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# Hydrolysis of *cis*- and *trans*platin: structure and reactivity of the aqua complexes in a solvent free environment†

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Singly aquated and diaquated species are key intermediates in the mechanism responsible for the antitumor activity of cisplatin. Aqua complexes  $[\text{PtX}(\text{NH}_3)_2(\text{H}_2\text{O})]^+$  ( $\text{X} = \text{Cl}, \text{OH}$ ), obtained in water by hydrolysis of cisplatin and of the inactive isomer transplatin, are transferred into the gas-phase by electrospray ionization. The so-formed ions, *cis*- and *trans*- $[\text{PtX}(\text{NH}_3)_2(\text{H}_2\text{O})]^+$ , have been allowed to react with selected ligands, representing platination targets in the biological environment. The reaction kinetics monitored in the gas-phase show consistently higher reactivity for the chloro complexes,  $[\text{PtCl}(\text{NH}_3)_2(\text{H}_2\text{O})]^+$ , with respect to the hydroxo counterparts,  $[\text{Pt}(\text{OH})(\text{NH}_3)_2(\text{H}_2\text{O})]^+$ . The latter species, both *cis*- and *trans*-isomers, have been assayed by IRMPD spectroscopy in the NH/OH stretching region and their vibrational and geometric features are compared with the ones pertaining to the already described chloro complexes, *cis*- and *trans*- $[\text{PtCl}(\text{NH}_3)_2(\text{H}_2\text{O})]^+$ .

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## 1. Introduction

Cisplatin, *cis*- $\text{PtCl}_2(\text{NH}_3)_2$ , is a leading antitumor drug of impressive efficacy, used both alone and in combination with other drugs for the treatment of various kinds of solid tumors.<sup>1–5</sup> The anticancer activity of cisplatin is due to a reaction with cellular DNA, leading to the formation of intrastrand cross-links with a bridging  $\text{Pt}(\text{NH}_3)_2$  unit. However, DNA bases do not replace the chloro ligand from platinum(II) directly but predominantly *via* a solvent assisted pathway. Cisplatin aquation (replacement of  $\text{Cl}^-$  with  $\text{H}_2\text{O}$ ) yields the singly aquated species, *cis*- $[\text{PtCl}(\text{NH}_3)_2(\text{H}_2\text{O})]^+$ . Also the diaquated *cis*- $[\text{Pt}(\text{NH}_3)_2(\text{H}_2\text{O})_2]^{2+}$  or its deprotonated form are suggested to be important contributors to DNA platination.<sup>6,7</sup> The positive charge on the platinum complex can attract the negatively charged nuclear DNA, thus favoring the formation of DNA lesions.<sup>1–5</sup> Interestingly, the charged aqua complexes are directly amenable to mass spectrometric investigation paving the way to an evaluation of their intrinsic properties and

hopefully enlightening useful issues for Pt-drug design.<sup>8</sup> In this context we have recently undertaken a study of the *cis*- $[\text{PtCl}(\text{NH}_3)_2(\text{H}_2\text{O})]^+$  complex from the primary aquation reaction.<sup>9</sup> The vibrational signatures of the isolated species have been assayed by infrared multiple photon dissociation (IRMPD) spectroscopy,<sup>10–15</sup> in parallel with a characterization of the *trans* isomer, *trans*- $[\text{PtCl}(\text{NH}_3)_2(\text{H}_2\text{O})]^+$ .<sup>9</sup> The experimental features have been interpreted by comparison with the calculated IR spectra for the computed stable structures, as typically performed.<sup>10–15</sup> Using this approach, the binding motifs in the naked adducts from cisplatin with nucleobases (adenine and guanine) have been shown to involve the N7 position of guanine and N1/N3 positions of adenine, with N7 attack to adenine notably missing.<sup>16</sup> Cisplatin binding to a DNA building block (2'-deoxyguanosine-5'-monophosphate) is found to lead to macrochelate species isolated in the gas-phase where IRMPD spectroscopy has shown the metal to be ligated to both N7 of guanine and a phosphate oxygen atom.<sup>17</sup> Gas-phase vibrational spectroscopy and ion chemistry have allowed to thoroughly characterize the ligand substitution reaction replacing the aqua ligand of *cis*- $[\text{PtCl}(\text{NH}_3)_2(\text{H}_2\text{O})]^+$  with simple molecules representing ubiquitous active species in the biological environment faced by the cisplatin drug.<sup>18</sup> A simple glycine-linked cisplatin,  $[(\text{Gly-H})\text{PtCl}_2]^-$ , has provided a case study for IRMPD experiments combined with a survey of hybrid theoretical approaches for determining structures and IR spectra.<sup>19</sup>

In this contribution we have investigated the aqua-hydroxo *cis*- and *trans*- $[\text{Pt}(\text{OH})(\text{NH}_3)_2(\text{H}_2\text{O})]^+$  complexes with regard to both structure and reactivity, in relation to the aqua-chloro *cis*- and *trans*- $[\text{PtCl}(\text{NH}_3)_2(\text{H}_2\text{O})]^+$  complexes whose features have already been reported.<sup>9,18</sup>

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Gas-phase studies have provided valuable insight in the mechanistic features of ion-molecule reactions of platinum(I/II) ions, ranging from simple mono- or diatomic species to multiply ligated complexes.<sup>20–24</sup> In solution, considerable interest has been devoted to correlations of kinetic and thermodynamic parameters for the ligand substitution reaction of square planar platinum(II) complexes which led to define nucleophilic reactivity constants for various entering nucleophiles.<sup>25–27</sup> The best nucleophiles are soft and polarizable and the charge on the substrate is found to affect the order of nucleophilicity. However, it is often difficult to discriminate among the various factors that may affect the reactivity of a metal complex in solution. For example, in examining the reactivities of different Pt<sup>II</sup>–solvento complexes, the influences of the nature of the incoming and leaving ligands and of the *trans* and *cis* groups as well as the steric properties of both the substrate complex and the entering nucleophile are not separable.<sup>28</sup> In this regards, the gas-phase may represent the ultimate simplified environment.

