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Construction and functional assessment of zein thin film incorporating spindle-like ZnO crystals

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The use of plastics has rapidly progressed during recent years in food packaging, which can be attributed to technological innovations in the synthesis of plastics and lower costs. However, the massive accumulation of plastic waste is often accompanied with their excessive consumption because of their poor degradability. Consequently, interest in the native materials is growing. It is noted that zein is a corn protein, has good compatibility and has been widely applied in the food industry. However, microbial contamination is a huge detriment to its long-term applications. This study aims to identify a method, which can provide antibacterial activity to a zein thin film. Monodispersed and spindle-like ZnO crystals made of nanocrystals were synthesized and added into zein thin films. EDX mapping confirmed the homogeneous distribution of ZnO in films. The antibacterial activities have been approved and showed great stability. In summary, the potential of zein thin films incorporating ZnO crystals as a functional packaging film has been demonstrated.

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1. Introduction

Food-borne diseases are becoming one of the serious problems faced by humans along with environmental contamination.¹ Technological innovations in food safety related to consumer confidence and human health are becoming extremely urgent. Traditional synthetic plastics have been widely applied in food packaging in an accelerated mode. Although they are from low-cost raw materials and have good mechanical properties, poor biodegradability is one of their fatal flaws and it is resulting in a massive accumulation of plastic waste and subsequent serious environmental problems.² Therefore, scientists are concentrating their efforts on finding new techniques and packaging materials.³

Biodegradable and natural polymers have recently aroused great attention for their relative abundance, film-forming ability or nutritional qualities.^{4,5} Especially, the appearance of slow-release technique drives the development of the native polymers-based active packaging, which is becoming a highlight in food industry.^{6–8} For example, Dalarbashi *et al.* fabricated a new active packaging film made from a soluble soybean polysaccharide incorporating ZnO nanoparticles,

which showed good antibacterial, antifungal and yeasticidal activities.⁹

This study aimed to prepare an active packaging made of zein and ZnO crystals. Herein, biomimetic synthesis, characterizing with environmental-friendly and facile conditions, has been used to fabricate spindle-like ZnO crystals. Zein, a predominant storage protein of corn, has exhibited great biodegradability and excellent biocompatibility, and can act as the structure-directing agent for nanocrystal synthesis and the formation of films.^{10–17} EDX mapping confirmed that ZnO crystals could homogeneously distribute in zein thin films. The zein thin film incorporating ZnO crystals could effectively inhibit the growth of *Escherichia coli* (*E. coli*) and *Staphylococcus aureus* (*S. aureus*), and these antibacterial activities could maintain a long-term effect. All of these results support the conclusion that zein thin films incorporating ZnO crystals are suitable to act as a functional packaging materials.

2. Material and methods

2.1 Materials

Zein was purchased from Sigma (St. Louis, MO, USA). Zinc chloride (>99.999%, MW = 136.3 g mol⁻¹, AR) was purchased from Tianjin Chemical Reagent Factory (Tianjin, China). All other solvents and chemicals were analytical grade.

2.2 Biomimetic synthesis of spindle-like zein-conjugated ZnO crystals with hierarchical structure

Spindle-like zein-conjugated ZnO crystals with a hierarchical structure were prepared by a modified biomimetic method using zein as the structure-directing agent. Briefly, 20 mL of

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a 6 mg mL⁻¹ zinc chloride aqueous solution was quickly added dropwise into 40 mL of a 2 mg mL⁻¹ zein alcoholic aqueous solution (60%) under stirring. The pH value of the mixture was adjusted to 8.0–8.9 using a 1 M NaOH solution and the mixture was kept static for 30 min at 37 °C. Finally, the as-prepared sample was separated by centrifugation for 10 min at 10 000 rpm. The collected product was washed with distilled water and lyophilized under vacuum for 48 h.

2.3 Fabrication of zein thin films incorporating ZnO crystals

Zein (600 mg) and zein-conjugated ZnO crystals (100 mg) were completely dispersed into 75% alcoholic aqueous solution. Then, 3 mg of cysteine, 100 μL 1-glycerol and 100 μL of polyethylene glycol were added in sequence and stirred for 35 min at 80 °C. Following this, the films were cast by pouring the mixture onto polystyrene Petri dishes and fumigated for 2 h at 70 °C in a water bath. The thin films were peeled from the casting surface and stored in a humidified incubator.

