1 Introduction

The CXCR4 chemokine receptor is a seven transmembrane helix protein, member of the G-protein-coupled receptor (GPCR) superfamily, whose own natural ligand is SDF-1α or CXCL12.1–4 The axis CXCR4/CXCL12 is implicated in the regulation of numerous biological processes in the human body that result in a variety of normal physiological responses, such as chemotaxis, cell survival and proliferation, intracellular calcium flux, and gene transcription. CXCR4 is also involved in many downstream pathways, such as inflammation,5 mobilisation of stem cells,6 and development of cancer cells metastasis.7,8 Furthermore, CXCR4 is a co-receptor for T-cell tropic strains of human immunodeficiency virus-1 (HIV-1) and allows fusion and entry of the virus into human white blood cells. Targeting specifically these receptors represents therefore a huge potential for many applications, such as characterization of a pathology, drug development, staging and restaging, following a therapy response, delineation and quantification of tumors in oncology. Nowadays, only one bis-cyclam derivative, AMD3100,9,10 is clinically used in oncology, in combination with GCSF (granulocyte colony stimulating factor) to mobilize haematopoietic stem cells and allow harvesting.11,12 Several AMD like compounds, differing by the size of the macrocycle and the nature of the linker, have been synthesized and studied,13,14,15 highlighting the tremendous importance of these parameters on the affinity of the bismacrocyclic compound for its target. Thus, it is of major interest to devise new synthetic routes for the construction of bismacrocyclic scaffolds analog to AMD3100, which could present better therapeutic properties.16,17

In this study we report the synthesis of a series of new bismacrocyclic compounds, structural analogues of biscyclam AMD3100, in which the two macrocycles are linked together through carbon atoms of the cycles. Several representatives of this new class of bicyclic derivatives were prepared by reacting C-aminomethyl-13aneN4 with aromatic dialdehydes. Preliminary in vitro studies were performed to evaluate the affinity of these compounds towards the co-receptor CXCR4.

2 Results and discussion

2.1 Synthesis

In previous work we have demonstrated that the primary amine function of C-aminomethyl-13aneN4 (I) readily reacts with one equivalent of aromatic aldehydes to form cyclic aminal derivatives involving an adjacent nitrogen atom of the cycle. Reduction of these aminal compounds with NaBH₄ gave access to the
corresponding macrocycles selectively alkylated on the pendant amino methyl group.\textsuperscript{20} It might be expected that such peculiar reactivity can be exploited to synthesize bismacrocycles by reacting the C-aminomethyl macrocycle (1) with dialdehydes. Indeed, by using 0.5 equivalent of various dialdehydes (isophthalaldehyde, terephthalaldehyde, pyridine-2,6-dicarbaldehyde, 2,2'-oxydibenzaldehyde), the corresponding bismacroyclic bisaminal derivatives were obtained in good yields (up to 82\%) (Scheme 1).

The reduction of these compounds with NaBH\textsubscript{4} followed by acidic treatment gave the hydrochloride salts of the aimed bistetraazacycloalkanes 5–8 as white powders.

The NMR spectra of the isolated bismacroyclic bisaminal derivatives 2–4 are very complicated due to the formation of a mixture of different isomers. Monocrystals of pure 2a were prepared after slow crystallization of 2 in diethyl ether (Fig. 1, left). The structure of compound 4a was also determined by X-ray diffraction (Fig. 1, right). An exhaustive description of each compound is given in the ESI.\textsuperscript{†}

Both compounds crystallize identically, they present the same symmetry (monoclinic) and the same conformation with a water molecule in the asymmetric unit. A RMSD\textsubscript{max} of 0.4 Å was found by superimposing both structures. Cremer–Pople parameters,\textsuperscript{21} which describe the conformation of a puckered ring in a quantitative mathematically well-defined manner, indicate that the five membered aminal rings have a slightly distorted chair geometry [compound 2a: N1–C9–C10–N5–C11, Q = 0.360(3) Å, \( \phi = 312.4(4) \)°, N6–C18–C19–C20, Q = 0.345(2) Å, \( \phi = 226.6(4) \)°; compound 4a: N4–C7–C10–N5–C11, Q = 0.348(4) Å, \( \phi = 314.1(7) \)°, N7, N7–C17–N8–C19–C18, Q = 0.363(4) Å, \( \phi = 24.5(6) \)°]. These five membered rings impose the half conformation of a part of the polyazacycloalkane. The second shape of the molecule is governed by intra- and intermolecular hydrogen bonds (see ESI\textsuperscript{†}).

