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Monolith-based ⁶⁸Ga processing: a new strategy for purification to facilitate direct radiolabelling methods†

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The post-processing of ⁶⁸Ga generator eluate by means of a novel high capacity cation-exchange silica monolith column has been validated in this work. Quantitative release of a purified ⁶⁸Ga solution in high concentration can be achieved using weak acidic solutions which can be directly used for chelator or conjugate labelling in an injectable form with improved radiochemical characteristics. The system has the potential to be incorporated in a flow based microfluidic system for dose-on-demand radiotracer synthesis.

⁶⁸Ga radiopharmaceuticals are of growing importance in clinical positron emission tomography (PET) with the success of octreotide derivatives for imaging neuroendocrine tumours, 1,2 and prostate specific membrane antigen (PSMA) derivatives receiving significant recent interest.3,4 68Ga generators have simultaneously been developed towards routine clinical use5,6 with the Eckert and Ziegler generator receiving recommendation from the EMA for pharmaceutical approval and FDA documents filed in 2014. This generator is based on a TiO2 solid phase in which the produced ionic ⁶⁸Ga³⁺ is eluted as GaCl₃ from the generator using a solution of 0.1 N HCl.⁷ However, current ⁶⁸Ge/⁶⁸Ga radionuclide generators do not have optimal properties for the direct synthesis of ⁶⁸Ga-labeled radiopharmaceuticals. The generator eluate has a large volume (5-10 mL) with a high concentration of H⁺ (pH < 1) and can contain metal ion impurities (commonly aluminium(III), iron(III) and zinc(II)).^{6,8} All of which can potentially hinder complexation formation and reduce specific activity (SA). Thus, the eluate from a ⁶⁸Ge/ ⁶⁸Ga generator cannot currently be directly used for preparing radiopharmaceuticals that are in routine clinical practice.

This can be overcome by either; (a) advanced chelator design to allow for direct labelling of eluate, a trait which is so far only proven successful with the highly promising tris(hydroxypyridinone) ligands developed by Blower, Ma and co-workers^{9,10} or; (b) post-processing of the generator eluate to allow for complexation, the approach nearly exclusively followed by groups worldwide and the only method possible for current clinically used tracers.

Several methods for the processing of the generator eluate have been attempted in order to overcome the issues mentioned above including ion-exchange chromatography and fractionation.^{7,8,11-13} All of which individually mitigate some of the limitations of ⁶⁸Ga complexation. However, to date, no method delivers on all requirements to release the ⁶⁸Ga purified from other metal ions into small volumes of low H+ concentration using a directly injectable solvent. Rösch and coworkers use a particle-packed cation-exchange resin column and ethanol (instead of the commonly used acetone) in the elution mixture for post-processing of ⁶⁸Ga eluate. ¹⁴ Ethanolbased post-processing provides a more acceptable reaction mixture for the preparation and formulation of directly injectable 68Ga-radiopharmaceuticals. However, the requirement for a higher concentration HCl solution (0.9 N) can suppress the reactivity of chelators and also needs higher concentrations of buffer to balance the pH.

Recently, miniaturised microfluidic-based systems have attracted much attention in PET tracer synthesis. 15,16 Such microfluidic systems are easily shielded and therefore reduce external impact and have significant potential to facilitate PET tracer synthesis, reduce consumption of expensive reagents and improve reactivity. Most importantly, microfluidicbased systems are compatible with automated dose-ondemand requirements. However, it is difficult to incorporate the current post-processing into microfluidic systems because particle-packed resin columns are challenging to integrate with microfluidic systems due to the relative high backpressures generated. This is an obstacle in the design of

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an integrated microfluidic system for the dose-on-demand tracer synthesis.