2.4 Characterization

The surface structure and morphology of the zein-conjugated ZnO crystals were observed using a field emission scanning electron microscope (Merin Compact, Zeiss, German) and the distribution of ZnO in the zein thin films was detected by EDX mapping. Furthermore, the inner structure of the ZnO crystals was observed using a JEM-2010 electron microscope working at 200 kV.

Fourier transform infrared spectroscopy (FTIR) was applied to confirm the organic composition of the zein-conjugated ZnO crystals. The analysis was carried out on a Bio-Rad FTS-40 Fourier transform infrared spectrophotometer in the wave-number range of 4000–400 cm⁻¹. The spectra were collected at a 2 cm⁻¹ resolution with 128 scans by preparing KBr pellets with a 3 : 100 “sample-to-KBr” ratio. Moreover, the organic content in the ZnO crystals was quantified by thermogravimetric-differential scanning calorimetry (TG/DTA) on an EXSTAR TG/DTA 6300 instrument (Seiko, Japan).

To confirm the composition of the inorganic component, XRD measurements were performed on a Bruker D8 & Advance X-ray powder diffractometer with graphite monochromatized Cu/Kα (γ = 0.15406 nm). A scan rate of 0.05 deg s⁻¹ was applied to record the pattern in the 2θ range of 10–80°.

The stability of the thin films was determined in an aqueous solution with different pH values. Briefly, the thin films were immersed into an aqueous solution at different time points and the Zn ion content in solution was quantified by atomic absorption spectrometry (AAS).

2.5 Enzymolysis

The zein thin film (diameter = 15 mm, n = 3) was immersed into 4 mL of an enzyme solution containing 12.5 U mL⁻¹ collagenase and incubated at 37 °C under static conditions. Each time the enzyme solution with the same volume as that taken out was added to the solution. The degradation rate was analysed spectrophotometrically according to our previous study.¹⁴

2.6 Water vapour permeability (WVP)

The WVP of the zein thin film was determined according to the report of Abdollahi *et al.*¹⁸ Briefly, glass permeation cups (diameter = 30 mm, n = 3) containing distilled water were covered with the zein thin film and kept static at 20 °C and 1.5% relative humidity. The weight of the cup was recorded at 1 h intervals for 8 h. The WVP was calculated as follows:

$$WVP = WVTR \times L/\Delta P$$

where WVTR is the measured water vapour transmission rate (g m⁻² s⁻¹) through the film, *L* is the mean film thickness (m), and Δ*P* is the partial water vapour pressure difference (Pa) across the two sides of the film.

2.7 Antibacterial activity

The antibacterial activities of the thin films were evaluated using the disc diffusion method and two microorganism strains of *E. coli* (Gram-negative bacteria) and *S. aureus* (Gram-positive bacteria) were chosen. Briefly, all microorganism pre-inoculum cultures were grown overnight at 37 °C in 20 mL of nutrient broth (made of 1 g L⁻¹ beef extract; 5 g L⁻¹ neutralized peptone; 2 g L⁻¹ yeast extract; 5 g L⁻¹ NaCl) and subjected to horizontal shaking at 100 rpm. The samples (diameter = 6 mm, n = 6) were placed onto the nutrient agar that was previously inoculated with different microorganisms. After incubation for 24 h, the presence of a zone of growth inhibition below and around the specimen was checked by visual inspection.

2.8 Statistics

All data represented as the mean ± SD and the number of the independent replicates is individually shown for each experiment. The analysis of data was carried out by one-way factorial analysis of variances (ANOVA) and multiple comparisons (Fisher's method as *post hoc* test, *p* < 0.05).