The characterization of the three different isomers was not further investigated since they all give the same compound after opening the aminal bridges by NaBH\textsubscript{4}. The new bismacrocycles 5–8 were fully characterized using mass spectrometry, \(^1\)H and \(^{13}\)C NMR and elemental analysis. It is noteworthy that the number of signals in the \(^{13}\)C NMR spectra of these compounds is low. This feature can of course be explained by an expected symmetry of the molecule. However, the presence of two identical chiral carbon atoms in these bismacrocycles should give rise to the formation of three stereoisomers, i.e. a meso compound and a couple of enantiomers since the starting C-aminomethyl macrocycle 1 was used as a racemic mixture. The simplicity of the NMR spectra seems to prove that the reaction is stereospecific, leading to either the meso compound or a racemic mixture of the two enantiomers.
The expected bisaminal bismacrocycle was also not obtained when reacting C-aminomethyl-13aneN4 (1) with 2,9-dicarboxaldehyde-1,10-phenanthroline (2,9-DCP). Again, this could be attributed to intra- and/or intermolecular reaction of the aldehyde with different amine functions, giving side products. These results prompted us to devise another route for the preparation of the desired bismacrocycles. In order to avoid the intramolecular reaction of the second aldehyde with secondary amine functions, we decided to use as starting material the bisaminal protected macrocycle 10 which is actually the precursor of C-aminomethyl macrocycle 1 which is actually the precursor of C-aminomethyl macrocycle 1

In this case the cyclic aminal cannot be formed and the reaction with 2,9-dicarboxaldehyde-1,10-phenanthroline followed by reduction with NaBH₄ gives the protected bismacrocycle 11. This compound was finally treated by HCl 37% in ethanol to yield the aimed bis-macrocycle 12.

2.2 Biological studies

Cellular binding assays of new AMD3100 analogues to the CXCR4 receptor were investigated in a competition assay with an anti-CXCR4 monoclonal antibody (mAb) (12G5 conjugated to phycoerythrin). Jurkat cells, a T-lymphocyte cell line which expresses high level of CXCR-4 expression, were used and binding assays of new compounds 2(a–c), 3(a–c), 4(a–c), 5, 6, 7, and AMD3100 (as reference) to the cells were analysed by flow cytometry. On the whole, synthesized compounds were less efficient than AMD3100, which maximum inhibition concentration was 1 μg mL⁻¹. Compounds 2(a–c), 3(a–c), 4(a–c) and 7, didn’t show any inhibition potency. However, at 5 μg mL⁻¹, the analogs 5 and 6 could inhibit the binding of the mAb (41.16% ± 8.1 and 54.83% ± 2.5, respectively) on Jurkat T cells (Fig. 3). This result was confirmed using DAUDI cells (data not shown). These preliminary results confirm the importance of both the nature of the macrocycle and of the bridge between the two.
macrocycles, and that small structural modifications can strongly reduce the affinity of the AMD analogues towards the CXCR4 receptor. Indeed, previous mutational analysis of the CXCR4 binding site for AMD3100 has identified amino acid Asp\textsuperscript{177} and Asp\textsuperscript{262} as key residues involved in the binding to the CXCR4 receptor\textsuperscript{22}. The carboxylate groups of the receptor residues form three (one strong, one intermediate and one weak) hydrogen bonds with protonated cyclam of AMD3100.

Changing the geometry of the macrocycle and the spacer can really influence the binding of the compounds, yielding in a decrease of the resulting interactions for the receptor. Therefore, not surprisingly, 5 and 6, which are structurally more resembling to AMD3100, present the highest affinity for the CXCR4 receptor.

Since the interaction between SDF-1 and CXCR4 has been shown to direct tumor cells to organ sites with high levels of SDF-1 expression, the effect of AMD3100 and its analogs on inhibiting CXCL12-induced migration of Jurkat T cells was estimated by a classical chemotaxis assay. As expected, Jurkat T cells migrated in response to CXCL12, even if only 20% migrated because of a low % of Jurkat T cells bound the chemokine. After AMD3100 treatment (10 μg mL\textsuperscript{-1}), chemotactic activity of Jurkat cells was totally inhibited. However, this property of AMD3100 to inhibit CXCL12-induced chemotaxis through CXCR4 was compromised by structural modifications as none of the analogs could strongly SDF-1-induced migration of Jurkat T cells (Fig. 4).