Owing to their small-size skeletons and mesoporous structures, monolith columns provide elevated permeability and favourable mass transfer compared to particle-packed columns. 17-20 Due to these characteristics monolith columns have shown their applications in a variety of chromatographic modes.^{21–24} Importantly, the integration of monolith columns with microfluidic systems has already been demonstrated in an enzymatic reaction module.25 If a cationexchange monolith can purify ⁶⁸Ga generator eluate, it would therefore be possible to build an integrated system comprising a monolith for ion-exchange and a microfluidic system ⁶⁸Ga performing flow-based preparation radiolabelling in an automated dose-on-demand system. To our knowledge there has been no reported use of a cationexchange monolith for the separation of ⁶⁸Ga directly from the ⁶⁸Ge/⁶⁸Ga generator eluate.

The aim of this work is to fabricate a cation-exchange silica monolith and evaluate its capability for efficient separation of ⁶⁸Ga directly from the generator eluate. The preparation of injectable ⁶⁸Ga-labeling pharmaceuticals can then be investigated to determine whether the characteristics are improved when compared to the best current method.

The silica monolith columns used in this work are porous rods consisting of silica skeletons and interconnecting mesopores that give much higher permeability and favourable mass transfer compared to particle-packed columns. 26,27 They were produced by the sol-gel method described in our previous study26 followed by surface modification with thiol groups and oxidation to transform -SH to -SO₃H (see ESI†). Therefore forming a sulfonic acid functionalised monolith capable of selectively trapping cations. BET nitrogen adsorption/desorption isotherm (see ESI† S5 and Fig. S3) shows typical hysteresis between adsorption and desorption, which is consistent with the disordered mesoporous structure displayed in the SEM (see Fig. 1) giving a specific surface area of 238 m² g⁻¹ and pore volume of 0.75 cm³ g⁻¹, with an average pore diameter of ~13 nm. The cation exchange capacity (CEC) quantifies the ability of a material to adsorb cations. Analysis of our -SO₃H monoliths gives a value of 180 μeq. g⁻¹, which is 30 times higher than a commercial solid phase extraction (SPE) cation-exchange resin (for further CEC discussion see ESI† S4).

Due to shrinkage during the sol-gel processing it is challenging to make a monolith column within a conventional stainless steel column without a void volume. Therefore, commercially available monolith columns are usually confined in heat-shrinkable polymer tubing. 17,28 To avoid damage to the functionalised monolith from the heating required to mount the column tightly inside heating-shrinkable tubing, the SiO2 monolith-SO3H rod was fixed in an empty Sep-Pak Vac tube and a void volume between the monolith rod and tube wall was packed with powdered functionalised monolith (ca. 10% wt of total monolith mass) and plastic frits fitted at both ends to form the SiO2 monolith-SO3H column (see Fig. 1).

As mentioned previously, the direct synthesis of ⁶⁸Ga tracers from the generator eluate is problematic. The most common approach to overcome this is the transfer of the initial ⁶⁸Ga eluate to a cation-exchange resin column. As it has a high cation-exchange distribution coefficient, ⁶⁸Ga can be quantitatively adsorbed on the resin from the acidic generator eluate and then eluted with HCl/acetone solutions.7,29,30 Although the use of HCl/acetone solutions can efficiently purify and concentrate the ⁶⁸Ga eluate, SPE is required for the removal of acetone and the recovery of the ⁶⁸Ga-labeled compounds. The use of ethanol in place of acetone provides the potential for preparation of injectable ⁶⁸Ga-radiopharmaceuticals. However, the reactivity of the ⁶⁸Ga solution released via current methodologies is inadequate for high SA tracer formation due to metal ion impurities and currently used technologies are not compatible with microfluidic reactors, preventing the design of automated compact dose-on-demand systems.

The SiO₂ monolith-SO₃H columns used in this work not only quantitatively (98 ± 1%) adsorbed ⁶⁸Ga³⁺ but also exhibited highly efficient release of the ⁶⁸Ga³⁺ in comparison to commercial polymeric cation-exchange resins. For the elution methods (see Table 1), a multi-fraction elution process showed a small advantage compared to single step method.