3. Results and discussion

The specific requirements for environment protection and food safety drive the innovation of food packaging with an emphasis on the roles of biodegradability and antibacterial properties.¹⁹ This study focused on understanding the feasibility to construct a zein thin film incorporating ZnO crystals. We firstly developed a modified biomimetic synthesis route to fabricate zein conjugated ZnO crystals. This route had two advantages. First, the entire synthesis route could be accomplished under eco-friendly and gentle conditions, which will reduce the dubious factors from an organic system and investment cost from special equipment. Second, the participation of zein in spindle-like ZnO crystals will be helpful for their better distribution in a zein thin film. With our interest in active food packaging, we envisaged that a zein thin film incorporating ZnO crystals will generate a new class of active food packaging that could be biodegradable and antibacterial.

The XRD analysis clearly demonstrates the formation of ZnO crystals as shown in Fig. 1A. It can be seen that eight



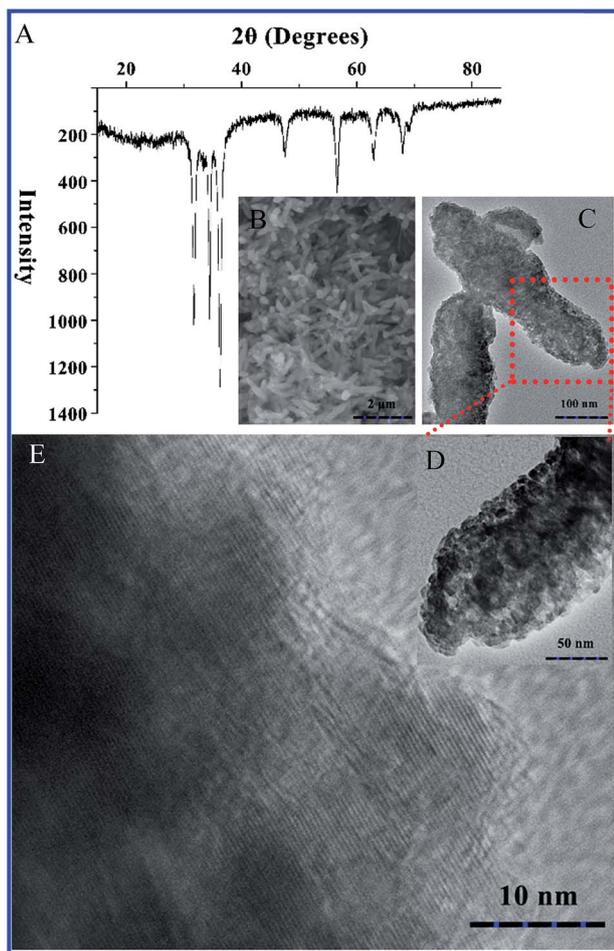


Fig. 1 Analysis on the inorganic component and morphologies of zein conjugated ZnO crystals. (A) XRD spectrum; (B) SEM observation; (C–E) TEM observation.

characteristic peaks for ZnO appear at 2θ at 31.769° , 34.421° , 36.252° , 47.538° , 56.602° , 62.862° , 66.378° , 67.961° and 69.098° , corresponding to ZnO (100), (002), (101), (102), (110), (103), (200), (110) and (201) diffractions. These peaks are in good agreement with the reported data of the JCPDS card 36-1451. In this study, we supplemented 60% ethanol into the reactive system instead of high temperature to promote the transformation from $\text{Zn}(\text{OH})_2$ to ZnO.²⁰ Moreover, biomimetic synthesis allows us to control the morphologies and size of nanocomposites using native molecules as templates.^{21,22} Both SEM and TEM clearly confirm that zein acts as a good transport carrier in Ostwald ripening and induces the formation of a spindle-like structure that is comprised of nanoparticles (Fig. 1B–E).²³

Fig. 2 shows the FTIR spectrum of pure ZnO crystals, pure zein and zein conjugated ZnO crystals. It can be seen that the typical Zn–O stretching band in pure ZnO crystals appears between 430 cm^{-1} and 520 cm^{-1} (Fig. 2A).^{24–26} As for pure zein, the peaks at 3396 cm^{-1} , 2958 cm^{-1} , 1651 cm^{-1} and 1539 cm^{-1} are clearly separated and can be assigned to the stretching vibration of a hydroxyl group, amide A', and amide I and amide