3 Conclusion

The work described herein reports the synthesis of a new family of bis-tetraazacycloalkanes formed by two same 13aneN4 units connected by an aromatic spacer. The reaction of two equivalents of C-aminomethyl-13aneN4 with commercial dialdehydes in ethanol followed by reduction with NaBH\textsubscript{4} resulted in the formation of bismacroyclic compounds, structural analogs of AMD3100. In some cases, with ortho-phthalaldehyde or 2,9 DCP, unexpected lactam or side products were obtained. To prepare the aimed bismacroycles, the protected macrocycle has been used instead of C-aminomethyl-13aneN4 in order to limit the formation of undesired compounds. Affinity assays with the CXCR4 receptor have been performed and showed an effective binding to the receptor for compounds 5 and 6. Further work is ongoing for the preparation of metal complexes of these bismacroycles. Indeed, previous studies have shown that metal ions incorporation could increase the affinity of bismacroycles such as AMD3100 or analogs for its receptor\textsuperscript{23–25}

4 Experimental section

4.1 Instruments

All analyses were performed at the “Plateforme d’Analyses Chimiques et de Synthèse Moléculaire de l’Université de Bourgogne”. High resolution and accurate mass measurements were carried out using a Bruker MALDI-TOF, microTOF-Q\textsuperscript{TM} ESI-TOF (Electro Spray Ionization-Time of Flight) and a Thermo Scientific\textsuperscript{®} LTQ Orbitrap mass spectrometer. \textsuperscript{1}H and \textsuperscript{13}C NMR spectra were recorded on a Bruker 300 and 500 NMR spectrometer. Chemical shifts were reported in parts per million using tetramethylsilane (TMS) as the internal standard. The following abbreviations are used; s: singlet, d: doublet, t: triplet, q: quartet, m: multiplet, b: broad.

X-ray equipment and refinement are detailed in ESL\textsuperscript{†} Data CCDC 1476681, 1476682 and 1476683 contain the supplementary crystallographic data for compound 2a, 4a and 9 respectively.

4.2 Biologicals

Chemokine and antibodies. Recombinant human SDF-1 (CXCL12) was purchased from PeproTech (Neuilly-Sur-Seine, France), Fluorokine\textsuperscript{®} biotinylated human SDF-1, monoclonal anti-human CXCR4-phycoerythrin (PE) antibody (clone 12G5) and mouse IgG2A isotype control-PE were purchased from R&D Systems.

Cell lines. Daudi or Jurkat cell line (clone E6-1) was obtained from the American Type Culture Collection (ATCC) (Molsheim, France). Cells were maintained in RPMI 1640 supplemented with 10% FBS, 2 mM L-glutamine, 100 μg mL\textsuperscript{-1} penicillin and 100 μg mL\textsuperscript{-1} streptomycin.

Flow cytometry. Daudi or Jurkat cells (105 cells in 25 μL) were incubated with or without AMD3100 or its analog molecules at different concentrations for 15 min on ice. 12G5 mAb (5 μg mL\textsuperscript{-1}) was then added for 30 min on ice. Cells were washed twice with buffer and acquired on a flow cytometer (LSR II, Becton Dickinson, San Jose, CA, USA). To determine the presence of SDF-1 on Jurkat cells, Fluorokine cells, Fluorokine\textsuperscript{®} kit was used. Briefly, 10 μL of biotinylated human SDF-1 was added to 25 μL of cells (4 × 10\textsuperscript{5} per mL) for 30 min on ice. 10 μL of avidin-FITC was then added and cells were further incubated for 30 min on ice. Cells were washed twice with Cell Wash Buffer and re-suspended for flow cytometry acquisition.

