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Preparation of nanosized coacervates of positive and negative starch derivatives intended for pulmonary delivery of proteins†

S. Barthold,^{ab} S. Kletting,^{ab} J. Taffner,^a C. de Souza Carvalho-Wodarz,^a E. Lepeltier,^a B. Loretz*^a and C.-M. Lehr^{ab}

Proteins and peptides represent a large fraction of the compounds currently in drug development pipelines. Their application however often depends on the use of carrier systems. Nanoparticles (NPs) are widely used such carrier systems for protein delivery. The aim of this study was to design a new drug delivery system (DDS), prepared under mild conditions in aqueous solution without the requirement of a stabilizer. The biodegradability and biocompatibility of the designed system was explored with a view to specifically determine its potential to facilitate the pulmonary delivery of proteins. As a first step, anionic and cationic water soluble starch-derivatives were synthesized. These starch polymers allowed for NP formation via coacervation, as well as protein loading. Physicochemical characterization of the prepared NPs was then carried out: NPs were found to have a narrow size distribution with an average size ranging from 140 to 350 nm, and a ζ -potential ranging from -10 to -35 mV, depending on the formulation conditions. In a proof of concept study, starch NPs were found to be readily degraded by the human enzyme α -amylase, and showed good biocompatibility with A549 cells after 4 h. Upon nebulization, NPs were seen to be internalized by air-liquid interface cultivated A549 cells as well as 16HBE14o⁻ cells. To evaluate the ability of starch NPs to load proteins of various characteristics, NPs were loaded with four model proteins/peptides possessing different molecular weights and isoelectric points – IgG1, RNase A, insulin, and vancomycin. The greatest loading was achieved in the case of vancomycin with up to 23% drug loading and 43% encapsulation efficiency, indicating an optimal loading of proteins with an isoelectric point close to the pH of the NP suspension. In conclusion, starch NPs prepared by the developed mild and straightforward technique show potential as a safe platform for pulmonary delivery of proteins and peptides.

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1 Introduction

Proteins and peptides represent a large fraction of the compounds currently in drug development pipelines,^{1,2} due to their high selectivity and predictable activity profiles. However, the successful delivery of such biopharmaceuticals is anything but easy.

^a Department Drug Delivery, Helmholtz-Institute for Pharmaceutical Research Saarland (HIPS), Helmholtz-Centre for Infection Research (HZI), Saarbrücken, Germany. E-mail: Brigitta.Loretz@helmholtz-hzi.de

^b Department of Pharmacy, Saarland University, Saarbrücken, Germany

† Electronic supplementary information (ESI) available: Fig. S1: ¹H-NMR of starch derivatives in D₂O. Fig. S2: GPC analysis of starch derivatives. Fig. S3: XRPD pattern of starch derivatives. Fig. S4: Stability study of starch NPs at RT or in the fridge. Fig. S5: FT-IR of unlabeled PosSt (black) and labeled PosSt_F. Fig. S6: Confocal laser scanning microscopy (CLSM) images of 250 μ L nebulized starch NP onto A549 cultivated under ALI. Incubation was at 4 °C for 1 h (A) or 4 h (B). Table S1: Characteristics of starch NPs prepared with different amounts of labeled PosSt_F in a molar ratio of 1 : 1 (PosSt_F : NegSt) and a concentration of 0.25 mg mL⁻¹. See DOI: 10.1039/c6tb00178e

The oral route of administration for example, although well-known and successful for small molecule drugs, presents significant challenges in the case of protein delivery. In contrast to small molecule drugs, proteins are characterized by a complex (and degradation-prone) secondary and tertiary structure, which is vital for their biological activity; proteins also possess a high molecular weight, meaning a low permeability across biological barriers.^{3,4} As a result, most protein and peptide formulations available on the market today are administered intravenously to overcome difficulties associated for example with oral delivery, such as enzymatic degradation and poor permeability. However, modern pharmaceutical technology has made significant progress in the design of non-invasive drug delivery strategies that act to stabilize protein during formulation and storage, and to maintain efficacy of the protein throughout the delivery process itself.⁵⁻⁷

Besides the oral route, one of the most promising non-invasive routes of administration is the pulmonary route.



Due to reduced systemic side effects and a rapid onset of action, administration *via* the lungs represents a valuable means of local drug delivery for treatment of diseases such as asthma, cystic fibrosis and lung cancer.

The use of nanoparticles (NPs) as drug delivery systems (DDS) for protein delivery is well-known. NPs offer several advantages over conventional therapeutic systems. The incorporation of proteins into NPs can be beneficial for their stability and can offer protection during storage and administration. Moreover, NPs are known to facilitate an enhanced cellular uptake of incorporated or associated actives, which could also be beneficial in the case of protein permeation through epithelial barriers.⁸

While several polymers have been investigated for use as components of DDS, the need for natural and semi-synthetic polymers with beneficial properties specifically for the delivery of proteins is increasing. PLGA for example has been widely utilized; however, although biodegradable mainly by hydrolytic degradation, the use of such a polymer in the case of a protein-DDS could prove detrimental to the encapsulated cargo, due to high generated levels of acidic degradation products.⁹ In addition to the commonly used PLGA, polysaccharides are among the most studied polymers for drug delivery applications.¹⁰ Alginate, for example, can form NPs of a wide size range, based on a controlled gelification process of alginate by calcium ions.¹¹ Chitosan-based systems are also well explored and characterized.^{12–16} Here again a polymer (chitosan) is interacting with a small ionic crosslinker, tripolyphosphate (TPP). Although chitosan is enzymatically degradable by chitosanase,^{17,18} this is not an enzyme that is ubiquitous in the human body. Starch, in contrast, is biocompatible, non-toxic, and readily degraded by α -amylase, a human enzyme. As such, it offers great potential as an excipient for DDS preparation. Moreover, the high amount of hydroxyl groups present within starch allows for different derivatizations. The use of propyl starch for the encapsulation of small compounds was for example studied by Santander-Ortega.¹⁹ Particles in this work were prepared by an emulsification diffusion technique; however, the use of ethyl acetate and a high speed homogenizer makes this preparation method challenging for the encapsulation of proteins, which are prone to degradation under these conditions. Another study from Yamada *et al.* has shown the formation of polyplexes from positively charged PEI-derivatized starch with pDNA, for improved transfection.²⁰ Thiele *et al.* investigated the formulation of NPs for targeted cancer therapy by the coacervation of a derivatized anionic starch copolymer, linked with pteric acid as cell-specific ligand, and a cationic cyclodextrin. In this work, the small ionic crosslinker as used for the systems of alginate and chitosan was replaced by a second polymer.²¹

The aim of this work was to prepare NPs *via* coacervation in aqueous medium from two oppositely charged starch derivatives, in order to take advantage of the excellent biocompatibility and biodegradability of these natural polymers. An anionic and a cationic starch derivative were first synthesized, as such compounds were hypothesized to readily facilitate the subsequent formation of NPs, suitable for loading of proteins with a range of sizes and charges.