## 2. Experimental

### 2.1 Sample preparation

Cisplatin, *cis*-PtCl<sub>2</sub>(NH<sub>3</sub>)<sub>2</sub>, and transplatin, *trans*-PtCl<sub>2</sub>(NH<sub>3</sub>)<sub>2</sub>, used in this work, were commercial products from Sigma-Aldrich and were used as received. Both compounds were dissolved in H<sub>2</sub>O LC-MS grade at a concentration of 10<sup>−3</sup> M. The aqueous solutions were incubated for one night before the analysis effected after dilution in H<sub>2</sub>O/MeOH 1 : 1 to obtain a final concentration of 10<sup>−5</sup> M. The end solution contained an appreciable fraction of [PtCl(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> ions. In order to favor the formation of hydrolyzed products, both cisplatin and transplatin dilute solutions were incubated two days longer leading to a conspicuous increase of the abundance of [Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> ions, at the expense of [PtCl(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup>.

### 2.2 IRMPD spectroscopy

IRMPD spectroscopy experiments were performed using an Optical Parametric Oscillator/Optical Parametric Amplifier (OPO/OPA) laser system (LaserVision) pumped by a Nd:YAG laser (Continuum Surelite II) operating at 10 Hz, coupled to a Paul ion trap mass spectrometer (Esquire 6000+, Bruker Daltonics) in an already described experimental setup.<sup>29</sup> The typical output energy from the OPO/OPA laser was *ca.* 27 mJ per pulse in the inspected spectral range with a deviation from the average of maximum 2 mJ per pulse. In the trap, ions were mass selected, stored for 30 ms and irradiated for 2 s. The H<sub>2</sub>O/MeOH solutions of cisplatin and transplatin were directly infused using a syringe pump at a flow rate of 180 μL h<sup>−1</sup> to the electrospray ionization (ESI) source. The laser induces the fragmentation of the ions stored in the trap when the IR frequency matches the energy of an IR active mode of the sampled species. The IRMPD spectra are obtained by plotting the photo-fragmentation yield  $R = -\log(I_p/(I_p + \Sigma I_f))$ , where  $I_p$  and  $\Sigma I_f$  are the parent and sum of the fragment ion intensity, respectively, as a function of the radiation wavenumber.

### 2.3 Ion-molecule reactions in FT-ICR mass spectrometry

Ion-molecule reactions have been examined in the cell of a Fourier transform-ion cyclotron resonance (FT-ICR) mass spectrometer (Bruker BioApex) equipped with an Apollo I ESI source and a 4.7 T magnet. Kinetic data were obtained at the nominal cell temperature of 300 K, as described in detail elsewhere.<sup>30</sup> The pseudo first order rate constants, obtained from the semilog plot of the parent ion decay with increasing reaction time, were divided by the neutral concentration to give the second order rate constant ( $k_{\text{exp}}$ ). The reaction efficiency ( $\text{Eff} = k_{\text{exp}}/k_{\text{coll}} \times 100$ , where  $k_{\text{coll}}$  is the collisional rate constant)<sup>31</sup> expresses the percentage of reactive collisions.

### 2.4 Density functional theory calculations

Quantum mechanics calculations have been employed to aid in interpreting the experimental data using density functional (DFT) methods as implemented in the Gaussian 09 Rev D.01 package.<sup>32</sup> The hybrid functional B3LYP has been used in combination with the aug-cc-pVTZ basis set for the light atoms. Relativistic effects affecting Pt were taken into account employing the pseudopotential aug-cc-pVTZ-PP.<sup>33,34</sup> The optimized structures were submitted to harmonic vibrational frequency analysis at the same level of theory in order to characterize the obtained structures as local minima or transition states and to obtain thermodynamic corrections to the electronic energies and IR spectra. The harmonic frequencies were scaled by a factor of 0.957, as used in previous studies.<sup>35,36</sup> Calculated linear IR spectra are convoluted with a Lorentzian line shape with a width (fwhm) of 5 cm<sup>−1</sup> for convenient comparison with the experimental spectra in the inspected IR region.

