Genetically encoded fluorophenylalanines enable insights into the recognition of lysine trimethylation by an epigenetic reader†

Yan-Jiun Lee, a M. J. Schmidt, b Jeffery M. Tharp, a Annmarie Weber, b Amber L. Koenig, c Hong Zheng, d Jianmin Gao, d Marcey L. Waters, c Daniel Summerer* e and Wenshe R. Liu* a

Fluorophenylalanines bearing 2–5 fluorine atoms at the phenyl ring have been genetically encoded by amber codon. Replacement of F59, a phenylalanine residue that is directly involved in interactions with trimethylated K9 of histone H3, in the Mpp8 chromodomain recombinantly with fluorophenylalanines significantly impairs the binding to a K9-trimethylated H3 peptide.

Due to the size similarity between hydrogen and fluorine atoms, most fluorinated amino acids closely resemble their canonical counterparts. When provided in nutrients, they are usually mistaken as canonical amino acids by the cellular translation system and integrated into proteins at corresponding amino acid sites, leading to mild to severe cellular toxicities.1 Biochemists have long been exploiting this promiscuity of the cellular translation system to generate fluorinated proteins.2 Owing to its high NMR signal that is sensitive toward surrounding environments, fluorine in proteins provides a unique probe to study protein structure and dynamics.3 Fluorine also has a more hydrophobic nature than hydrogen, which endows fluorinated proteins with unique features such as high resistance to denaturants.4 Although convenient in making fluorinated proteins, this residue-specific noncanonical amino acid (ncAA) mutagenesis approach typically leads to partial replacement of a native amino acid as a fluorinated amino acid at multiple sites in a protein and generates a highly heterogeneous final product that leads to complexity in subsequent studies. Consequently, methods for the synthesis of proteins bearing fluorinated amino acids at user-defined sites are of high interest. A successful strategy at this front is the use of amber suppression that in the context of fluorinated amino acids was pioneered by Furter and later expanded by others.5

We previously reported two rationally designed, polyspecific mutants of Methanosarcina mazei pyrrolysyl-tRNA synthetase (PyIRS) that enable the effective aminoacylation of tRNA with a large variety of phenylalanine-derived ncAA for their incorporation into proteins by amber suppression. Of these, mutant N346A/C348A/Y306A/Y384F (PyIRS-AAAF) accepted phenylalanine derivatives with large substituents at the para position as substrates,6 whereas N346A/C348A (PyIRS-AA) accepted small phenylalanine derivatives,7 including the NMR probe m-trifluoromethyl-phenylalanine.

To get insights into the origin of polyspecificity of PyIRS-AA and into its lack of phenylalanine recognition, we determined the crystal structure of the C-terminal catalytic fragment (amino acids 188–454)8 in complex with the ATP analog adenosine-5′-(β,γ-imido)triphosphate (AMPPNP) at a resolution of 1.5 Å (the structure of PyIRS-AAAF was also solved, for data collection and refinement statistics, see the ESI†).9 In wild type PyIRS (PyIRS_wt), N346 of the amino acid binding pocket serves as a gatekeeper residue that is engaged in a variety of direct and water-mediated hydrogen bonds (Fig. 1A). This includes donation of one bond to the N-carbonyl group of pyrrolysine adenylate, which is believed to be critical for pyrrolysine recognition.10 C348 forms part of the pocket bottom (Fig. 1A). In the structure of PyIRS-AA, the mutation of both residues to alanine results in an inability of hydrogen bonding, combined with an enlargement of the pocket, in particular at position 346 (Fig. 1B). A related observation has been made in the context of a different mutant that accepts O-methyl-tyrosine as a substrate.11

PyIRS belongs to the aminoaeryl-tRNA-synthetase subclass Iic that also includes PheRS from that PyIRS has directly evolved, and both show similarity in their overall fold and in the organization of the core domain (Fig. 1C).9 Our structure of PyIRS-AA reveals similarities with PheRS in the front pocket dimension and polarity, despite marked differences in the type and orientation of the involved residues (Fig. 1D and E). However, the overall pocket dimensions of PyIRS-AA (in particular in the rear part) are larger, and in superimposed structures,
both the Phe ligand and the pocket surface of PheRS can easily be accommodated in the pocket of PyrRS-AA (Fig. 1E). Specifically, in PheRS, residues including E210, F248, F250, and A294 form a very compact binding pocket for phenylalanine (Fig. 1D). Consequently, though PheRS tolerates only unsubstituted Phe may account for the observed selectivity of PyrRS-AA (Fig. 5, ESI†).