2 Experimental section

Materials

Partially hydrolyzed potato starch (M_w 1 300 000 g mol⁻¹, approx. 33% amylose content) was a gift from AVEBE (Netherlands). Negatively (NegSt) and positively (PosSt) charged starches were synthesized with a weight average molecular weight of 87 000 and 76 000, respectively. IgG1 and insulin were kindly donated by Boehringer Ingelheim (Germany) and Sanofi (France), respectively. Vancomycin hydrochloride (vancomycin), Ribonuclease A from bovine pancreas (RNase A), α -amylase from porcine pancreas and thiazolyl blue tetrazolium bromide (MTT reagent) were bought from Sigma Aldrich (USA). Bodipy[®] FL-C5 NHS Ester (succinimidyl ester), cell culture medium (RPMI 1640, minimum essential medium eagle (MEM)), fetal bovine serum (FBS), phosphate buffered saline (PBS) and Hank's balanced salt solution (HBSS) were obtained from Life Technologies (USA). Non-essential amino acids (NEAA) were sourced from GE Healthcare Life-Sciences. A549 cells (No. CCL-185) were purchased from American Type Culture Collection. 16HBE14o⁻ cells were a kind gift from Dr Dieter C. Gruenert (Department of Otolaryngology, University of California, San Francisco, CA, USA). Purified water was produced by a Milli-Q water purification system (Merck Millipore, USA). All other reagents were of analytical grade.

Synthesis of starch derivatives

Negatively charged starch (NegSt) was synthesized *via* oxidation of potato starch.^{21,22} Briefly, 20 g of dried potato starch was suspended in 800 mL of purified water and heated at 95 °C for one hour. After cooling down to room temperature (RT), TEMPO ((2,2,6,6-tetramethylpiperidin-1-yl)oxidanyl; 4 mg g⁻¹ potato starch) was added to the suspension. 112 mL of sodium hypochlorite solution (12%) was then added (1 mL min⁻¹). During the addition of the sodium hypochlorite solution, the pH was kept at 8.5 by adding 1 M sodium hydroxide. To stop the reaction, sodium borohydride was slowly added. The resulting solution was stirred overnight and purified by ultrafiltration (Vivaflow 200; 5 kDa Hydrosart membrane, Sartorius Stedim Biotech GmbH, Germany). The product was dried by lyophilization (Alpha 2-4, Martin Christ GmbH, Germany). For the synthesis of PosSt, 13 g of NegSt (free acid form) was dissolved in 650 mL DMSO at 50 °C under a reflux condenser, with stirring overnight. Coupling reagents *N,N'*-dicyclohexylcarbodiimide (DCC) and 1-hydroxybenzotriazole (HOBt; each 1.5 molar equivalents, according to the degree of oxidation of NegSt) were then added and stirred for another 24 h. Subsequently, 10 molar equivalents of ethylenediamine (according to the degree of oxidation of NegSt) were slowly added. The reaction was continued for 48 h at 50 °C, with stirring. After filtration, the solution was lyophilized, subsequently dissolved in water, subject to extensive ultrafiltration (Vivaflow 200; 5 kDa Hydrosart membrane, Sartorius Stedim Biotech GmbH, Germany) and again lyophilized.

Characterization of starch derivatives

Fourier transform infrared (FT-IR) measurements were carried out in solid state with a Spectrum 400 FT-IR/FT-NIR spectrometer



(PerkinElmer, UK) between 600 and 4000 cm^{-1} . $^1\text{H-NMR}$ spectra were collected at room temperature on a Bruker Biospin spectrometer NMR Magnet System 400 MHz Ultrashield plus (Bruker, USA) in D_2O (64 scans). The weight average molecular weights of starch derivatives were analysed by gel permeation chromatography (GPC, HLC-8320 GPC, Tosoh, Japan), equipped with online viscometer (ETA-2010, PSS, Germany) on SUPREMA 1000 and 30 columns (NegSt) or SUPREMA-MAX 1000 and 30 columns (PosSt) (PSS, Germany). A flow rate of 1 mL min^{-1} at 35°C in 1 M sodium nitrate (NegSt) or 1% formic acid (PosSt) was employed. Calibration was performed with Pullulan and PVP standards. The degree of oxidation of NegSt was determined with the Blumenkrantz assay, a colorimetric method which detects uronic acid content.²³ X-ray scans were taken by a diffractometer of the type Bruker D8 Advance, equipped with a 1D-detector 'Lynxeye' and variable divergence slit, using $\text{CuK}\alpha$ radiation.

Preparation of starch nanoparticles

Starch NPs were prepared by charge-mediated coacervation between positively charged amine groups (PosSt) and negatively charged carboxylate groups (NegSt) of starch derivatives in aqueous solution. Briefly, materials were dissolved in purified water to obtain solutions of 1 mg mL^{-1} , 0.5 mg mL^{-1} and 0.25 mg mL^{-1} and filtered. Different molar ratios (1 : 3, 1 : 1, 3 : 1) of NegSt and PosSt solutions were prepared by adding adequate volumes of NegSt (1, 0.5, 0.25 mg mL^{-1}) solution to PosSt solution (1, 0.5, 0.25 mg mL^{-1}) under gentle stirring at RT. NPs formed spontaneously and were analyzed after 15 min of stirring for equilibration. The final pH value of all tested formulations was 7.4.

Physicochemical characterization of nanoparticles

Particle size, size distribution (PdI), and ζ -potential of starch NPs in preparation medium were determined using the Zetasizer Nano ZSP (Malvern Instruments, UK) with a scattering angle of 173° . Particle sizes were intensity based z -average values and standard deviation was of 3 measurements. Morphology of NPs was analyzed by transmission electron microscopy (TEM) using a JEOL JEM 2011 microscope (Oxford Instruments, UK). $20 \mu\text{L}$ of sample was incubated on a copper grid for 30 min. After positive staining with 1% (w/v) of phosphotungstic acid (PTA) for 30 s, samples were dried overnight (excess fluid was directly removed with filter paper) and analyzed. Physicochemical stability of NPs was monitored with respect to size, PdI and ζ -potential for 14 days. Samples were stored either at RT (21°C) or in the fridge (4°C).

Biodegradability of starch nanoparticles

To study the biodegradation of starch NPs, 10 mL of NP suspension was incubated with $25 \mu\text{L}$ α -amylase solution ($36.37 \text{ units per } \mu\text{L}$) at 37°C for 2 h, 4 h, 6 h, and 8 h under gentle stirring. As control, 10 mL of starch NP suspension was incubated at 37°C under gentle stirring. Particle concentrations were then analyzed by nanoparticle tracking analysis (NanoSight LM 10, Malvern Instruments, UK). For the measurement, 0.5 mL was taken from each sample. Results shown are the particle concentration (after correction for the dilution

factor needed for valuable measurement) of treated and non-treated samples (each $n = 3$), respectively.

