. The XRD patterns in Fig. S10[†] suggested another possible sensing mechanism. MnO₄⁻ may have weak interactions with the framework of MOF(Eu). Meanwhile, MnO₄⁻ reacted with some CDs initially, resulting in the formation of

MnO₂, which could accelerate the interaction between MnO₄⁻ and MOF(Eu). Otherwise, this sensor was comparable with other MOFs for sensing MnO₄⁻, and the addition of CDs made the performance of MOF(Eu) almost identical or even better than that of similar materials (Table S2[†]).

Applications in real samples

Doxycycline is often used to treat people or animals. Hence, TBS buffer solution was used to simulate a biological environment to detect doxycycline. MnO₄⁻ is known to be a common pollutant, so accurate quantitative determination of MnO₄⁻ in tap water is very important. As shown in Tables 1 and 2, the relative standard deviation of the results was <3%, indicating that these data were reliable. The measured concentration was calculated according to the previous linear equations (C: mol L⁻¹):

$$\ln(F_{616 \text{ nm}}/F_{436 \text{ nm}}) = 0.2964 + 0.0332 \times C_{\text{dox}} \quad (1)$$

Table 1 Analytical results of doxycycline detection in a simulated biological system (TBS)

Sample	Spiked concentration (μM)	Measure (μM)	Recovery (%)	RSD (%), n = 3
1	15.8	14.85	93.95	0.70
2	27	27.08	100.31	1.41
3	35.6	36.41	102.28	1.29
4	44.2	44.36	100.37	0.66
5	56.6	54.44	96.19	0.54



Table 2 Analytical results of MnO_4^- detection in tap water

Sample	Spiked concentration (μM)	Measure (μM)	Recovery (%)	RSD (%), $n = 3$
1	10.4	10.10	97.15	2.48
2	17.6	17.71	100.61	0.92
3	29.4	29.91	101.72	2.61
4	46.2	46.59	100.85	0.60
5	75	74.91	99.88	1.54

$$F_0/F - 1 = -0.04573 + 0.03641 \times C_{\text{MnO}_4^-} \quad (2)$$

Good detection recoveries of doxycycline and MnO_4^- were acquired (93.95–102.28%). Such accuracy and reliability further confirmed that this composite was a promising and efficient sensor for the detection of doxycycline and MnO_4^- in real samples.

Conclusions

We synthesized a hybrid material which had dual-emission fluorescence for sensing doxycycline and MnO_4^- . This work presents, for the first time, a rapid, low-cost way to detect doxycycline using test paper. The latter is more convenient and faster than other methods mentioned in the literature and can be observed by the naked eyes. This sensor can also detect MnO_4^- with a lower LOD among other anions compared with the original MOF(Eu). The proposed hybrid material in this work could have wider applications in the field of sensors.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was supported by the National Natural Science Foundation of China (21371103).