Larger amounts of ethanol and hydrochloric acid in the washing solution (i.e. 85% EtOH/0.2 N HCl) result in a modest amount (8%) of ⁶⁸Ga being lost in the washing step. When using 80% EtOH/0.1 N HCl and 90% EtOH/0.5 N HCl

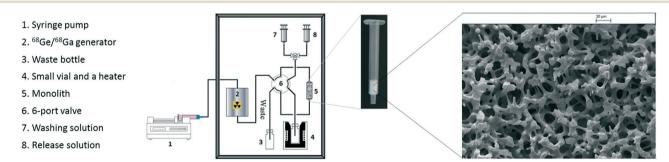


Fig. 1 Schematic representation of the monolith based system for ⁶⁸Ga processing and radiolabelling (left), photograph of cation-exchange silicamonolith column (centre) and SEM image of SiO₂ monolith-SO₃H (right, scale bar: 20 μm).

Table 1 Ethanol-based post-processing of 68 Ga on SiO_2 monolith- SO_3H (average of n=3)

Entry	Activity% ^a	Washing solution ^b	Activity% ^c	Elution solution	Volume	Activity% ^d
1	98	85% EtOH/0.2 N HCl	8.0	93% EtOH/0.9 N HCl	1 mL	90
2	99	80% EtOH/0.1 N HCl	0.3	96% EtOH/0.5 N HCl	1 mL	98
3	99	80% EtOH/0.1 N HCl	0.3	90% EtOH/0.9 N HCl	0.4 mL	94^e
4	99	80% EtOH/0.1 N HCl	0.3	90% EtOH/0.5 N HCl	0.4 mL	93 ^e
5	99	80% EtOH/0.1 N HCl	0.3	90% EtOH/0.5 N HCl	$4 \times 0.2 \text{ mL}$	98
6	99	80% EtOH/0.1 N HCl	0.6	90% EtOH/0.5 N HCl	0.5 mL	98
7	99	80% EtOH/0.1 N HCl	0.6	90% EtOH/0.2 N HCl	1 mL	94
8^f	98	80% EtOH/0.2 N HCl	0.1	90% EtOH/0.9 N HCl	1 mL	46
9^f	98	80% EtOH/0.1 N HCl	0.1	90% EtOH/0.2 N HCl	1 mL	16

^a Activity trapped in monolith. ^b Volume of washing solution was 1 mL. ^c Activity in washing solution. ^d Activity in elution solution. ^e Release activity increases to ≥98% with 1 mL. f Commercial cation-exchange resin (SCX). Solvent compositions are given in v/v.

Table 2 Summary of metal ion impurities before and after processing^a

	$\begin{array}{l} Al^{3+} \\ \left(\mu g \ L^{-1}\right) \end{array}$	$Fe^{3+} \atop \left(\mu g \ L^{-1}\right)$	$\begin{array}{l}Zn^{2^{+}}\\ \left(\mu g\;L^{^{-1}}\right)\end{array}$	$\begin{array}{l} Ti^{4^+} \\ \left(\mu g \ L^{-1}\right) \end{array}$	$\begin{array}{l} Ge^{4^+} \\ \left(\mu g \ L^{-1}\right) \end{array}$
Before	1140	74	320	420	25
After	35	52	6	1	Undetected

^a 80% EtOH/0.1 N HCl and 90% EtOH/0.5 N HCl were used as washing and elution solutions respectively.

as washing and elution solutions respectively, the SiO₂ monolith-SO₃H does not only efficiently purify ⁶⁸Ga from other metal ions (see Table 2) but also almost quantitatively (93%) recovers ⁶⁸Ga from the eluate. This compares to only 25% recovery for the AG50W column using the same conditions.¹⁴

It is postulated that the lower release efficiencies for the resin indicate the strong affinity for ⁶⁸Ga³⁺ within the cationexchange resin pores.31 These results suggest that the microenvironment of the monolith columns used in this work favours ion-exchange processing for adsorption and desorption of ⁶⁸Ga³⁺.