Synthesis of fluorescently labeled PosSt (PosSt_F)

For uptake studies of starch NPs into A549 and 16HBE140⁻ cells, PosSt was labeled with Bodipy[®] FL-C5 NHS Ester. Briefly, 30 mg of polymer was dissolved in 5 mL purified water and diluted slowly with 5 mL of methanol. 2 mL of Bodipy[®] FL-C5 NHS Ester solution in methanol (0.5 mg mL^{-1}) was added slowly to obtain a molar ratio starch : dye of 50 : 1. The sample was stirred for one hour at RT under light protection and precipitated afterwards with 1 M sodium hydroxide and ethanol. Washing was performed with ethanol:methanol (1 : 1) until the supernatant did not show any fluorescence. Coupled starch was then dissolved in purified water and freeze dried.

Nanoparticle preparation with fluorescently labeled PosSt

For uptake studies, solutions of 0.25 mg mL^{-1} and a molar ratio of 1 : 1 of PosSt:NegSt were chosen. Rather than PosSt alone, a 1 : 1 mixture of PosSt_F:PosSt was used for particle preparation.

Stability of nanoparticles upon aerosolization

To study the aerosolization behaviour of starch NPs, starch 0.25–1 : 1 was nebulized with the AeronebLab nebulizer (Aerogen Ltd, Ireland) and collected to be analyzed for size, PdI and ζ -potential with the Zetasizer Nano ZSP (Malvern Instruments, UK).

Cell culture

A549 cells were cultivated in RPMI 1640 medium containing 10% FBS (v/v). For the conduction of cell experiments, A549 cells were seeded into 96 well plates (cell viability assay) or Transwell[®] plates ($3460, 1.12 \text{ cm}^2$; uptake studies). To test the uptake also with a non-cancer cell line of bronchial origin, 16HBE140⁻ cells were used. These cells were cultivated in MEM medium containing 10% FBS (v/v), 1% NEAA and 300 mg of glucose. 1% (v/v) Penicillin/Streptomycin (P/S, $10\,000 \text{ U mL}^{-1}$, Gibco life technologies, USA) was added to each medium before cell viability and uptake studies.

Cell viability studies

Cell viability was evaluated by using the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT)-assay. 2×10^5 cells were seeded into 96 well plates. The assay was performed at 90% confluence. Cell medium was removed and cells were washed once with $200 \mu\text{L}$ HBSS (composed of 1.12 mM CaCl_2 , $0.49 \text{ mM MgCl}_2 \cdot 6\text{H}_2\text{O}$, $0.41 \text{ mM MgSO}_4 \cdot 7\text{H}_2\text{O}$, 5.33 mM KCl , $0.44 \text{ mM KH}_2\text{PO}_4$, 4.17 mM NaHCO_3 , 137.93 mM NaCl , $0.34 \text{ mM Na}_2\text{HPO}_4$, $5.55 \text{ mM D-Glucose}$, pH 7.4). Cells were incubated with starch NP suspension diluted with RPMI medium, PosSt or NegSt or water soluble potato starch, dissolved in RPMI medium (concentrations of $133, 67, 33, 17$, and $8 \mu\text{g mL}^{-1}$) for 4 h and 24 h, respectively. Afterwards, the supernatant was aspirated and the cells were washed once with $200 \mu\text{L}$ HBSS buffer. $100 \mu\text{L}$ fresh HBSS buffer, containing 10% (v/v) MTT reagent (5 mg mL^{-1}) was added and incubated for further 4 h. HBSS buffer was aspirated and $100 \mu\text{L}$ DMSO was added and



incubated for 20 min. Absorbance was measured at 550 nm with the Tecan Infinite[®] 200 microplate reader (Tecan Deutschland GmbH, Germany). All incubation steps were done at 37 °C, under gentle shaking and light protection. Cell viabilities were calculated in comparison to negative control (untreated cells as 100% value of cell viability) and positive control (1% TritonX-100 solution as 0% value of cell viability).

Uptake of starch nanoparticles into pulmonary cell lines

For the evaluation of starch NP uptake into A549 and 16HBE140⁻ cells, 1×10^5 and 0.6×10^5 cells per well, respectively, were seeded on a Transwell[®] membrane. After 24 h A549 cells were transferred to the air-liquid interface (ALI), *i.e.* medium was aspirated, and 500 μL of fresh medium was added to the basolateral compartment only. Two days later 250 μL of labeled starch NPs were deposited onto A549 cells with the AeronebLab nebulizer (Aerogen Ltd, Ireland). 16HBE140⁻ cells were transferred to ALI after 48 h in culture. 250 μL starch NPs were nebulized as for deposition on A549 or 16HBE140⁻ cells, respectively. In both cases, after 1 h, 4 h or 24 h of incubation at 37 °C, cells were washed with PBS and fixed with 3% paraformaldehyde (PFA) for 30 min at RT. After fixation, cells were permeabilized with a BSA/saponin solution, followed by an anti-phalloidin (actin) staining (1:143 in A549 and 1:100 in 16HBE140⁻ in PBS/BSA/saponin) for 30 min. After washing, the samples were counterstained with nuclei-staining DAPI (1:50 000 in PBS) and subsequently mounted with DAKO mounting medium (DAKO, USA). The samples were analyzed by a confocal laser scanning microscope (Zeiss LSM710, Zeiss, Germany). Lasers at 405 nm (DAPI), 488 nm (starch NPs) and 633 nm (actin) were used for detection. Microscopic images of fixed samples were acquired at 1024×1024 resolution, using a $63\times$ water immersion objective and z-stacks of 6 μm . Confocal images were analyzed using Zen 2012 software (Carl Zeiss Microscopy GmbH) and Fiji Software (a distribution of ImageJ, available at <http://fiji.sc>).

Loading of starch nanoparticles with proteins/peptides

In order to assess the use of starch NPs as DDS, a typical test set of four proteins varying in size, net charge, and hydrophilicity were chosen for loading experiments. Proteins that could actually act as active pharmaceutical ingredients (API) were chosen for use, rather than model proteins (such as ovalbumin (OVA) or bovine serum albumin (BSA)). In detail, these proteins were IgG1 (M_w : 150 kDa, IEP: 8.5), RNase A (M_w : 14 kDa, IEP: 9.6), insulin (M_w : 6 kDa, IEP: 5.3), and vancomycin (M_w : 1.5 kDa, IEP: 7.5). For the loading experiments, each protein sample was prepared as 5 mg mL⁻¹ stock solution in purified water, and added to the NP suspension to obtain final protein concentrations of 17.2 $\mu\text{g mL}^{-1}$, 34.2 $\mu\text{g mL}^{-1}$, and 51.2 $\mu\text{g mL}^{-1}$.