Structures and energies pertaining to the potential energy surface (PES) for the reaction of *cis*- and *trans*-[PtCl(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> with thioanisole were calculated at the ω-B97XD/6-311+G\*\* level with the LanL2TZ pseudopotential for the platinum atom, in order to be consistent with previous work.<sup>18</sup>

## 3. Results and discussion

### 3.1 Vibrational features and optimized structures of *cis*- and *trans*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> ions

*cis*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> and *trans*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> obtained by stepwise hydrolysis of cisplatin in aqueous solution have been assayed by IRMPD spectroscopy in order to unveil vibrational features of the bare mass selected ions. The mass spectrum of *trans*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> ions isolated in the ion trap mass spectrometer is shown in Fig. 1a. The isotopic pattern is distinctive of a species containing a platinum atom, excluding the contribution of formally isobaric peaks of [PtCl(NH<sub>3</sub>)<sub>2</sub>]<sup>+</sup> composition. The elemental composition of the ions is thus verified, agreeing with the calculated ion abundances of the isotopic cluster. Further confirmation of the ion composition is obtained by FT-ICR mass spectrometry operated in high resolution mode. As shown in Fig. S1 in the ESI,<sup>†</sup> the observed  $m/z$  values for the isotopic peaks are within ±2 ppm of the theoretical  $m/z$  value. The mass spectrum of *cis*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> is in



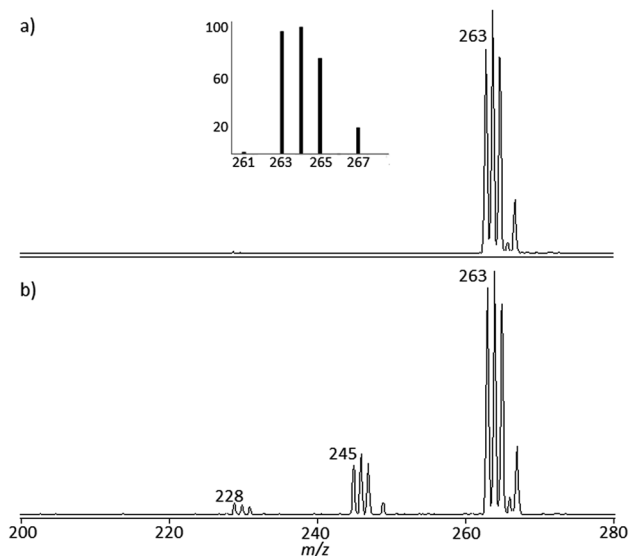


Fig. 1 Mass spectrum showing isolated *trans*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> ions at *m/z* 263 prior to (a) and after (b) irradiation by IR photons at 3600 cm<sup>-1</sup> (Esquire 6000). The inset shows the calculated isotope pattern for the given atom composition.

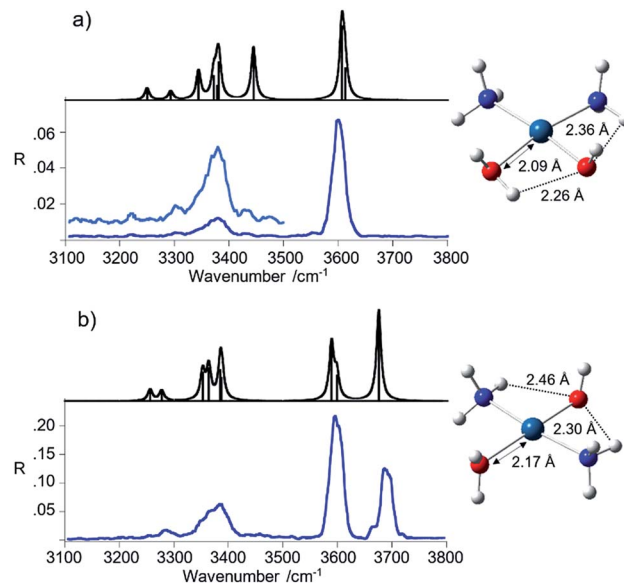


Fig. 2 IRMPD spectra of *cis*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> (a) and *trans*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> (b) (blue profiles) compared with the calculated linear IR spectra (black profiles). The optimized geometries are shown beside the respective spectra.

everything similar. For the sake of simplicity, henceforth isotopic cluster ions are named by the mass of the <sup>194</sup>Pt isotope containing species. The mass spectrometric behavior, namely the collision induced dissociation (CID) mass spectrum of the two species, is quite comparable, impeding any structural discrimination (see CID mass spectra reported in Fig. S2 in the ESI†). IR spectroscopy was therefore turned to as potential structural probe. Both *cis*- and *trans*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup>, when submitted to irradiation by IR photons in resonance with an active vibrational mode, undergo water loss yielding ions at *m/z* 245 (Fig. 1b). Additional NH<sub>3</sub> loss is also observed, albeit to a minor extent (*m/z* 228). The fragment ion profiles present similar features in both *m/z* 245 and *m/z* 228 channels and IRMPD spectra are obtained plotting the IRMPD yield as a function of the IR wavenumber.

The experimental spectra shown in blue in Fig. 2a and b are different for the two isomers. To begin with, the photofragmentation yield is different, the process being more efficient in the case of the *trans* isomer relative to the *cis*. This behavior is traced to the diverse energy threshold for the loss of water. DFT calculations providing optimized geometries and thermodynamic data (see Scheme S1 and Tables S1–S3 in the ESI†) show that the process is uphill by 149 and 119 kJ mol<sup>-1</sup> ( $\Delta H$  at 298 K) for the *cis* and *trans* isomer, respectively.