A main benefit of our method is that the ⁶⁸Ga can be concentrated up to ten-fold and the ⁶⁸Ga elution requires a lower concentration of hydrochloric acid (0.5 N) when compared with the best published example.14 Decreasing the concentration of H⁺ allows lower concentration buffers to be used in radiolabelling reactions. Metallic radiolabelling characteristics are generally directly proportional to the concentration of the two reagents in solution.9 Increase of 68Ga concentration in this manner could have significant impact on future tracer production by decreasing the amount of precursor required for synthesis, therefore dramatically increasing the SA. In order to ensure the purified ⁶⁸Ga could still be used in standard labelling protocols and behaved analogously to reported chelator reactions, two common gallium(III) chelators (DOTA and NOTA) were selected for comparison of radiolabelling characteristics (see Table 3 and Scheme 1). The monolithbased system for ⁶⁸Ga processing and labelling is shown in Fig. 1. A yield of 97% was achieved for DOTA (95 °C, 10 min, 5 μM) which offers a two-fold decrease in the concentration required for near quantitative conversion. The results are more pronounced for NOTA, with 95% incorporation at 1 µM (room temperature, 5 min). This is a ten-fold improvement over the reported concentrations of 10 µM giving 80% RCY.9 Finally, a demonstration of clinical applicability was provided by ⁶⁸Ga radiolabelling of the licensed somatostatin targeted agent DOTATOC, which produced near quantitative yields within 10 minutes using standard kit precursor amounts.14

Complete synthesis of ⁶⁸Ga-DOTATOC can be carried out, from elution of the generator to delivery of the tracer, in 25 minutes. If all the generator eluate is used (740 MBq), the tracer can be produced with a final SA of 41 GBq µmol⁻¹,

Table 3 ⁶⁸Ga-radiolabelling of different chelators after post-processing a (n = 3)

Entry	Trap (%)	Release (%)	Labelling conditions				
			Chelator	Concentration (µM)	Temperature (°C)	Time (min)	Yield (%)
1	99	98	DOTA	1	95	10	0
2	99	98	DOTA	5	95	10	97
3	99	98	DOTA	10	95	10	98
4	99	98	DOTA	20	95	5	98^b
5 ^c	99	97	DOTA	10	95	10	99
6	99	98	NOTA	5	RT^d	5	99
7	98	97	NOTA	1	RT^d	5	95
8	99	97	NOTA	0.5	RT^d	5	56
9	99	95	DOTATOC	14	95	10	97
10	99	95	DOTATOC	14	95	15	>99

^a Washing solution was 1 mL of 80% ethanol/0.1 N HCl, elution solution was 0.5 mL of 90% ethanol/0.5 N HCl, buffer was 0.5 mL of NH₄OAc (1 M, pH 5). The 68Ga elution was preheated for 10 min at 95 °C. The monolith was reused (SiO₂ monolith–SO₃H). Room temperature.

Scheme 1 Radiochemical formation of ⁶⁸GaDOTA

significantly higher than commonly reported values of 10-22 GBq μ mol⁻¹.^{32,33}

These improved radiolabelling characteristics and shorter reaction times will permit clinical application following a simple pre-injection dilution of the radiopharmaceutical when using ethanol as a solvent.12 The application of the novel monolith column in this work showed that the ⁶⁸Ga radiolabelling characteristics of the processed solution could be significantly improved, reducing the amount of chelator and the reaction time required. It was also found that the monolith could be reused with no significant reduction in labelling yield observed. This has not been reported for commercial polymeric cation-exchange resin in ethanol-based processing of ⁶⁸Ga generator eluate.