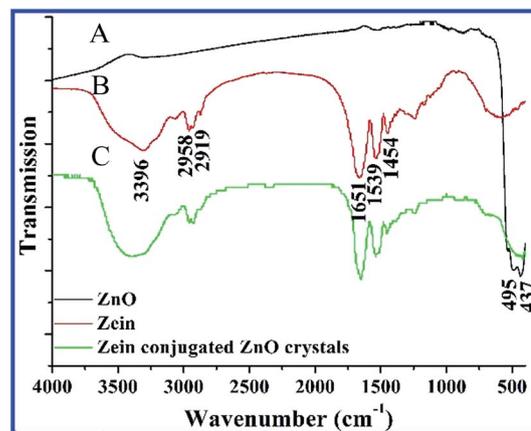


Fig. 2 Analysis on the organic component of zein conjugated ZnO crystals by FTIR spectra. (A) Pure ZnO, (B) pure zein, and (C) zein conjugated ZnO crystals.

II, which are the typical protein absorption peaks (Fig. 2B). By comparing Fig. 2A and B, the FTIR spectrum of zein conjugated ZnO crystals supplies direct evidence of zein participating in the formation of spindle-like ZnO crystals (Fig. 2C).

According to TG/DTA analysis, we could easily quantify the zein content in ZnO crystals (Fig. 3A). The total weight loss of the ZnO crystals decreases to 44.42% from $250\text{ }^\circ\text{C}$ to $650\text{ }^\circ\text{C}$ according to the TG curve. In the DTA curve, there are three exothermic peaks, appearing at $365\text{ }^\circ\text{C}$, $485\text{ }^\circ\text{C}$ and $531\text{ }^\circ\text{C}$,

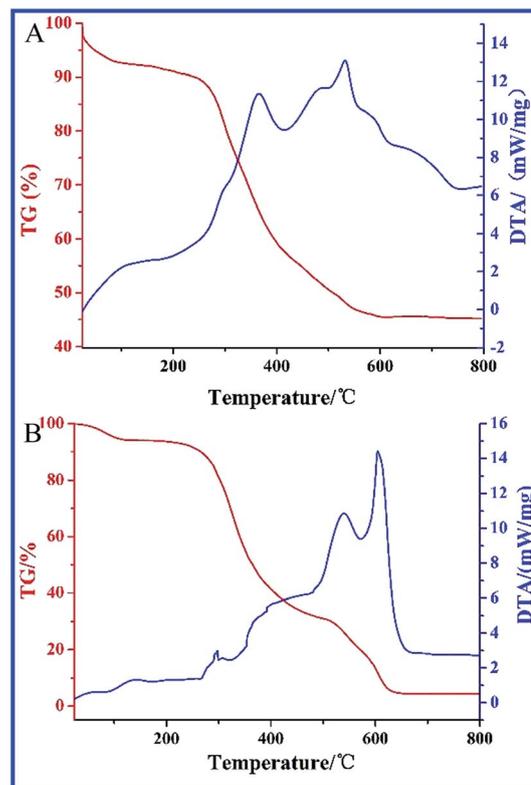


Fig. 3 Analysis on the content of zein in zein conjugated ZnO crystals by TG/DTA. (A) Zein conjugated ZnO crystals; (B) pure zein.



respectively, which correspond to the carbonization, side chain-burning and main carbon-burning of zein, respectively.²⁷ However, by comparison with that of pure zein (Fig. 3B), the peaks of the abovementioned exothermic process appear at 404 °C, 538 °C and 603 °C, respectively, suggesting the stimulative effect of ZnO on zein burning.