The spectrum of the *cis* isomer presents a strong signal at 3600 cm<sup>-1</sup> and less pronounced broad peaks from 3200 to 3450 cm<sup>-1</sup>, in the range expected for the asymmetric and symmetric N–H stretches of the ammonia ligands. A similar pattern appears in the spectrum of *trans*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup>, where the band at 3600 cm<sup>-1</sup> is however accompanied by another one at 3683 cm<sup>-1</sup> of comparable intensity. In order to gain insight into the observed spectral features, the geometries of *cis*- and *trans*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> have been optimized at B3LYP/aug-cc-pVTZ level using the aug-cc-pVTZ-PP pseudopotential for Pt.

The structures are also reported in Fig. 2 while thermodynamic data and Cartesian coordinates are given in Tables S1 and S3 in the ESI†, respectively. Harmonic frequency calculations were performed at the same level of theory and the computed spectra are compared to the experimental ones in Fig. 2, while vibrational frequencies and IR intensities are reported in Table 1.

Table 1 Vibrational frequencies for *cis*- and *trans*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> complexes

Vibrational mode <sup>a</sup>	Exp. <sup>b</sup>	Theor. <sup>b,c</sup>
<b><i>cis</i>-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup></b>		
$\nu$ OH	3600	3612 (61)
$\nu_{as}$ H <sub>2</sub> O		3606 (142)
$\nu_s$ H <sub>2</sub> O	3442	3444 (101)
$\nu_{as}$ NH <sub>3</sub>	3379	3381 (72)
$\nu_{as}$ NH <sub>3</sub>		3378 (27)
$\nu_{as}$ NH <sub>3</sub>		3372 (46)
$\nu_{as}$ NH <sub>3</sub>		3344 (55)
$\nu_s$ NH <sub>3</sub>	3300	3293 (17)
$\nu_s$ NH <sub>3</sub>	3222	3250 (22)
<b><i>trans</i>-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup></b>		
$\nu_{as}$ H <sub>2</sub> O	3683	3674 (166)
$\nu$ OH	3600	3598 (46)
$\nu_s$ H <sub>2</sub> O		3588 (103)
$\nu_{as}$ NH <sub>3</sub>	3384	3387 (42)
$\nu_{as}$ NH <sub>3</sub>		3385 (57)
$\nu_{as}$ NH <sub>3</sub>		3363 (60)
$\nu_{as}$ NH <sub>3</sub>		3352 (52)
$\nu_s$ NH <sub>3</sub>	3284	3277 (19)
$\nu_s$ NH <sub>3</sub>		3256 (21)

<sup>a</sup>  $\nu_s$  and  $\nu_{as}$  stand for symmetric and asymmetric stretching modes, respectively. <sup>b</sup> In cm<sup>-1</sup>. <sup>c</sup> Computed harmonic intensities (km mol<sup>-1</sup>) are reported in brackets.



The calculated linear IR spectra account well for the major signatures observed in the IRMPD spectra of both *cis*- and *trans*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> isomers. The matching is underlined in the data summarized in Table 1. Few structural and vibrational features may be highlighted. A notable difference between the two geometries is in the length of the Pt–OH<sub>2</sub> bond. The shorter distance in the *cis* isomer may be related to the presence of a H-bond interaction between the hydrogen of the water ligand lying on the molecular plane and the adjacent hydroxyl oxygen. This feature bears consequence in the vibrational spectrum.

The IRMPD spectrum of the *trans*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> isomer shows two bands in the OH stretching region at 3683 and 3600 cm<sup>−1</sup> corresponding to the H<sub>2</sub>O asymmetric stretching and a combination of the OH and H<sub>2</sub>O symmetric stretching modes, respectively. Interestingly, the O–H stretching modes in the spectrum of *trans*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> well reproduce the situation observed in the IRMPD spectrum of *trans*-[PtCl(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup>, where absorbances at 3683 and 3596 cm<sup>−1</sup> have been reported.<sup>9</sup> This finding highlights the comparable effect of either chloro or hydroxo substituents on the vibrational modes of the *trans* water ligand. A difference in relative intensity, enhancing the band at 3600 cm<sup>−1</sup> compared to the one at 3683 cm<sup>−1</sup> in *trans*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup>, is proportional to the additive contribution in the former by the OH stretching and H<sub>2</sub>O symmetric stretching oscillators.

In the *cis* isomer, one hydrogen atom of the water ligand on the molecular plane is oriented towards the adjacent hydroxyl oxygen, as already noticed. This interaction causes an important red shift of the water stretching modes. In particular, the asymmetric stretching shifts to 3606 cm<sup>−1</sup>, coalescing with the OH stretching calculated at 3612 cm<sup>−1</sup>, accounting for the band observed at 3600 cm<sup>−1</sup>. The symmetric H<sub>2</sub>O stretching is calculated at 3444 cm<sup>−1</sup> and barely emerges at 3442 cm<sup>−1</sup> in the IRMPD spectrum. However, it is not unusual to find an unexpectedly low intensity associated to stretching modes involved in hydrogen bonding. Possible reasons for this behavior include anharmonicity effects as well as effects related to the stepwise excitation in the IRMPD process, disrupting the hydrogen bond and removing the affected mode from resonance.<sup>36–39</sup>