In conclusion, a rapid, simple and efficient processing of TiO₂-based ⁶⁸Ge/⁶⁸Ga generator eluate was developed utilising a novel cation-exchange silica-monolith column. It was demonstrated that with an ethanol/HCl eluent the cationexchange silica-monolith could not only quantitatively recover the ⁶⁸Ga but also efficiently purify the ⁶⁸Ga from other metal ion impurities present in the initial generator eluate. The advantage of monolith-based processing compared to the optimum method reported in literature¹⁴ is a ca. 50% reduction in both the eluent volume and the HCl concentration required. The higher ⁶⁸Ga concentration and purity facilitates efficient radiolabelling with the potential for significant increases in specific activity which could dramatically improve image quality for some tracers. The processed ⁶⁸Ga was directly used for radiolabelling of commonly used chelators DOTA and NOTA, showing significant improvement in reactivity compared to other ⁶⁸Ga³⁺ purification methods. Comparative radiolabelling of DOTATOC was performed to demonstrate the utility of this method with a clinically relevant example. The low back pressure of monolith based systems compared to polymeric cation-exchange resins allows, for the first time, the possibility to efficiently integrate an ionexchange component with a microfluidic reactor and the production of automated modules for continuous ⁶⁸Ga elution, preparation and labelling. These applications and integrated synthesis unit designs are under investigation.

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Notes and references

- 1 M. Hofmann, H. Maecke, A. R. Borner, E. Weckesser, P. Schoffski, M. L. Oei, J. Schumacher, M. Henze, A. Heppeler, G. J. Meyer and W. H. Knapp, Eur. J. Nucl. Med., 2001, 28, 1751-1757.
- 2 I. Velikyan, Theranostics, 2014, 4, 47-80.
- 3 A. Afshar-Oromieh, A. Malcher, M. Eder, M. Eisenhut, H. G. Linhart, B. A. Hadaschik, T. Holland-Letz, F. L. Giesel, C. Kratochwil, S. Haufe, U. Haberkorn and C. M. Zechmann, Eur. J. Nucl. Med. Mol. Imaging, 2013, 40, 486-495.
- 4 M. Eiber, T. Maurer, M. Souvatzoglou, A. J. Beer, A. Ruffani, B. Haller, F.-P. Graner, H. Kuebler, U. Haberhorn, M. Eisenhut, H.-J. Wester, J. E. Gschwend and M. Schwaiger, J. Nucl. Med., 2015, 56, 668-674.
- 5 M. Fani, J. P. Andre and H. R. Maecke, Contrast Media Mol. Imaging, 2008, 3, 67-77.
- 6 B. P. Burke, G. S. Clemente and S. J. Archibald, J. Labelled Compd. Radiopharm., 2014, 57, 239-243.
- 7 K. P. Zhernosekov, D. V. Filosofov, R. P. Baum, P. Aschoff, H. Bihl, A. A. Razbash, M. Jahn, M. Jennewein and F. Rösch, J. Nucl. Med., 2007, 48, 1741-1748.
- 8 A. A. Larenkov, A. B. Bruskin and G. E. Kodina, Radiochemistry, 2014, 56, 57-65.
- 9 D. J. Berry, Y. Ma, J. R. Ballinger, R. Tavare, A. Koers, K. Sunassee, T. Zhou, S. Nawaz, G. E. D. Mullen, R. C. Hider and P. J. Blower, Chem. Commun., 2011, 47, 7068-7070.
- 10 M. T. Ma, C. Cullinane, C. Imberti, J. B. Torres, S. Y. A. Terry, P. Roselt, R. J. Hicks and P. J. Blower, Bioconjugate Chem., 2016, 27, 309-318.
- 11 W. A. P. Breeman, M. de Jong, E. de Blois, B. F. Bernard, M. Konijnenberg and E. P. Krenning, Eur. J. Nucl. Med. Mol. Imaging, 2005, 32, 478-485.
- 12 J. Seemann, E. Eppard, B. P. Waldron, T. L. Ross and F. Rösch, Appl. Radiat. Isot., 2015, 98, 54-59.
- 13 G. J. Meyer, H. Macke, J. Schuhmacher, W. H. Knapp and M. Hofmann, Eur. J. Nucl. Med. Mol. Imaging, 2004, 31, 1097-1104.
- 14 E. Eppard, M. Wuttke, P. L. Nicodemus and F. Rösch, J. Nucl. Med., 2014, 55, 1023-1028.
- 15 P. Y. Keng, S. Chen, H. Ding, S. Sadeghi, G. J. Shah, A. Dooraghi, M. E. Phelps, N. Satyamurthy, A. F. Chatziioannou, C.-J. C. J. Kim and R. M. van Dama, Proc. Natl. Acad. Sci. U. S. A., 2012, 109, 690-695.
- 16 V. Arima, G. Pascali, O. Lade, H. R. Kretschmer, I. Bernsdorf, V. Hammond, P. Watts, F. De Leonardis, M. D. Tarn, N. Pamme, B. Z. Cvetkovic, P. S. Dittrich, N. Vasovic, R. Duane, A. Jaksic, A. Zacheo, A. Zizzari, L. Marra, E. Perrone, P. A. Salvadori and R. Rinaldi, Lab Chip, 2013, 13, 2328-2336.
- 17 K. Cabrera, J. Sep. Sci., 2004, 27, 843-852.
- 18 N. Tanaka, H. Kobayashi, N. Ishizuka, H. Minakuchi, K. Nakanishi, K. Hosoya and T. Ikegami, J. Chromatogr. A, 2002, 965, 35-49.
- 19 M. D. Tarn, D. Maneuski, R. Alexander, N. J. Brown, V. O'Shea, S. L. Pimlott, N. Pamme and S. J. Archibald, Chem. Commun., 2016, 52, 7221-7224.