Currently, inorganic nanomaterials in the prevention of bacterial infections are gaining an importance due to low drug resistance and stable physical and chemical properties compared to traditional antimicrobials. Literature studies have demonstrated that ZnO could effectively inhibit the growth of microorganisms by oxidative stress and the interaction of zinc with the bacteria membrane.^{28,29} Moreover, these antibacterial activities are species-dependent.^{30,31} For example, Baek and An demonstrated that *S. aureus* was more sensitive than *E. coli* to ZnO nanoparticles.³⁰ Sultana *et al.* reported that ZnO nanoparticles exhibited stronger antibacterial activity against *S. aureus* and *Bacillus subtilis* compared to *E. coli* and *Pseudomonas aeruginosa*.³¹ In this study, we also tested the antibacterial activities of zein conjugated ZnO crystals against *S. aureus* and *E. coli* (Fig. 4). The results indicate that the ZnO crystals keep a positive correlation with ZnO concentrations. At the same time, *S. aureus* shows higher sensitivity than *E. coli* to ZnO crystals, with 1.14 $\mu\text{g mm}^{-2}$ and 1.89 $\mu\text{g mm}^{-2}$ of minimum inhibitory concentration, respectively.

Observations of the monodispersed and hierarchical structure and excellent antibacterial activity of zein conjugated ZnO crystals prompted us to focus on the fabrication of zein thin films incorporating ZnO crystals. The obtained thin films are yellow and transparent as shown in Fig. 5A. Moreover, we could easily control the thickness by adjusting the content of raw materials. Fig. 5B and C show the microstructure of the thin film observed by SEM and there are a few remaining air bubbles in the film. As one of the typical elements of a protein, N could be detected by EDX analysis (Fig. 5D). In addition, we also can detect Zn, which is from ZnO crystals. By the AAS test, the loading of ZnO in a thin film can get to 0.66 mg cm^{-2} . The homogeneity of the ZnO crystals in a thin film was directly observed by EDX mapping based on the distribution of Zn and N. As shown in Fig. 5E–G, zein has a homogeneous distribution according to N mapping,

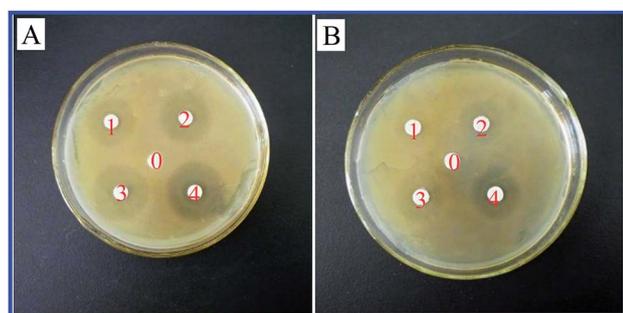


Fig. 4 The antibacterial activity of zein conjugated ZnO crystals with different concentrations against *S. aureus* (A) and *E. coli* (B) using the disc diffusion method. "0": control; "1": 2.83 $\mu\text{g mm}^{-2}$; "2": 8.49 $\mu\text{g mm}^{-2}$; "3": 14.15 $\mu\text{g mm}^{-2}$; "4": 28.31 $\mu\text{g mm}^{-2}$.

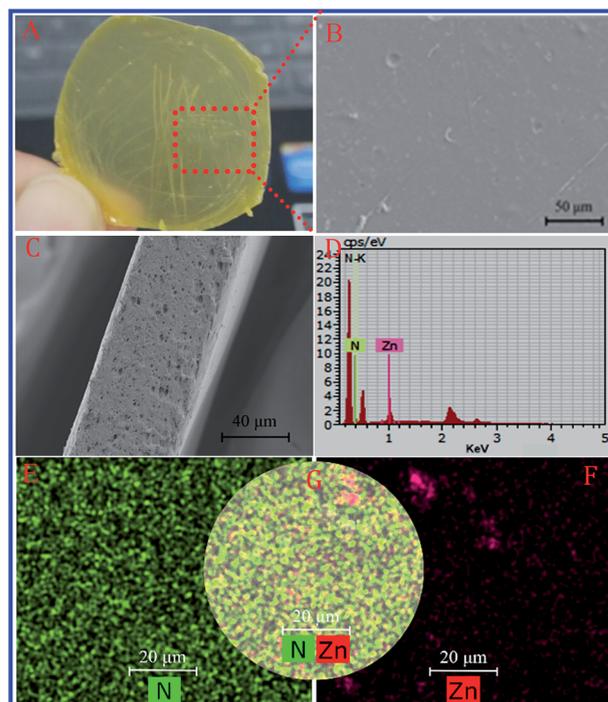


Fig. 5 Morphological observations, elemental composition and phase distribution detection of zein thin films incorporating these crystals. (A) Visual observation; (B) SEM image (facade); (C) SEM image (profile); (D) EDX analysis; (E) EDX mapping on N; (F) EDX mapping on Zn; (G) EDX mapping on N and Zn.

while the presence of only a slight aggregate of Zn signals suggests a homogeneous distribution of ZnO in a thin film.