Stretching modes of the ammonia ligands appear below 3400 cm<sup>−1</sup>. These modes are not particularly influenced by the relative position of the other ligands. Indeed, only few differences emerge from the analysis of the NH<sub>3</sub> stretching vibrations. The asymmetric stretching modes give rise to a broad feature with maximum at *ca.* 3380 cm<sup>−1</sup> in the spectra of both isomers. The presence of multiple absorptions close in frequency is compatible with the broad signal, well interpreted by the calculations. The symmetric stretching modes, instead, are poorly active and are observed at lower frequencies. It is interesting to note that in the IRMPD spectrum of *cis*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> the small band at 3300 cm<sup>−1</sup> matches the NH<sub>3</sub> symmetric stretching calculated at 3293 cm<sup>−1</sup>, while in the spectrum of the *trans* isomer the corresponding band is found at 3284 cm<sup>−1</sup> (calculated value 3277 cm<sup>−1</sup>). This slightly lower frequency is likely due to the interaction of both NH<sub>3</sub> ligands with the hydroxyl oxygen, a stronger binding motif than the one with water. In the *cis* isomer only one ammonia molecule is

allowed to interact with the OH group while the second one may interact only with H<sub>2</sub>O, yielding resonances calculated at 3250 cm<sup>−1</sup> and 3293 cm<sup>−1</sup>, respectively.

### 3.2 Reactivity of [PtX(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> (X = Cl, OH) complexes in ligand substitution reactions

*cis*- and *trans*-[PtX(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> (X = Cl, OH) complexes, formed in solution by stepwise hydrolysis of the respective precursors, cisplatin and transplatin, *cis*- and *trans*-PtCl<sub>2</sub>(NH<sub>3</sub>)<sub>2</sub>, have been allowed to react with selected ligands of appropriate volatility introduced at stationary concentration in the cell of an FT-ICR mass spectrometer. The ligands are endowed with functional groups that are well established biomolecular platinum sites, for example the thioether functional group of methionine, the aza group present in pyridine and in nucleobases, the oxygen atoms of phosphoric acid derivatives.<sup>4,29</sup>

The reaction common to all complexes [PtX(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> is ligand exchange whereby a water molecule is replaced by the incoming L, with L equal to trimethylphosphate (TMP), pyridine (Py), thioanisole (TA), and dimethylsulfide (DMS) (reaction (1)). Fig. 3 shows an example of kinetic plot for the reaction of *cis*-[PtCl(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> with DMS.

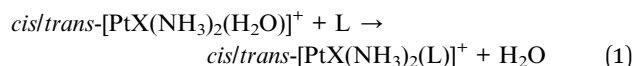


Table 2 summarizes the second order rate constants and corresponding efficiencies, representing the fraction of reactive collisions. Pyridine, the most basic among the selected ligands, displays also proton transfer reactivity and ligand addition to some extent. All reactions show rather low efficiencies. This finding is in line with the behavior of platinum(II) complexes, characterized by typically slow ligand exchange kinetics.<sup>25</sup>

The most notable feature emerging from the kinetic data in Table 2 is the consistently higher reactivity of [PtCl(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> complexes with respect to the hydroxo counterparts [Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup>. For example, reaction efficiency ratios for the reaction of TA with *cis*-[PtX(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> and

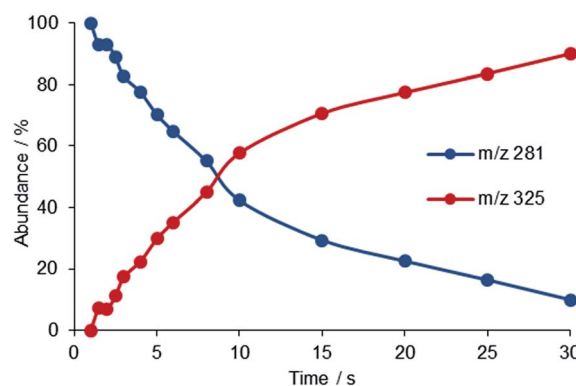


Fig. 3 Time dependence of ion abundances for the reaction of *cis*-[PtCl(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> (*m/z* 281) with DMS forming [PtCl(NH<sub>3</sub>)<sub>2</sub>(DMS)]<sup>+</sup> at *m/z* 325 in the FT-ICR cell at the DMS pressure of  $1.5 \times 10^{-7}$  mbar.





reach the TS for ligand exchange has a more pronounced variation. The two calculated TS structures are very similar, making the interpretation of the observed difference a difficult task. Two points should be highlighted: the presence of the chlorine atom in *trans* to the exchanging ligands leads to slightly longer bonds by *ca.* 0.07 Å suggesting the effect of the *trans* ligand not to be negligible. Both transition states present multiple interactions between the two ligands engaged in the substitution and the ligands in the *cis*-sites that can be implicated in the calculated difference. The listed elements probably contribute to give the final results that are anyway in fair agreement with the experiments. The reaction efficiency of the *trans*-isomer being 5.7 times higher with respect to the *cis*-one (6.3 *versus* 1.1, see Table 2) is, in fact, well interpreted by the higher energy difference between the separated reactants and the transition state for the *trans* with respect to the *cis* configuration (54 and 46 kJ mol<sup>-1</sup>, respectively). The *trans* isomer is thus more prone to undergo ligand substitution in view of the relatively more energy demanding back dissociation to reactants. The latter process remains, however, entropically favored which accounts for the low values of reaction efficiency, well below collision control.