This structural comparison intrigued us to test the recognition of pentafluoro-phenylalanine (F5F, Fig. 2A) by PyrRS-AA. As expected, PyrRS-AA does recognize F5F. E. coli BL21 cells coding PyrRS-AA, tRNA51, and sfGFP2TAG (superfolder green fluorescent protein (sfGFP) with an amber mutation at its S2 position) expressed full-length sfGFP when F5F was provided in the GMML medium, albeit with a low level (Fig. S1, ESI†). In order to identify a PyrRS mutant that in coordination with tRNA51 shows an enhanced amber suppression rate in E. coli for more efficient incorporation of F5F into proteins, we constructed a small PyrRS-AA-based mutant library by randomizing A348. A348 is spatially close to E174 in PheRS that locks phenylalanine restrictedly at the PheRS active site. By randomizing A348, we deemed that a better mutant with tighter binding of F5F could be identified. Screening all mutants led to the final identification of the mutant with S348 (coined as PyrRS-AS) that in coordination with tRNA51 provided a higher efficiency of amber suppression in E. coli in the presence of F5F (Fig. S2 and S3, ESI†).

To test the fidelity of PyrRS-AS for the genetic incorporation of F5F in response to the amber codon, E. coli BL21 cells coding PyrRS-AS, tRNA51, and sfGFP2TAG were grown in GMML medium with or without supplementing F5F. Cells grown in the presence of F5F produced full-length sfGFP (sfGFP-F5F) with an expression level of 10 mg L⁻¹, markedly contrasting to a negligible expression of full-length sfGFP in the absence of F5F (Fig. 2B). This demonstrated that PyrRS-AS accepts F5F as substrate but discriminates against canonical amino acids including Phe. Electrospray ionization mass spectrometry (ESI-MS) analysis of the purified sfGFP-F5F displayed a molecular weight of 27 817 Da that agreed well with the theoretical mass of 27 819 Da. The single dominant ESI-MS peak also indicated that F5F was not recognized...
by *E. coli* PheRS, which would lead to replacement of 12 Phe residues in sfGFP during translation. Therefore the genetic encoding of F₃F by amber codon is orthogonal to the endogenous translation system.

We next tested the ability of PyrRS-AS for the acceptance of other fluorophenylalanines including 2,3,4,5-tetrafluorophenylalaine (F₄F), 3,4,5-trifluorophenylalanine (F₃F), 3,5-difluorophenylalanine (F₂F), and 3,4-difluorophenylalanine (F₂F'). When these fluorophenylalanines were present in the growth medium, *E. coli* BL21 cells coding for PyrRS-AS, tRNAₐ₅⁵ and sfGFP2TAG expressed full-length sfGFP (Fig. 2B). Expression levels under these conditions were similar to the condition with F₂F. Molecular weights of purified full-length sfGFP proteins expressed in presence of F₂F, F₃F, and F₄F (sfGFP-F₂F, sfGFP-F₃F, and sfGFP-F₄F, respectively) determined by ES-MS agreed well with theoretical molecular weights of these proteins (Fig. 2C and Table 1). All three proteins exhibited a single dominant ES-MS peak, establishing the orthogonality of genetic ncAA incorporation in respect to the endogenous translation system. However, the full-length sfGFP with F₂F incorporated (sfGFP-F₂F') displayed multiple peaks in its ES-MS spectrum. The smallest peak at 27 765 Da matched the theoretical mass of 27 763 Da. However, other peaks were all about multiples of 36 Da addition to the theoretical mass, clearly indicating that F₂F' displaced regular phenylalanine residues in sfGFP. This result demonstrated that the genetic encoding of F₂F' by amber codon is not orthogonal to the endogenous translation system, although PyrRS-AS does recognize it as a substrate.

In addition to being used as a NMR probe and to improve protein folding, genetically encoded fluorophenylalanines in proteins could potentially be for the investigation of cation–pi interactions such as in the recognition of lysine methylation in histones by epigenetic readers. Being part of epigenetic regulation of histone function, histone lysine methylation induces interactions with effector proteins and subsequently regulates DNA replication, repair, and transcription. The recognition of methylated lysine typically involves an aromatic cage that has been found in the chromodomain (Fig. 3A), the PHD finger, and the Tudor domain, and appears to be mediated by cation–pi interactions between the methylammonium moiety and aromatic residues in the cage. The cation–pi interaction is predominantly electrostatic, occurring between a cation and the quadrupole moment of an aromatic π system (Fig. 3B). As the quadrupole moment places partial negative charge above each face of the aromatic ring, favorable interactions with a cation occur perpendicular to the aromatic plane within a typical van der Waals distance.