Loading efficiency

For the determination of protein encapsulation efficiency (EE) and loading rate, NPs and unincorporated protein were separated, after 15 min equilibration, by ultracentrifugation ($45\,000 \times g$ for 90 min; Optima L-XP, Beckman Coulter, UK). The amount of free

protein in the resulting supernatant was determined with a BCA (bicinchoninic acid) Assay Kit (Sigma Aldrich, USA). The principle of the assay relies on the formation of Cu²⁺-protein complexes that lead to the reduction of Cu²⁺ to Cu¹⁺ under alkaline conditions. BCA then forms a purple complex with Cu¹⁺ in alkaline conditions that can be detected by measurement of absorbance at 562 nm; absorbance values can then be correlated to protein concentrations. EE and loading rate of proteins in the NPs were defined and calculated according to the following equations:

$$\text{EE} [\%] = \frac{\text{mass}(\text{protein})_{\text{total}} - \text{mass}(\text{protein})_{\text{free}}}{\text{mass}(\text{protein})_{\text{total}}} \times 100$$

$$\text{Loading rate} [\%] = \frac{\text{mass}(\text{protein})_{\text{total}} - \text{mass}(\text{protein})_{\text{free}}}{\text{mass}_{\text{NP}}} \times 100$$

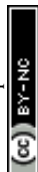
For the calculation of the particle yield (mass NPs), fixed volumes of NP suspension were filtered with Vivaflow 50 (100 kDa molecular weight cut off (MWCO), PES (Polyethersulfone) membrane, Sartorius, Germany), freeze dried and weighed.

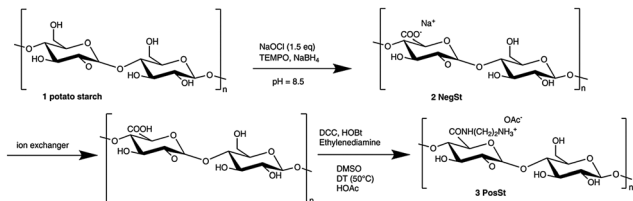
3 Results and discussion

Synthesis and characterization of starch derivatives

The raw material used for the synthesis of starch derivatives was a partially hydrolyzed potato starch with a molecular weight of 1 300 000 g mol⁻¹ and an amylose content of approximately 33%. Starch derivatives were prepared by TEMPO mediated oxidation²² of potato starch, resulting in a negatively charged starch derivative (NegSt). This well-known first oxidation step was also employed in order to further synthesize a positively charged starch derivative, *via* subsequent reaction of the carboxylic group of NegSt with the amine group of ethylenediamine (Scheme 1). The advantage of this initial oxidation process lies within the regioselectivity of the C-6 oxidation, which leads ultimately to a very controlled coupling product (PosSt). Positively charged starch (PosSt) was synthesized as mentioned by covalent coupling of ethylenediamine to the free acid form of oxidized starch, *via* a carbodiimide-mediated amidation. The method was adopted from Sheehan.²⁴ The benefit of this reaction is the selectivity of the coupling between carboxylic and amine groups, so that no protection of the hydroxyl groups is necessary. Additionally, there are no toxic side products/impurities, as is the case for synthesis of cationic starches in the paper industry, where the substitution is generally performed with epoxyalkylamines^{25,26} or halogenalkylamines.²⁷

After the oxidation of potato starch, ¹H-NMR spectra of NegSt (Fig. S1, ESI[†]) showed δH values between 5.28–5.58 ppm, that were assigned to the anomeric 1H atom. The signals between 3.20–4.20 ppm were ascribed to the ring protons 2H, 3H, 4H, 5H.^{21,28} Following the reaction, the degree of oxidation (DS_{COONa}) was determined by a colorimetric assay, according to Blumenkrantz *et al.*²³ Briefly, the Blumenkrantz assay detects uronic acid content (carboxylic groups arising from the TEMPO C-6 oxidation) by building a chromogen when heated in concentrated sulphuric acid/tetraborate and treated with *m*-hydroxydiphenyl. NegSt showed an oxidation degree of $40.32 \pm 0.01\%$. The yield of





Scheme 1 Synthesis of negatively charged starch (NegSt) and positively charged starch (PosSt). Potato starch is illustrated as amylose.

this process was 80.6%. The weight average molecular weight, determined by GPC was $87\,000 \pm 1800$ (Fig. S2, ESI[†]).

The coupling reaction could be observed by ¹H-NMR (Fig. S1, ESI[†]). PosSt showed the typical pattern between 5.10–5.62 ppm of the anomeric 1H atom. The ring protons were visible between 3.30–4.30 ppm. Further, the value between 3.00–3.22 ppm was ascribed to the coupled ethylenediamine (–NH–CH₂–CH₂–NH₂). FT-IR spectra confirm these results (Fig. 1): the typical peaks at 1600 cm^{-1} (C=O stretching of the carboxyl anion COONa) and 1732 cm^{-1} (C=O stretching from the COOH group, after ion exchange), were absent in PosSt. These were replaced by two peaks at 1660 cm^{-1} and at 1550 cm^{-1} , resulting from amide bond formation – the amide I peak (1660 cm^{-1}) is mainly associated with the C=O stretching vibration, while the amide II peak (1550 cm^{-1}) results from the N–H bending vibration and from the C–N stretching vibration. The reaction yield for the coupling with ethylenediamine was found to be 59.6% and the degree of coupling was 34.6%, determined by ¹H-NMR. GPC measurements showed a weight average molecular weight of $76\,000 \pm 4300$ for PosSt (Fig. S2, ESI[†]), which is in the same range as that found for NegSt.

Starch nanoparticle preparation

Starch NPs were prepared *via* charge-mediated coacervation in aqueous solution at RT, a process similar to the preparation of chitosan–TPP nanoparticles, introduced by Calvo *et al.*^{12,29} In contrast to the chitosan system, where a polymer (chitosan) is interacting with a small molecule (TPP), the starch NP formation involved the interaction of two polymeric materials of approximately the same molecular weight. Mixing the two oppositely charged starch derivatives under stirring led to the spontaneous formation of NPs, suggesting that partial charge neutralization occurred due to the reaction of positively

charged amine groups from PosSt with negatively charged carboxylic groups of NegSt. This process is very mild, as organic solvents, sonication, and high temperatures are avoided. Further, no additional excipients such as stabilizers are necessary for the NP preparation.