Few other tested ligands proved to be unreactive, including H<sub>2</sub><sup>18</sup>O, which suggests a sizeable activation barrier for the water exchange process. Fig. 5 shows the symmetrical profile for reaction (2) with *cis*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> as reactant and product ion (thermodynamic data and coordinates of all species are given in Tables S1 and S3 in the ESI,<sup>†</sup> respectively). Here the collision complexes RC and PC are symmetrical, apart from the <sup>18</sup>O-isotope distribution. Their geometry is characterized by two hydrogen bonds linking a hydrogen atom of ligated water with an O atom of the external water and a H atom of the external H<sub>2</sub>O with the hydroxyl oxygen. In forming the RC complex, however, the hydrogen bond interaction between the in plane water hydrogen and the hydroxyl oxygen in *cis*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> is impaired, which may account for the relatively small binding energy of 72 kJ mol<sup>-1</sup>. The activation barrier for the

ligand exchange (RC → TS) is 91 kJ mol<sup>-1</sup>, protruding above the energy level of the isolated reactant pair. This feature implies that, in the single collision regime prevailing in the FT-ICR cell, the reaction will not occur, as experimentally observed. The RC complex would require an additional source of Δ*E* = 91 – 72 = 19 kJ mol<sup>-1</sup> for the reaction to proceed in the isolated, low pressure environment.

For comparison purposes, the transition state for the substitution of water from *cis*-[PtCl(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> by TMP, TA and Py has been found to lie 67, 46, and 43 kJ mol<sup>-1</sup> below the energy of the reactants, respectively, giving rise to productive collisions as shown in Table 2.<sup>18</sup> However, entropic factors account for reaction efficiencies (2.5, 1.1, and 0.41, for the tested series of ligands, TMP, TA, and Py, respectively) that are considerably below 100.

The whole set of calculations performed is thus consistent in interpreting the experimental results as a function of multiple factors. The reactivity of square planar complexes is often related to the presence of certain ligated atoms in *trans* position. The evidence presently obtained points in a different direction showing a remarkable importance of the *cis*-sites in entangling interactions with the reacting ligands all along the entire reaction. Significantly, groups possessing multiple binding sites, *e.g.* TMP, present the worst agreement with the qualitative indications suggested by the *trans* effect.

## 4. Conclusions

The early hydrolysis products from *cis* and *trans*platin, *cis*- and *trans*-[PtX(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> (X = Cl, OH), have been obtained by ESI and isolated in the gas-phase where their structure and reactivity properties have been comparatively examined. The gaseous environment permits to unambiguously characterize the selected ions. This task is hard to achieve in water solution where prototropic equilibria occur and the formation of hydroxo-bridged polynuclear complexes is an important competing process.<sup>48</sup> Even the use of buffered solutions presents the drawback of interferences due to reaction of hydrolyzed cisplatin with buffer components.<sup>7,44</sup>

IRMPD spectroscopy in the NH/OH stretching range has revealed the vibrational signatures of bare *cis*- and *trans*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> ions. While the IR spectrum of *trans*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> is very similar to the one of the chloro complex, the spectra of the *cis* complexes are rather different. The major variation in frequency affects the water OH stretching modes. Asymmetric and symmetric resonances are at 3624 and 3531 cm<sup>-1</sup> in the chloro complex and 3600 and 3442 cm<sup>-1</sup> in the hydroxo complex, respectively. Hydrogen bonding interactions are responsible for the different behavior that is manifested in the orientation of the water ligand, establishing a bisected hydrogen bonding interaction with the adjacent Cl in *cis*-[PtCl(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> while privileging a single H-bond between an in plane water hydrogen atom and the hydroxyl oxygen in *cis*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup>.

Both hydrolyzed complexes, [PtCl(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> and [Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup>, have been allowed to react with model compounds of biological ligands. All reactions show rather low

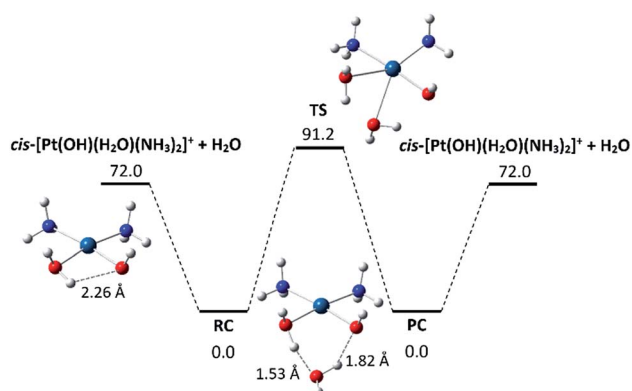


Fig. 5 Computed profile for the degenerate reaction of *cis*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> with H<sub>2</sub>O. Optimized geometries of *cis*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup>, of the collision complex RC/PC (*cis*-[Pt(OH)(NH<sub>3</sub>)<sub>2</sub>(H<sub>2</sub>O)]<sup>+</sup> · H<sub>2</sub>O) and of the transition state TS are shown. Relative enthalpies at 298 K (kJ mol<sup>-1</sup>) are reported.



efficiencies. This finding is in line with the behavior of platinum(II) complexes, characterized by relatively slow ligand exchange kinetics.<sup>25</sup> This feature is critically related to anti-tumor activity because it allows cisplatin and related drugs to reach the cell intact and effectively bind to cellular DNA rather than to extracellular binding sites.<sup>25,49</sup>