Chemotaxis. Jurkat cells were serum-starved in migration medium (RPMI 1640 containing 1% BSA, 10 mM HEPES buffer, pH 6.9) for 4 h. Migration assays were performed in duplicate in transwell plates of 6.5 mm diameter, with 5.0 μm pore polycarbonate membrane (VWR international, France). A volume of 0.6 mL migration medium, with or without 100 ng mL\textsuperscript{-1} of recombinant human SDF-1, was added to the lower chamber of

Fig. 4 Inhibition of SDF-1 induced migration of Jurkat cells by AMD3100 and its analogs. Results shown are average of duplicate ± SE. This experiment is representative of two different experiments.
and compound as a white powder (\(+H\)). Elemental analysis: C 28H52N10, H2O calculated: C 50.3, 50.5, 50.6, 50.8, 51.0, 51.1, 51.2, 51.3, 51.5, 51.6, 51.7, 51.8, 51.9, 52.0, 52.1, 52.3, 52.4, 52.5, 52.6, 52.7, 52.8, 52.9, 53.0, 53.1, 53.2, 53.3, 53.4, 53.6. 1H NMR of mixture of (1H) NMR (75 MHz, CDCl3, 300 K) \(\delta\) ppm: 1.05 (bs, 1H), 2.08 (m, 5H), 2.70–4.01 (m, 4H), 4.43 (bs, 5H), 7.40 (m, 2H), 7.83 (m, 1H). 13C{1H} NMR (150 MHz, D2O, 300 K) \(\delta\) ppm: 22.3, 41.2, 43.1, 44.2, 44.3, 46.0, 47.6, 48.4, 51.6, 51.7, 123.4, 139.5, 150.4. ESI-TOF (m/z) calculated: 576.732, found: 576.738 [M + 2H]+.

2,2′-(Oxybis(2,1-phenylene))bis(N\((1,4,7,10\text{-tetrachlorobenzocyclododec-5-yl})\) methyl)ethanamine hydrochloride salt (7). This compound was synthesized according to the procedure used for the synthesis of 5 starting from compound (a-c) (0.37 g, 0.70 mmol), and NaBH4 (0.26 g, 7.0 mmol). Compound 7 was obtained as a white powder after protonation by HCl 37% \((m = 0.5 g, 0.56 mmol, yield = 80\%)\). 1H NMR (600 MHz, D2O, 300 K) \(\delta\) ppm: 1.23–2.02 (m, 8H), 2.23–3.35 (m, 38H), 4.33 (m, 2H), 7.21 (m, 2H), 7.60 (m, 1H). ESI-TOF (m/z) calculated: 530.441, found: 530.442 [M]+.
solid (diethyl ether (20 mL). Compound solution was obtained as an orange powder (yield = 55%). 1H NMR (300 MHz, CDCl3, 300 K) δ (ppm): 2.16 (m, 1H), 2.10–2.84 (m, 16H, 3.18 (m, 4H), 1.13 (m, 6H), 2.16 (m, 1H), 3.49 (m, 1H), 4.54 (m, 1H), 7.46 (m, 3H), 7.81 (m, 1H). 13C{1H} NMR (75 MHz, CDCl3, 300 K) δ (ppm): 28.0, 45.7, 47.7, 48.2, 48.9, 50.9, 51.1, 51.9, 53.3, 77.2, 124.2, 124.9, 129.3, 130.9, 133.5, 141.3, 166.7. HRMS-MALDI-TOF (m/z): calculated: 329.221; found: 329.701 [M]+. Elemental analysis: C18H27N5O.

Bisaminal-butanedione-N,N’-(1,10-phenanthroline-2,9-diyl)bis(methylene)bis(1-(4,7,10-tetraazacyclodecan-5-yl) methanamine) (11). 1.10 g of 1,10-phenanthroline-2,9-dicarbaldehyde (4.7 mmol) were added to a solution of 10 (2.47 g, 9.4 mmol) in ethanol (100 mL). The mixture was stirred at room temperature during 2 hours. Then the solvent was removed and the residual oil was purified by recrystallization in hexane/CH2Cl2 (eluant (98/2): CH2Cl2/MeOH). A compound 9 was obtained as a brown solid (yield = 8.3 Hz, 2H). 13C{1H} NMR (75 MHz, CDCl3, 300 K) δ (ppm): 28.9, 46.1, 47.7, 48.7, 49.1, 49.2, 49.7, 49.8, 51.2, 51.9, 57.0, 117.8, 123.4, 128.5, 130.4, 131.0, 155.1. HRMS-MALDI-TOF (m/z): calculated: 624.495, found: 625.319 [M + H]+.

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