Different PosSt and NegSt concentrations (1 mg mL^{-1} , 0.5 mg mL^{-1} , 0.25 mg mL^{-1}) were employed for the preparation process. NegSt was added to PosSt in different molar ratios (1 : 3, 1 : 1, 3 : 1) with the help of a syringe pump under stirring. Important properties of the formed NPs, such as size, PDI and ζ -potential are described in Table 1, in relation to the used material concentrations and molar ratios. The PDI was always below 0.2, indicating a narrow particle size distribution for all formulations. Particle size ranged from approximately 140 to 350 nm, depending on both the molar ratio of components and concentration of solutions. The smallest particle size of $138.9 \pm 0.3\text{ nm}$ was obtained for 0.25 mg mL^{-1} concentrations and a molar ratio of 1 : 3. Keeping the molar ratio constant, but increasing the material concentrations to 0.5 and 1 mg mL^{-1} resulted in increased particle sizes of 160.5 ± 0.6 and $181.6 \pm 1.7\text{ nm}$, respectively. This is in accordance with Calvo *et al.*, who obtained similar results (an increase of particle size with increasing material concentrations) for the interaction of chitosan with TPP.¹² The utilized molar ratio was also seen to influence particle size. Looking again at 0.25 mg mL^{-1} concentrations, an increase in particle size from $138.9 \pm 0.3\text{ nm}$ to $152.0 \pm 0.2\text{ nm}$, and further to $209 \pm 2.8\text{ nm}$ was observed for molar ratios of 1 : 3, 1 : 1 and 3 : 1, respectively. This indicated that with an increase in the content of PosSt, the particle size also increased.

In the investigated ratios, the ζ -potential was between -35 and -10 mV , and was found to vary depending only on the molar ratio of PosSt and NegSt. For all three tested material concentrations, the ζ -potential was approximately -35 mV for a molar ratio of 1 : 3. A molar ratio of 1 : 1 resulted in a ζ -potential of around -25 mV , while starch NPs prepared in a molar ratio of 3 : 1 showed a ζ -potential of around -10 mV ; therefore, the greater the amount of positively charged starch (PosSt) in the formulation, the closer the ζ -potential was to 0. All formulations showed a negative ζ -potential, due to the fact that the degree of coupling with ethylenediamine was about 34.6%, meaning there are still free carboxylic groups left in PosSt (oxidation degree of NegSt $\sim 40\%$). NP formulations of 0.25 mg mL^{-1} were chosen for further experiments as a particle size of around 150 nm was desired.

For evaluating the storage stability of starch NPs, 0.25 mg mL^{-1} formulations with a molar ratio of 1 : 3, 1 : 1 and 3 : 1 were chosen, in order to investigate the influence of NP surface charge. Starch NPs showed different storage stabilities when stored after preparation: while starch 0.25–1 : 3 and starch 0.25–1 : 1 showed stability in size over 14 days, the formulation starch 0.25–3 : 1, with a ζ -potential close to zero showed aggregation tendencies when stored at RT or at $4\text{ }^{\circ}\text{C}$ (Fig. S4, ESI[†]). Obviously, for this formulation the weak surface charge is not able to electrostatically stabilize the NP suspension *via* repulsive forces.³⁰

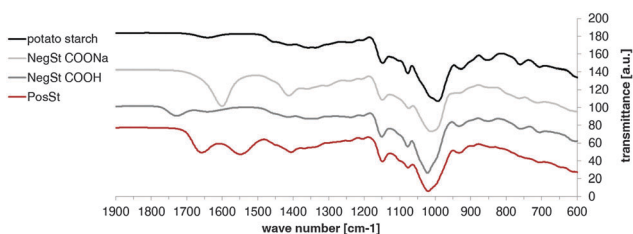


Fig. 1 FT-IR spectra of potato starch (black) and starch derivatives (dark red: PosSt; light grey: NegSt COONa; dark grey: NegSt COOH).



Table 1 Physicochemical characteristics of starch NPs related to concentration of solutions used for complexation and molar ratio of components. The last three formulations (shown in bold) were used for further evaluation of starch NPs, of which one (^a) was used for biodegradation, protein loading, and uptake studies

Formulation name	PosSt conc. [mg mL ⁻¹]	NegSt conc. [mg mL ⁻¹]	PosSt:NegSt [molar ratio]	Size [d nm]	PdI	ζ-Potential [mV]
Starch 1-1:3	1	1	1:3	181.6 ± 1.7	0.104	-31.1 ± 0.3
Starch 1-1:1	1	1	1:1	206.9 ± 0.7	0.078	-22.2 ± 0.1
Starch 1-3:1	1	1	3:1	346.9 ± 6.6	0.211	-10.3 ± 0.1
Starch 0.5-1:3	0.5	0.5	1:3	160.5 ± 0.6	0.088	-33.4 ± 1.6
Starch 0.5-1:1	0.5	0.5	1:1	179.0 ± 1.6	0.077	-23.5 ± 0.8
Starch 0.5-3:1	0.5	0.5	3:1	230.5 ± 4.2	0.084	-9.1 ± 0.1
Starch 0.25-1:3	0.25	0.25	1:3	138.9 ± 0.3	0.105	-37.0 ± 3.1
Starch 0.25-1:1^a	0.25	0.25	1:1	152.0 ± 0.2	0.077	-24.1 ± 0.6
Starch 0.25-3:1	0.25	0.25	3:1	209.3 ± 2.8	0.035	-8.5 ± 0.3

Morphology

TEM images of starch NPs, prepared from 0.25 mg mL⁻¹ solutions, incubated on a copper grid and stained with 1% PTA are shown in Fig. 2. For all formulations, black particles with a spherical, solid and consistent shape could be observed. Indicated sizes are comparable to gained Zetasizer data. Due to their gel-like, sticky character, NPs tended to aggregate during the drying process on the copper grid, forming a gel layer (data not shown). Therefore, starch NPs were positively stained and stabilized with 1% PTA during the drying process. While a considerable number of nanoparticles could be found for formulations starch 0.25-3:1 and starch 0.25-1:1, the formulation of starch 0.25-1:3 only showed few particles. The ζ-potential of -37.0 ± 3.1 mV probably hindered the ability of these particles to efficiently interact with the slightly negatively charged TEM grids during the incubation time.

Cell viability assay

For evaluating cell viability, A549 cells, representing a human pulmonary adenocarcinoma cell line,³¹ were incubated with starch NPs for 4 h and 24 h at 37 °C. In order to investigate the influence of molar ratio on cell viability, three different starch NP formulations were studied: Starch 0.25-3:1, starch 0.25-1:1, and starch 0.25-1:3. For comparison, NegSt, PosSt, and a water soluble potato starch (PSs) were tested. Concentrations were in a range from 8 to 133 μg mL⁻¹. Viability data of A549 cells after treatment with aforementioned samples for 4 h and 24 h can be found in Fig. 3.

At the 4 h measurement, PSs and NegSt showed cell viabilities of around 100%. PosSt showed a decrease in cell viability with increasing concentration, with a level of 65.4 ± 5.5% noted for the highest tested concentration. Similar results were

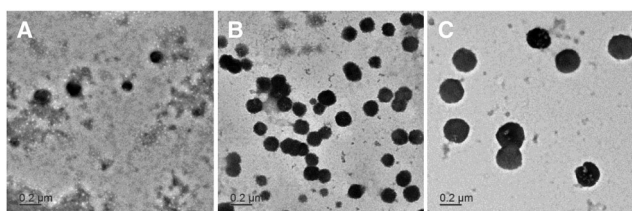


Fig. 2 TEM images of starch NPs, positively stained with 1% PTA. (A) Starch 0.25-1:3; (B) starch 0.25-1:1; (C) starch 0.25-3:1.