Protein-based materials are known to undergo naturally-controlled degradation processes. Zein, the predominant storage protein of corn, has exhibited great biodegradability. In previous studies several groups including our group, have demonstrated that zein could be degraded *via* enzymolysis.^{14,15,32} In order to understand the biodegradability of a zein thin film, we further studied its degradation process *in vitro*. A zein thin film ($d = 1.5$ cm) was immersed into 4 mL of an enzyme solution containing 12.5 U mL^{-1} collagenase and incubated at 37 °C in an incubator under static conditions. As shown in Fig. 6, the degradation rate of the zein thin film increases gradually in 5 days and reaches 66.8%, suggesting its good biodegradability. In addition, we also tested the water vapour permeability of the zein thin film. The result shows that the WVP of the zein thin film incorporating ZnO crystals reaches 4.15 ± 0.12 ($\text{g ms}^{-1} \text{ Pa}^{-1}$) 10^{-11} . The relatively lower permeability might result from the water-insoluble property of zein.²

Active food packaging is currently a focus of attention that is linked to the development of the shelf-life and safety of packaged food.^{33,34} Herein, we investigated the antibacterial activities of a zein thin film incorporating ZnO crystals against various bacterial pathogens. As shown in Fig. 7, thin films can effectively inhibit the growth of *S. aureus* and *E. coli*. Furthermore, the inhibition zone size reached 25.39 ± 0.90 mm against



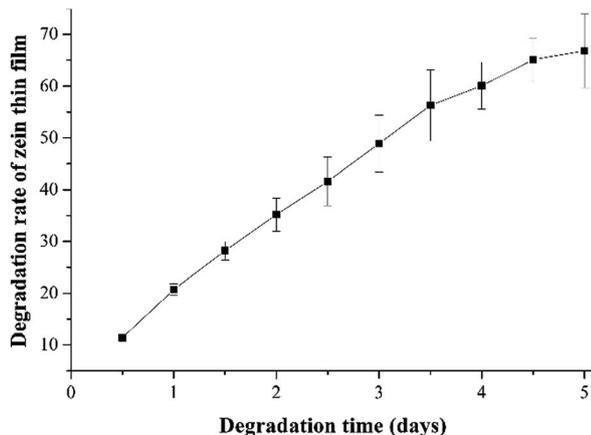


Fig. 6 Degradation of zein thin film by 12.5 U mL⁻¹ collagenase.

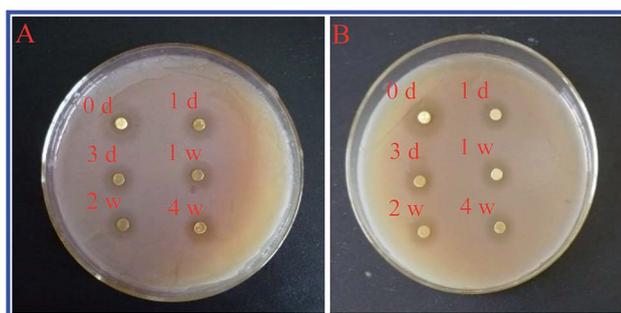


Fig. 7 The antibacterial activity of a zein thin film incorporating ZnO crystals after immersion in an aqueous solution at different times. (A) *S. aureus*, (B) *E. coli*.

S. aureus and 21.09 ± 1.89 mm against *E. coli*. Interestingly, the zein thin films incorporating ZnO crystals exhibited continuous antibacterial activities. After 4 weeks of immersion in the aqueous solution, the thin film still maintained a high inhibitory effect on both bacterial pathogens.

In order to understand the antibacterial behaviours, we analysed the stability of zein thin films incorporating ZnO crystals in an aqueous solution with different pH values (Fig. 8).