- 20 J. H. Smith and H. M. McNair, J. Chromatogr. Sci., 2003, 41, 209-214.
- 21 E. Klodzinska, D. Moravcova, P. Jandera and B. Buszewski, J. Chromatogr. A, 2006, 1109, 51-59.
- 22 C. Legido-Quigley, N. D. Marlin, V. Melin, A. Manz and N. W. Smith, Electrophoresis, 2003, 24, 917-944.
- 23 M. P. Taggart, M. D. Tarn, M. M. N. Esfahani, D. M. Schofield, N. J. Brown, S. J. Archibald, T. Deakin, N. Pamme and L. F. Thompson, Lab Chip, 2016, 16, 1605-1616.
- 24 F. Svec, J. Sep. Sci., 2005, 28, 729-745.
- 25 P. He, J. Davies, G. Greenway and S. J. Haswell, Anal. Chim. Acta, 2010, 659, 9-14.
- 26 P. D. I. Fletcher, S. J. Haswell, P. He, S. M. Kelly and A. Mansfield, J. Porous Mater., 2011, 18, 501-508.

- 27 K. Nakanishi and N. Soga, J. Non-Cryst. Solids, 1992, 139, 1-13.
- 28 P. He, S. J. Haswell, P. D. I. Fletcher, S. M. Kelly and A. Mansfield, J. Flow Chem., 2012, 2, 47-51.
- 29 J. S. Fritz and T. A. Rettig, Anal. Chem., 1962, 34, 1562-1566.
- 30 F. W. E. Strelow, A. H. Victor, C. R. Vanzyl and C. Eloff, Anal. Chem., 1971, 43, 870-876.
- 31 S. D. Chambers, K. M. Glenn and C. A. Lucy, J. Sep. Sci., 2007, 30, 1628-1645.
- 32 M. Luisa Soto-Montenegro, S. Pena-Zalbidea, J. Maria Mateos-Perez, M. Oteo, E. Romero, M. Angel Morcillo and M. Desco, PLoS One, 2014, 9.
- 33 Y. Menda, L. L. B. Ponto, M. K. Schultz, G. K. D. Zamba, G. L. Watkins, D. L. Bushnell, M. T. Madsen, J. J. Sunderland, M. M. Graham, T. M. O'Dorisio and M. S. O'Dorisio, Pancreas, 2013, 42, 937-943.