Although a number of theoretical and experimental studies have been carried out to address the importance of the cation–pi interaction in the recognition of lysine methylation, it is not clear to what degree the cation–pi interaction contributes to the recognition specificity. A particularly interesting target protein to address this question is the Mpp8 chromodomain (Mpp8C). Mpp8 is a heterochromatin component that specifically recognizes and binds trimethylated K9 of histone H3 and promotes recruitment of proteins that mediate epigenetic repression. In Mpp8C, F59 is part of the aromatic cage that directly binds to trimethylated K9 of H3. Replacing this residue with fluorophenylalanines (in particular with F₅F that has a strongly reduced partial negative charge above each face of the aromatic side chain) is expected to significantly reduce the binding of Mpp8C to trimethylated K9 of H3 in the case that the cation–pi interaction plays a dominant role. Otherwise, binding would not be strongly affected or might increase due to the more hydrophobic nature of fluorophenylalanines than phenylalanine.

Using our currently developed approach, Mpp8C with F₅9 replaced by the three derivatives F₂F, F₃F, and F₄F were expressed. The incorporation of F₂F in Mpp8C was independently confirmed with the detection of three ¹⁹F NMR singals in the finally purified protein (Fig. S6, ESI†). Together with wild type Mpp8C, interactions of these proteins with a fluorescein-conjugated N-terminal histone H3 peptide with trimethylation at the K9 position (FAM-H3(1-15)K9me3) were studied using fluorescent polarization changes. As shown in Fig. 3C and Table 2, wild type Mpp8C interacts with FAM-H3(1-15)K9me3 strongly, with a determined Kₐ value around 0.8 μM that agrees...
Table 2  Determined dissociation constants between MPP8C proteins and FAM-H3(1-15)K9me3

<table>
<thead>
<tr>
<th></th>
<th>$K_d$ (μM)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Wild type MPP8C</td>
<td>0.8 ± 0.1</td>
</tr>
<tr>
<td>MPP8C-F59F_F</td>
<td>12 ± 5</td>
</tr>
<tr>
<td>MPP8C-F59F_F</td>
<td>&gt;100</td>
</tr>
<tr>
<td>MPP8C-F59F_F</td>
<td>&gt;200</td>
</tr>
</tbody>
</table>

with previously reported values. This binding was decreased 15-fold when F59 was replaced with F3F and continued to drop when F59 was replaced with F4F and F5F (Fig. 3D–F and Table 2). Due to the low binding of FAM-H3(1-15)K9me3 to both F59F and F59F mutants of MPP8C, no sufficient data could be collected to determine accurate $K_d$ values between these two proteins and FAM-H3(1-15)K9me3. This continuous decrease of binding of Mpp8C to FAM-H3(1-15)K9me3 when a growing number of fluorine substituents are added to F59 strongly suggests that the cation–pi interaction plays a dominant role in the binding of trimethylated K9 of H3 to Mpp8C. Though hydrophobic interactions may contribute to the binding, they appear to be not significant, since adding hydrophobicity to F59 does not improve binding.

In summary, a method for the genetic incorporation of fluorophenylalanines with fluorine substituents at the side chain phenyl ring ranging from 2 to 5 has been developed. This was based on a polyspecific PyrRS mutant, its crystal structural analysis, and its further reengineering. The engineered PyrRS mutants display recognition of fluorophenylalanines and discriminate against canonical amino acids including phenylalanine, assuring their specific incorporation in response to the amber codon. Using this method, we synthesized Mpp8C, a chromodomain with fluorophenylalanines replacing the critical active site residue F59 that directly interacts with trimethylated K9 of H3 for its binding to Mpp8C. We showed that replacing F59 with fluorophenylalanines significantly weakens the binding of Mpp8C to trimethylated K9 of H3. This result strongly supports a critical involvement of the cation–pi interaction in the recognition of lysine trimethylation by a chromodomain.

This work was supported by National Institute of Health (grants CA161158 to WSL and GM102735 to JG), National Science Foundation (grant CHE-1148684 to WSL, CHE-1112188 to JG, and CHE-1309777 to MLW, and DGE-1144081 to ALK), Welch Foundation (grant A-1715 to WSL), and the Deutsche Forschungsgemeinschaft (SU 726/6-1 in SPP1623).

Notes and references