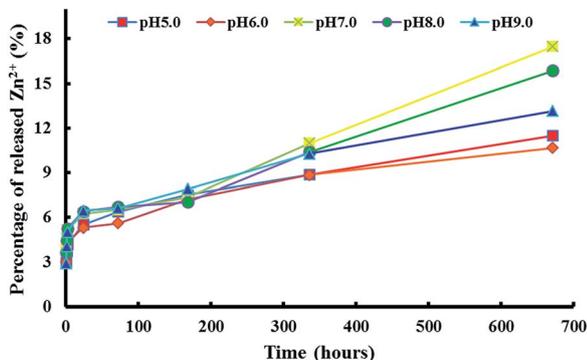


Fig. 8 Zinc ion released from a zein thin film incorporating ZnO crystals in an aqueous solution with different pH values.

It can be seen that the thin films kept a high stability under different pH conditions ranging from 5.0 to 9.0 and the highest percentage of released Zn²⁺ from the thin films was only 17.4% in 4 weeks. It has been well documented that ZnO can release Zn²⁺ and the excess zinc can penetrate through the bio-membrane, affect the antioxidant defence system and further induce toxicity.^{9,35,36} Therefore, this result can explain why the thin film still has high antibacterial activities after 4 weeks of immersion. In summary, it will be very interesting to further study the application of a zein thin film incorporating ZnO crystals in food packaging due to its better film-forming effect and excellent antibacterial activities.

4. Conclusion

In summary, this study prepared a zein thin film incorporating ZnO crystals by the solution-casting method. Spindle-like zein conjugated ZnO crystals were synthesized *via* a green and gentle route. The participation of zein in zein conjugated ZnO crystals was key for the homogeneous distribution of ZnO crystals in the final zein thin film. At the same time, the synthesized zein thin film incorporating ZnO crystals showed high and long-term antibacterial activities against both Gram-negative and Gram-positive pathogens. Moreover, the bacteriostatic effect is due to the released zinc ion from the thin film. These data can serve as a guide for the application of a zein thin film incorporating ZnO crystals to food packaging material.

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Notes and references

- 1 N. B. Johnson, L. D. Hayes, K. Brown, E. C. Hoo and K. A. Ethier, *MMWR Surveillance Summaries: Morbidity and Mortality Weekly Report*, 2014, vol. 63, suppl. 4, p. 3.
- 2 H. Rostamzad, S. Y. Paighambari, B. Shabanpour, S. M. Ojagh and S. M. Mousavi, *Food Packaging and Shelf Life*, 2016, vol. 7, p. 1.
- 3 N. N. V. Long, C. Joly and P. Dantigny, *Int. J. Food Microbiol.*, 2016, **220**, 73.
- 4 C. Pires, C. Ramos, G. Teixeira, I. Batista, R. Mendes, L. Nunes and A. Marques, *J. Food Eng.*, 2011, **105**, 422.
- 5 P. Tongnuanchan, S. Benjakul, T. Prodpran and P. Songtipy, *Int. J. Biol. Macromol.*, 2011, **48**, 758.
- 6 P. Appendini and J. H. Hotchkiss, *Innovative Food Sci. Emerging Technol.*, 2002, **3**, 113.
- 7 L. Wang, F. Liu, Y. Jiang, Z. Chai, P. Li, Y. Cheng, H. Jing and X. Leng, *J. Agric. Food Chem.*, 2011, **59**, 12411.