The reactivity of the chloro complexes,  $[\text{PtCl}(\text{NH}_3)_2(\text{H}_2\text{O})]^+$ , is consistently higher than for the hydroxo-ligated counterparts,  $[\text{Pt}(\text{OH})(\text{NH}_3)_2(\text{H}_2\text{O})]^+$ , the chloro substituent likely enhancing the electrophilicity of the metal. This finding may be related to the observation that the majority of the binding of cisplatin to DNA occurs *via* the mono-aquated species  $\text{cis-}[\text{PtCl}(\text{NH}_3)_2(\text{OH}_2)]^{+1-7}$ . In comparing the reactivity of different ligands, caution is due to the contribution of the reactant pair binding energy that affects the reaction efficiency together with the ligand exchange activation barrier. As expected from the soft character of platinum(II) an S-nucleophile like TA is fairly reactive, in accord with the function of S-donor compounds both as Pt-reservoirs in the body and as “rescue agents”.<sup>40–45</sup> Notably, also TMP, an O-donor ligand, shows relatively high efficiency in ligand exchange, thus supporting a role for phosphate groups in cisplatin binding and delivery.

It may be underlined here that the present report on the gas phase reactivity of *cis*- and *trans*- $[\text{PtX}(\text{NH}_3)_2(\text{H}_2\text{O})]^+$  complexes does not take into account the effect of the solvent that has to be included, for example, also when theoretical work is applied to interpret reactions occurring in solution, as in the case of cisplatin hydrolysis itself.<sup>50</sup> Still, it is interesting to observe here a manifestation of the higher intrinsic reactivity of transplatin towards sulfur nucleophiles at the level of the naked species. This result is reminiscent of the more extensive reaction of transplatin with glutathione in red blood cells.<sup>51</sup> In this regard, further efforts will be devoted to gain insight into the structural and reactivity features that affect the distinct behavior of *cis* and *trans* platinum(II) complexes in a variety of contexts,<sup>52,53</sup> most importantly in binding to DNA within the cell nucleus, using the tool of isolating from solution the key intermediates to be examined as long lived species in the gas phase.

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## References

- 1 T. C. Johnstone, K. Suntharalingam and S. J. Lippard, *Chem. Rev.*, 2016, **116**, 3436–3486.
- 2 B. W. Harper, A. M. Krause-Heuer, M. P. Grant, M. Manohar, K. B. Garbutcheon-Singh and J. R. Aldrich-Wright, *Chem.–Eur. J.*, 2010, **16**, 7064–7077.
- 3 D. Wang and S. J. Lippard, *Nat. Rev. Drug Discovery*, 2005, **4**, 307–320.
- 4 J. Reedijk, *Proc. Natl. Acad. Sci. U. S. A.*, 2003, **100**, 3611–3616.
- 5 J. D. White, M. M. Haley and V. J. DeRose, *Acc. Chem. Res.*, 2016, **49**, 56–66.
- 6 J. Kozelka, *Inorg. Chim. Acta*, 2009, **362**, 651–668.
- 7 M. S. Davies, S. J. Berners-Price and T. W. Hambley, *Inorg. Chem.*, 2000, **39**, 5603–5613.
- 8 A. Springer, C. Buergel, V. Boehrsch, R. Mitric, V. Bonacic-Koutecky and M. W. Linscheid, *ChemPhysChem*, 2006, **7**, 1779–1785.
- 9 A. De Petris, A. Ciavardini, C. Coletti, N. Re, B. Chiavarino, M. E. Crestoni and S. Fornarini, *J. Phys. Chem. Lett.*, 2013, **4**, 3631–3635.
- 10 J. Oomens, B. G. Sartakov, G. Meijer and G. Von Helden, *Int. J. Mass Spectrom.*, 2006, **254**, 1–19.
- 11 N. C. Polfer, *Chem. Soc. Rev.*, 2011, **40**, 2211–2221.
- 12 J. Roithova, *Chem. Soc. Rev.*, 2012, **41**, 547–559.
- 13 T. D. Fridgen, *Mass Spectrom. Rev.*, 2009, **28**, 586–607.
- 14 J. R. Eyler, *Mass Spectrom. Rev.*, 2009, **28**, 448–467.
- 15 L. MacAleese and P. Maitre, *Mass Spectrom. Rev.*, 2007, **26**, 583–605.
- 16 B. Chiavarino, M. E. Crestoni, S. Fornarini, D. Scuderi and J.-Y. Salpin, *J. Am. Chem. Soc.*, 2013, **135**, 1445–1455.
- 17 B. Chiavarino, M. E. Crestoni, S. Fornarini, D. Scuderi and J.-Y. Salpin, *Inorg. Chem.*, 2015, **54**, 3513–3522.
- 18 D. Corinti, C. Coletti, N. Re, B. Chiavarino, M. E. Crestoni and S. Fornarini, *Chem.–Eur. J.*, 2016, **22**, 3794–3803.
- 19 C. C. He, B. Kimutai, X. Bao, L. Hamlow, Y. Zhu, S. F. Stroehn, J. Gao, G. Berden, J. Oomens, C. S. Chow and M. T. Rodgers, *J. Phys. Chem. A*, 2015, **119**, 10980–10987.
- 20 D. Schröder and H. Schwarz, *Can. J. Chem.*, 2005, **83**, 1936–1940.
- 21 B. Butschke and H. Schwarz, *Chem.–Eur. J.*, 2012, **18**, 14055–14062.
- 22 M.-E. Moret and P. Chen, *Organometallics*, 2007, **26**, 1523–1530.
- 23 R. Liyanage, M. L. Styles, R. A. J. O’Hair and P. B. Armentrout, *J. Phys. Chem. A*, 2003, **107**, 10303–10310.
- 24 S. Wee, R. A. J. O’Hair, A. J. Richard and W. D. McFadyen, *Rapid Commun. Mass Spectrom.*, 2004, **18**, 1221–1226.
- 25 D. T. Richens, *Chem. Rev.*, 2005, **105**, 1961–2002.
- 26 U. Belluco, L. Cattalini, F. Basolo, R. G. Pearson and A. Turco, *J. Am. Chem. Soc.*, 1965, **87**, 241–246.
- 27 B. Pitteri, L. Canovese, G. Chessa, G. Marangoni and P. Uguagliati, *Polyhedron*, 1992, **11**, 2363–2373.
- 28 S. Elmroth, Z. Bugarcic and L. I. Elding, *Inorg. Chem.*, 1992, **31**, 3551–3554.
- 29 R. K. Sinha, P. Maitre, S. Piccirillo, B. Chiavarino, M. E. Crestoni and S. Fornarini, *Phys. Chem. Chem. Phys.*, 2010, **12**, 9794–9800.
- 30 F. Angelelli, B. Chiavarino, M. E. Crestoni and S. Fornarini, *J. Am. Soc. Mass Spectrom.*, 2005, **16**, 589–598.
- 31 T. Su and W. J. Chesnavich, *J. Chem. Phys.*, 1982, **76**, 5183–5185.
- 32 M. J. Frisch, *et al.*, Complete reference is given in the ESI data.†
- 33 D. Figgen, K. A. Peterson, M. Dolg and H. Stoll, *J. Chem. Phys.*, 2009, **130**, 164108–164119.
- 34 B. Pinter, V. Van-Speybroeck, M. Waroquier, P. Geerlings and F. De-Proft, *Phys. Chem. Chem. Phys.*, 2013, **15**, 17354–17365.
- 35 P. Sinha, S. E. Boesch, C. Gu, R. A. Wheeler and A. K. Wilson, *J. Phys. Chem. A*, 2004, **108**, 9213–9217.