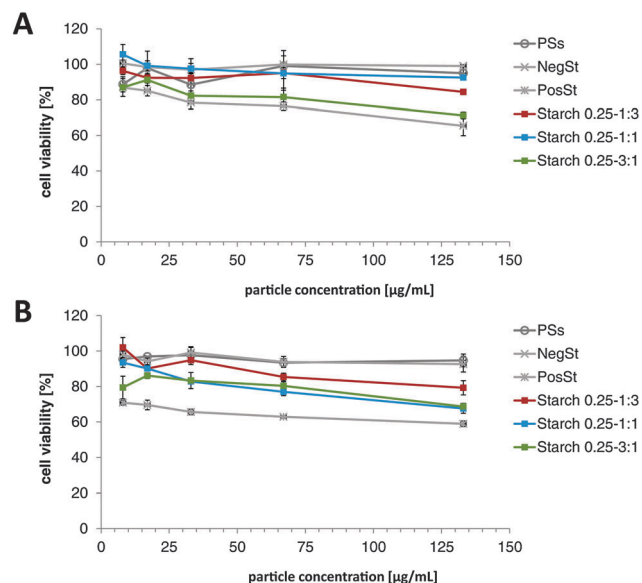


Fig. 3 Viability data of A549 cells after exposure to starch NPs, or starch derivatives (PosSt, NegSt), or a soluble potato starch (PSs) for 4 h (A) or 24 h (B). Data represent two independent experiments ($n = 3$ in each).

obtained for starch 0.25-3:1 with cell viabilities of around 71.3 ± 1.9%. Decreasing the PosSt amount in the NP formulation led to an increase in cell viability, which is not surprising bearing in mind that positively charged polymers usually show higher toxicities.^{20,32} The increase in toxicity after 24 h should be taken into consideration, but should not be overestimated, as degradation products usually show less toxicity compared to pure polymer – this could be shown by Yamada *et al.*,²⁰ with respect to the toxicity of starch-PEI derivatives and their degradation products. Further, the synthesized starch derivatives and prepared DDS showed higher cell viabilities as the starch-PEI derivatives from Yamada *et al.* for a comparable incubation time of 4 h, indicating an advantage of our newly developed DDS. Considering that the 1:1 ratio showed good cell viability and good particle stability over time, this formulation was chosen for further experiments.

Biodegradation

When talking about health care applications of NPs, one important point that has to be considered is the biodegradation



of the system. This is due to the fact that safety of NPs is not only related to their size, but also depends, amongst a number of other factors (toxicity of raw material, dose frequency, *etc.*), on their persistence in the body. The longer NPs stay in the body without any biodegradation, the higher the potential risk of adverse effects, *e.g.* due to unintentional stimulation of the immune system.³³ Further, degradation products of NPs must also be sufficiently safe, as otherwise they may contribute to unwanted effects such as non-specific immune responses. Therefore, we selected starch derivatives as material for DDS preparation. Starch itself consists of glucose monomers only, which are known to be safe. Only some monomer units of PosSt show coupled ethylenediamine (34.6%). This is further reduced by using a NP system, consisting of positively and negatively charged starch derivatives. In a proof of concept study, to show whether such NPs are degradable not only by unspecific degradation processes but also by a specific enzymatic degradation, we tested the biodegradability of starch nanoparticles by α -amylase – an enzyme found for example in saliva, pancreatic juice and serum.³⁴ It is even secreted in the lungs in a diseased state from carcinomas with adenomatous differentiation,³⁵ which could additionally confer a targeting effect on the starch NPs.

Particle concentration was determined with the help of nanoparticle tracking analysis after several time points. Compared to a control group (starch NPs stored at 37 °C without enzyme), which showed particle concentrations fluctuating between 40–55 $\times 10^8$ particles per mL over time, a drastic decrease in particle concentration, could be observed already after 2 h incubation with α -amylase at 37 °C (Table 2). While it might be expected that modified starch is less effectively degraded by α -amylase, and indeed Yamada *et al.* found that the biodegradation of modified starch-PEI derivatives by α -amylase was slower compared to the degradation of unmodified starch; however, the starch derivatives were still able to be degraded by the enzyme.²⁰ Furthermore, synthesized starch derivatives (NegSt, PosSt) in the current work represent a smaller change to the molecular structure of unmodified starch than starch-PEI derivatives, which have a relatively long molecular side chain. Accordingly, the incubation of starch NPs with α -amylase showed that also after charge-mediated coacervation, NegSt and PosSt could not only be degraded by unspecific degradation processes for polysaccharides or potentially by intracellular enzymes,^{36,37} but also by the specific human enzyme α -amylase.

Stability of nanoparticles upon aerosolization

The aerosolization properties of unlabeled and labeled starch NPs were investigated by nebulizing starch NP suspension, and

measuring NP physicochemical properties before and after nebulization. In Table 3 it can be seen that unlabeled starch NPs showed the same values before and after nebulization. The labeled starch NPs showed the same size and PdI, but exhibited a decrease in ζ -potential following nebulization. The fluorescent dye may change the hydrophobicity of the NPs, and thus the tightness of the surface binding of the ion layers which contribute to the zeta potential.

Uptake studies

In order to evaluate uptake of starch NPs into A549 and 16HBE140⁻ cells, PosSt was labeled with a green fluorescent dye (Bodipy[®] FL C5 NHS Ester), and starch NPs were prepared afterwards from 0.25 mg mL⁻¹ solutions in a ratio of 1 : 1. The size of these labeled starch NPs was 145.2 \pm 0.1 nm, with a PdI of 0.1, indicating a narrow size distribution. The ζ -potential was -26.4 ± 2.8 mV. Starch NPs were deposited onto A549 cells and further incubated for 1 h, 4 h and 24 h at 37 °C. As control, A549 cells were grown under ALI conditions without treatment. Endocytosis, as an energy-driven uptake mechanism, is highly reduced in cells at 4 °C; therefore, NP incubation was also evaluated at 4 °C for the A549 cell line (Fig. S6, ESI[†]). If NPs are taken up by the cells at 4 °C, it can be concluded that the uptake is not by an endocytosis-mediated mechanism. The uptake of starch NPs by A549 cells can be seen in Fig. 4A–D. Particles (green) already showed a level of uptake into A549 cells after 1 h of incubation at 37 °C. There was also some uptake after 1 h of incubation at 4 °C, although to a lower extent, suggesting that at least a part of the uptake mechanism is energy dependent (*i.e.* occurs by endocytosis). The cell associated fluorescence as observed at 4 °C could also be due to mere binding to the cell membrane, in addition to or instead of indicating some non-specific, energy independent uptake. Such nonspecific uptake is however known for A549 cells, suggesting that this could be a cell-dependent phenomenon.³⁸