Notes and references

- P. J. Ramesh, K. Basavaiah and N. Rajendraprasad, *Acta Pharm.*, 2010, **60**, 445–454.
- M. Y. Xue, L. L. Zhang, Z. H. Zhan, M. B. Zou, Y. Huang and S. L. Zhao, *Talanta*, 2016, **150**, 324–330.
- K. Smith and J. J. Leyden, *Clin. Ther.*, 2015, **27**, 1329–1342.
- D. Vega, L. Agüí, A. González-Cortés, P. Yáñez-Sedeño and J. M. Pingarrón, *Anal. Bioanal. Chem.*, 2007, **389**, 951–958.
- S. H. Li, J. P. Li, Q. Y. Lin and X. P. Wei, *Analyst*, 2015, **140**, 4702–4707.
- Y. S. Kim, J. H. Kim, I. A. Kim, S. J. Lee, J. S. Jung and M. B. Gu, *Biosens. Bioelectron.*, 2010, **26**, 1644–1649.
- Y. L. Tang, J. C. Zhang, J. H. Liu, I. Gapparov, S. Wang, Y. W. Dong, H. J. Su and T. W. Tan, *Anal. Methods*, 2017, **9**, 1133–1140.
- M. Granados, M. Encabo, R. Compano and M. D. Prat, *Chromatographia*, 2005, **61**, 471–477.
- H. T. Lu, Y. Jiang, H. B. Li, F. Chen and M. H. Wong, *Chromatographia*, 2004, **60**, 113–116.
- T. Tylová, J. Olšovská, P. Novák and M. Flieger, *Chemosphere*, 2010, **78**, 353–359.
- T. Charoenraks, S. Palaharn, K. Grudpan, W. Siangproh and O. Chailapakul, *Talanta*, 2004, **64**, 1247–1252.
- P. Masawat, S. Liawruangrath and S. Upalee, *Maejo Int. J. Sci. Technol.*, 2008, **2**, 201–209.
- W. L. Shi, F. Guo, M. M. Han, S. L. Yuan, W. S. Guan, H. Li, H. Huang, Y. Liu and Z. H. Kang, *J. Mater. Chem. B*, 2017, **5**, 3293–3299.
- J. P. Song, J. Li, Z. Y. Guo, W. Liu, Q. Ma, F. Feng and C. Dong, *RSC Adv.*, 2017, **7**, 12827–12834.
- J. Tashkhourian, G. Absalan, M. Jafari and S. Zare, *Spectrochim. Acta, Part A*, 2016, **152**, 119–125.
- J. Y. Liu, G. C. Lv, W. L. Gu, Z. H. Li, A. W. Tang and L. F. Mei, *J. Mater. Chem. C*, 2017, **5**, 5024–5030.
- R. Zhou, Q. Zhao, K. K. Liu, Y. J. Lu, L. Donga and C. X. Shan, *J. Mater. Chem. C*, 2017, **5**, 1685–1691.
- J. Tao, P. Zhao and Q. Zeng, *J. Mater. Chem. B*, 2016, **4**, 4258–4262.
- C. Li, W. J. Liu, Y. J. Ren, X. B. Sun, W. Pan and J. P. Wang, *Sens. Actuators, B*, 2017, **240**, 941–948.
- J. Zong, X. L. Yang, A. Trinchì, S. Hardin, I. Cole, Y. H. Zhu, C. Z. Li, T. Muster and G. Wei, *Biosens. Bioelectron.*, 2014, **51**, 330–335.
- L. B. Li, B. Yu and T. Y. You, *Biosens. Bioelectron.*, 2015, **74**, 263–269.
- S. Chandra, D. Laha, A. Pramanik, A. R. Chowdhuri, P. Karmakar and S. K. Sahu, *Luminescence*, 2016, **31**, 81–87.
- W. J. Lu, Y. Li, R. J. Li, S. M. Shuang, C. Dong and Z. W. Cai, *ACS Appl. Mater. Interfaces*, 2016, **8**, 12976–12984.
- A. Cayuela, M. L. Soriano and M. Valcárcel, *Anal. Chim. Acta*, 2013, **804**, 246–251.
- M. Zheng, Z. G. Xie, D. Qu, D. Li, P. Du, X. B. Jing and Z. C. Sun, *ACS Appl. Mater. Interfaces*, 2013, **5**, 13242–13247.
- R. K. Das and S. Mohapatra, *J. Mater. Chem. B*, 2017, **5**, 2190–2197.
- F. Qu, Z. Sun, D. Y. Liu, X. E. Zhao and J. M. You, *Microchim. Acta*, 2016, **183**, 2547–2553.
- Y. Zhou, H. H. Chen and B. Yan, *J. Mater. Chem. A*, 2014, **2**, 13691–13697.
- X. Y. Xu and B. Yan, *ACS Appl. Mater. Interfaces*, 2015, **7**, 721–729.
- J. N. Hao and B. Yan, *J. Mater. Chem. A*, 2014, **2**, 18018–18025.
- Y. Q. Dong, J. H. Cai, Q. Q. Fang, X. You and Y. W. Chi, *Anal. Chem.*, 2016, **88**, 1748–1752.
- X. Y. Xu and B. Yan, *J. Mater. Chem. C*, 2016, **4**, 1543–1549.
- J. X. Wu and B. Yan, *J. Colloid Interface Sci.*, 2017, **504**, 197–205.
- B. Ding, S. X. Liu, Y. Cheng, C. Guo, X. X. Wu, J. H. Guo, Y. Y. Liu and Y. Li, *Inorg. Chem.*, 2016, **55**, 4391–4402.
- M. Bagheri, M. Y. Masoomi and A. Morsali, *Sens. Actuators, B*, 2017, **243**, 353–360.



- 36 B. L. Chen, Y. Yang, F. Zapata, G. Lin, G. D. Qian and E. B. Lobkovsky, *Adv. Mater.*, 2007, **19**, 1693–1696.
- 37 Q. Peng, F. J. Hou and C. Q. Jiang, *Spectrochim. Acta, Part A*, 2006, **65**, 62–66.
- 38 J. Li, J. K. Liu, X. J. Zhu, Q. Peng and C. Q. Jiang, *J. Lumin.*, 2005, **113**, 305–313.
- 39 C. Q. Jiang and N. Zhang, *J. Pharm. Biomed. Anal.*, 2004, **35**, 1301–1306.
- 40 F. L. Zu, F. Y. Yan, Z. J. Bai, J. X. Xu, Y. Y. Wang, Y. C. Huang and X. G. Zhou, *Microchim. Acta*, 2017, **184**, 1899–1914.
- 41 L. Li, A. R. O. Raji and J. M. Tour, *Adv. Mater.*, 2013, **25**, 6298–6302.
- 42 L. Zhang, H. D. Liu, H. B. Ruan, Y. Y. Su, R. Hu, L. L. Tian, Z. L. Hu and J. Li, *Int. J. Electrochem. Sci.*, 2016, **11**, 10815–10826.
- 43 M. Chen, W. M. Xu, J. Y. Tian, H. Cui, J. X. Zhang, C. S. Liu and M. Du, *J. Mater. Chem. C*, 2017, **5**, 2015–2021.