- 8 A. Llorens, E. Lloret, P. Picouet and A. Fernandez, *Int. J. Food Microbiol.*, 2012, **158**, 113.
- 9 D. Salarbashi, S. A. Mortazavi, M. S. Noghabi, B. S. F. Bazzaz, N. Sedaghat, M. Ramezani and I. Shahabi-Ghahfarrokhi, *Carbohydr. Polym.*, 2016, **140**, 220.
- 10 J. Dong, Q. S. Sun and J. Y. Wang, *Biomaterials*, 2004, **25**, 4691.
- 11 H. J. Wang, Z. X. Lin, X. M. Liu, S. Y. Sheng and J. Y. Wang, *J. Controlled Release*, 2005, **105**, 120.
- 12 H. J. Wang, S. J. Gong, Z. X. Lin, J. X. Fu, S. T. Xue, J. C. Huang and J. Y. Wang, *Biomaterials*, 2007, **28**, 3952.
- 13 Z. H. Qu, H. J. Wang, T. T. Tang, X. L. Zhang, J. Y. Wang and K. R. Dai, *Acta Biomater.*, 2008, **4**, 1360.
- 14 X. M. Liu, Q. S. Sun, H. J. Wang, L. Zhang and J. Y. Wang, *Biomaterials*, 2005, **26**, 109.
- 15 S. J. Gong, H. J. Wang, Q. S. Sun, S. T. Xue and J. Y. Wang, *Biomaterials*, 2006, **27**, 3793.
- 16 P. Argos, K. Pedersen, M. D. Marks and B. A. Larkin, *J. Biol. Chem.*, 1982, **257**, 9984.
- 17 A. S. Tatham, J. M. Field, V. J. Morris, K. J. I'Anson, L. Cardle, M. J. Dufton and P. R. Shewry, *J. Biol. Chem.*, 1993, **268**, 26253.
- 18 M. Abdollahi, M. Rezaei and G. Farzi, *J. Food Eng.*, 2012, **111**, 343.
- 19 D. S. Cha and M. S. Chinnan, *Crit. Rev. Food Sci. Nutr.*, 2004, **44**(4), 223.
- 20 B. Liu and H. C. Zeng, *Langmuir*, 2004, **20**, 4196.
- 21 A. Vyalikh, P. Simon, E. Rosseeva, J. Buder, U. Scheler and R. Kniep, *Sci. Rep.*, 2015, **5**, 15797.
- 22 F. C. Meldrum, *Int. Mater. Rev.*, 2003, **48**(3), 187.
- 23 W. Z. Ostwald, *J. Phys. Chem.*, 1900, **34**, 495.
- 24 M. M. H. Farooqi and R. K. Srivastava, *J. Alloys Compd.*, 2017, **691**, 275.
- 25 R. Kripal, A. K. Gupta, R. K. Srivastava and S. K. Mishra, *Spectrochim. Acta, Part A*, 2011, **79**, 1605.
- 26 K. Raja, P. S. Ramesh and D. Geetha, *Spectrochim. Acta, Part A*, 2014, **131**, 183.
- 27 H. J. Wang, X. J. Yu, Y. Cao, B. Zhou and C. F. Wang, *J. Inorg. Biochem.*, 2012, **113**, 40.
- 28 Y. N. Chang, M. Zhang, L. Xia, J. Zhang and G. Xing, *Materials*, 2012, **5**(12), 2850.
- 29 L. C. Ann, S. Mahmud, S. K. M. Bakhori, A. Sirelkhatim, D. Mohamad, H. Hasan, A. Seenii and R. A. Rahman, *Ceram. Int.*, 2014, **40**(2), 2993.
- 30 Y. W. Baek and Y. J. An, *Sci. Total Environ.*, 2011, **409**(8), 1603.
- 31 S. Sultana, M. Z. Rafiuddin, M. Z. Khan and M. Shahadat, *J. Environ. Chem. Eng.*, 2015, **3**(2), 886.
- 32 H. J. Wang, J. C. Huang, L. Hou, T. Miyazawa and J. Y. Wang, *J. Mater. Sci.: Mater. Med.*, 2016, **27**, 92.
- 33 L. Gutiérrez, A. Escudero, R. Battle and C. Nerin, *J. Agric. Food Chem.*, 2009, **57**, 8564.
- 34 L. Gutierrez, R. Battle, S. Andujar, C. Sanchez and C. Nerin, *Packag. Technol. Sci.*, 2011, **24**, 485.
- 35 U. I. Walther, S. C. Walther and O. Temruck, *Toxicol. In Vitro*, 2007, **21**, 380.
- 36 B. Wihelm, U. I. Walther and B. Fichtl, *Arch. Toxicol.*, 2001, **75**, 388.