Additionally, the interaction of starch NPs with 16HBE140⁻ cells, a cell line generated by transformation of normal human bronchial epithelial cells that does not show carcinoma like properties,^{39,40} was also tested. The results of this study can be found in Fig. 4E–H. The size of starch NPs in this case was 114.6 \pm 2 nm, with a PdI of 0.12 and a ζ -potential of -28.1 ± 0.6 mV. As can be seen from Fig. 4E–H, the nucleus staining was more intense, whereas the actin staining showed a decreased fluorescence in comparison to the A549 cells. No particle uptake was seen after 1 h of incubation. However, after

Table 2 Biodegradation of starch NPs by α -amylase at 37 °C

	Starch NPs + amylase [$\times 10^8$ particles per mL]	Starch NPs [$\times 10^8$ particles per mL]
Before	44.5 \pm 0.2	41.1 \pm 0.7
2 h	2.7 \pm 0.6	56.1 \pm 8.1
4 h	1.7 \pm 1.1	45.8 \pm 1.7
6 h	1.8 \pm 0.8	55.4 \pm 10.4
8 h	0.9 \pm 0.6	50.2 \pm 4.4

Table 3 Physicochemical properties of unlabeled and labeled starch NPs before and after nebulization, prepared from 0.25 mg mL⁻¹ solutions in a molar ratio of 1 : 1. Fluorescently labeled PosSt_F was 50% of the PosSt amount

Starch NPs	Size [d nm]	PdI	ζ -Potential [mV]
Before	168.7 \pm 2.1	0.09	-19.4 ± 4.9
After	167.4 \pm 7.2	0.09	-21.2 ± 2.2
Labeled before	145.2 \pm 0.7	0.1	-26.4 ± 2.8
Labeled after	145.2 \pm 2.2	0.09	-14.9 ± 6.6



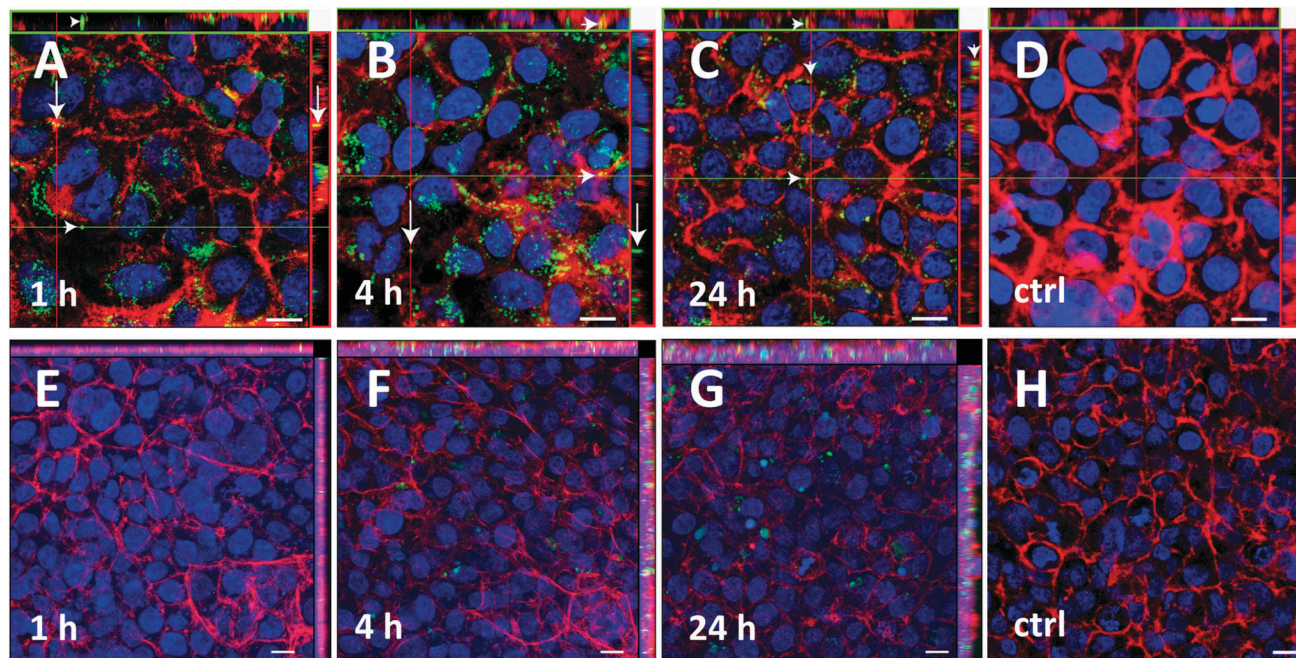


Fig. 4 Confocal laser scanning microscopy (CLSM) images of 250 μL nebulized starch NP onto A549 (upper row) and 16HBE140⁻ (lower row) cells, cultivated under ALI at 37 °C. (A) After 1 h incubation; (B) after 4 h incubation; (C) after 24 h incubation; (D) control; (E) after 1 h incubation; (F) after 4 h incubation; (G) after 24 h incubation; (H) control. Cells were washed with PBS, fixed with 3% PFA and stained. Blue: DAPI; red: actin; green: starch NP; scale bar: 10 μm ; arrow: extracellular NPs; arrow head: intracellular NPs.

an incubation time of 4 h or longer some particles could be observed and seem to be localized below the cell membrane, despite the non-optimal membrane staining. The apparent lower extent of particle uptake into 16HBE140⁻ cells compared to A549 cells should be judged with caution.

The confocal microscopy analysis shown here, provides qualitative data. The amount of uptake cannot be directly compared between the two cell types since the particle batch and the instrument settings (*e.g.* laser power) were not identical. An important factor in uptake studies is the proliferation rate of the cell line. A549 is a fast growing cancer cell line with a doubling time of 22 h. The non-cancerous 16HBE140⁻ cells are somewhat slower in growth with 26 h doubling time, a fact which may contribute to the apparently different uptake rate and speed. Despite being non quantitative, these experiments show a potential of uptake of starch NPs into two cell lines of pulmonary origin.