- 36 A. Bouchet, M. Schütz, B. Chiavarino, M. E. Crestoni, S. Fornarini and O. Dopfer, *Phys. Chem. Chem. Phys.*, 2015, **17**, 25742–25754.
- 37 F. Turecek, C. L. Moss, I. Pikalov, R. Pepin, K. Gulyuz, N. C. Polfer, M. F. Bush, J. Brown, J. Williams and K. Richardson, *Int. J. Mass Spectrom.*, 2013, **354**, 249–256.
- 38 N. Heine, T. I. Yacovitch, F. Schubert, C. Brieger, D. M. Neumark and K. R. Asmis, *J. Phys. Chem. A*, 2014, **118**, 7613–7622.
- 39 B. Chiavarino, M. E. Crestoni, M. Schütz, A. Bouchet, S. Piccirillo, V. Steinmetz, O. Dopfer and S. Fornarini, *J. Phys. Chem. A*, 2014, **118**, 7130–7138.
- 40 Z. D. Bugarcic, J. Bogojewski, B. Petrovic, S. Hochreuther and R. van Eldik, *Dalton Trans.*, 2012, **41**, 12329–12345.
- 41 A. Casini and J. Reedijk, *Chem. Sci.*, 2012, **3**, 3135–3144.
- 42 G. Ferraro, L. Messori and A. Merlino, *Chem. Commun.*, 2015, **51**, 2559–2561.
- 43 H. Li, Y. Zhao, H. I. A. Phillips, Y. Qi, T. Y. Lin, P. J. Sadler and P. B. O'Connor, *Anal. Chem.*, 2011, **83**, 5369–5376.
- 44 J. Vinje, E. Sletten and J. Kozelka, *Chem.–Eur. J.*, 2005, **11**, 3863–3871.
- 45 O. Pinato, C. Musetti, N. P. Farrell and C. Sissi, *J. Inorg. Biochem.*, 2013, **122**, 27–37.
- 46 M. L. Chabiny, S. L. Craig, C. K. Regan and J. I. Brauman, *Science*, 1998, **279**, 1882–1886.
- 47 S. Gronert, *Chem. Rev.*, 2001, **101**, 329–360.
- 48 S. J. Berners-Price and T. G. Appleton, The Chemistry of Cisplatin in Aqueous Solution, in *Platinum Based Drugs in Cancer Therapy*, ed. L. R. Kelland and N. Farrell, Humana Press Inc., Totowa, N. J., U. S. A., 2000, ch. 1.
- 49 J. Reedijk, *Eur. J. Inorg. Chem.*, 2009, **10**, 1303–1312.
- 50 Y. Zhang, Z. Guo and X.-Z. You, *J. Am. Chem. Soc.*, 2001, **123**, 9378–9387.
- 51 S. J. Berners-Price and P. W. Kuchel, *J. Inorg. Biochem.*, 1990, **38**, 327–345.
- 52 Z. Tao, Y. Xie, J. Goodisman and T. Asefa, *Langmuir*, 2010, **26**, 8914–8924.
- 53 G. McGowan, S. Parsons and P. J. Sadler, *Inorg. Chem.*, 2005, **44**, 7459–7467.