Loading

To investigate the use of starch NPs as DDS, a protein test set was created. This included four different proteins of different isoelectric points (IEPs) and molecular weights, which could also be potential drug candidates. Such proteins were to be tested *in vitro* for their efficiency in a next step. The selected proteins were IgG1 (M_w : 150 kDa, IEP: 8.5), RNase A (M_w : 14 kDa, IEP: 9.6), insulin (M_w : 6 kDa, IEP: 5.3), and vancomycin (M_w : 1.5 kDa, IEP: 7.5). For this experiment, starch NPs from 0.25 mg mL⁻¹ solutions and a molar ratio of 1 : 1 were chosen, as these showed the best stability and a medium level, negative ζ -potential of around -25 mV. Table 4 shows the physical

properties of protein-loaded starch NPs as well as EE and loading rate, as determined indirectly with the BCA assay after separating loaded NPs from free protein *via* ultracentrifugation. The EE and loading rate were different for all four proteins. The EE was affected by the initial amount of protein: the lower the amount, the higher the EE. Conversely, the loading rate was enhanced by increasing the initial amount of protein. Calvo *et al.* observed the same phenomenon for the encapsulation of BSA into chitosan NPs.¹² A narrow size distribution (low PDI) could be observed for all formulations, except for the formulation loaded with 500 μg IgG1. An increase in size, combined with a decrease in ζ -potential indicated that the loading with IgG1 was successful. It seems that the loading can be increased from approximately 4.5% (125 μg and 250 μg IgG1) up to 12.0% for the 500 μg formulation. However, when taking into account the broad size distribution of 0.39, such data must be interpreted cautiously. As the EE was obtained *via* indirect measurement, protein aggregation could have biased the results. A saturation of IgG1 loading could already be seen for the 125 μg formulation, as no difference could be found between the encapsulated mass and the loading rate of the 125 μg loaded NP and the 250 μg loaded NP. For these formulations the encapsulated mass was found to be 63 and 72 μg , respectively, and the loading was determined as 4.4 and 4.9% respectively.

Starch NPs loaded with RNase A showed no saturation, as the loading could be increased from 4.9% for the 250 μg loaded formulation up to 9.6% for the 750 μg formulation, however, the EE dropped from 27.9% down to 18.1%. The same could be observed for insulin: an EE of 24.9% for the loading with 250 μg of insulin decreasing to 15.9% for the loading with 750 μg



Table 4 Physicochemical properties, encapsulation efficiency (EE) and loading of starch nanoparticles loaded with different amounts of IgG1, RNase A, insulin or vancomycin

Starch NP (loaded with)	Mass _{initial} protein (μg)	Size [d nm]	PdI	ζ-Potential [mV]	Mass _{encaps} protein (μg)	EE [%]	Loading rate [%]
Blank	—	157.7 ± 2.2	0.08	-23.7 ± 0.7	—	—	—
IgG1 (<i>M_w</i> 150 kDa, IEP 8.5)							
	125	179.6 ± 6.1	0.10	-18.0 ± 0.2	63.2 ± 14.7	50.6 ± 11.7	4.4 ± 1.0
	250	237.4 ± 15.6	0.17	-13.1 ± 0.3	71.9 ± 1.9	28.8 ± 0.8	4.9 ± 0.1
	500	556.5 ± 49.5	0.39	-11.6 ± 0.9	174.3 ± 2.4	35.9 ± 0.5	12.0 ± 0.2
RNase A (<i>M_w</i> 14 kDa, IEP 9.6)							
	250	171.5 ± 0.3	0.07	-21.6 ± 0.6	72.0 ± 3.4	27.9 ± 1.3	4.9 ± 0.2
	500	172.9 ± 1.3	0.05	-17.7 ± 2.4	103.5 ± 3.1	20.1 ± 0.6	7.1 ± 0.2
	750	195.0 ± 31.5	0.13	-20.7 ± 0.5	139.8 ± 20.5	18.1 ± 2.7	9.6 ± 1.4
Insulin (<i>M_w</i> 6 kDa, IEP 5.3)							
	250	157.6 ± 0.7	0.09	-22.7 ± 1.2	62.4 ± 3.1	24.9 ± 1.2	4.3 ± 0.2
	500	162.3 ± 0.4	0.06	-21.0 ± 0.3	73.9 ± 4.7	14.8 ± 0.9	5.1 ± 0.3
	750	160.3 ± 0.9	0.07	-20.4 ± 0.5	119.2 ± 4.3	15.9 ± 0.6	8.2 ± 0.3
Vancomycin (<i>M_w</i> 1.5 kDa, IEP 7.5)							
	250	163.9 ± 0.4	0.08	-23.4 ± 0.7	205.2 ± 7.7	81.1 ± 3.0	14.2 ± 0.5
	500	164.7 ± 0.8	0.05	-21.2 ± 0.3	258.6 ± 0.2	51.1 ± 0.1	17.8 ± 0.1
	750	164.5 ± 0.6	0.06	-21.2 ± 0.4	330.1 ± 4.1	43.5 ± 0.5	22.8 ± 0.3

insulin, together with an increase in loading from 4.3% up to 8.2%. The most promising loading, however, was achieved for vancomycin, which showed loading efficiencies of up to 22.8% for the 750 μg loaded formulation, together with a high EE of 43.5%.

The differences in EE and loading rate can be ascribed to the differences in IEP and molecular weight. Calvo *et al.*, among others, described the phenomenon of protein loading as electrostatic interaction between protein and polysaccharide.¹² As the loading of the proteins into the starch NPs is a result of the electrostatic interactions between negatively charged NPs and protein, the IEP of each protein at the pH of the starch NP suspension (pH 7.4) has to be taken into account. Both IgG1 and RNase A are positively charged at pH 7.4, whereas insulin is negatively charged and vancomycin is neutral. A high EE and loading for IgG1 and RNase A would therefore be expected. However, IgG1 and RNase A loaded starch NP showed rather low loading capacities of 4.9% and 9.6%, respectively. The slightly higher loading of RNase could be due to the fact that the system loaded with IgG1 was already saturated in the case of the 250 μg formulation. IgG1 is an antibody with a molecular weight of around 150 kDa, and therefore requires more space when binding to the NP surface than *e.g.* RNase A, with a molecular weight of around 14 kDa. Insulin showed a similar behavior to RNase A: a loading of 8.2% could be achieved for the 750 μg formulation, although insulin is negatively charged at pH 7.4. Vancomycin, which is neutral at pH 7.4, showed the highest loading of 22.8%, probably due to a combined effect of low water solubility at this pH, as well as a low molecular weight of 1.5 kDa.

4 Conclusions

The derivatization of potato starch into water soluble, positively and negatively charged starches enabled the formation of starch NPs in aqueous medium *via* charge-mediated coacervation.

NPs with a spherical shape could be obtained. Physicochemical properties of NPs could be tuned by varying concentration and molar ratio of the two starch components. In a proof of concept study, the chosen NP formulation was seen to be biodegradable by α-amylase, and showed little cytotoxicity after 4 h. NPs were stable during nebulization and were observed to be taken up by A549 cells, largely by endocytosis. Further, the preparation of starch NPs allowed for loading of proteins/peptides of different molecular weights and IEPs under mild conditions. Vancomycin showed the highest loading rate (23%) and EE (44%), probably favored by the combination of the low molecular weight and the hydrophobicity of that particular compound. It can therefore be concluded that starch NPs prepared by such an eco-friendly and easy technique have the potential to be further explored as safe platform for pulmonary delivery of proteins and peptides. To further ensure protein stability during storage and administration and a high alveolar deposition, NPs could be embedded into microparticles.

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Notes and references

- 1 S. Lawrence, *Nat. Biotechnol.*, 2005, **23**, 1466.
- 2 G. Walsh, *Nat. Biotechnol.*, 2006, **24**, 769–776.



- 3 S. Mitragotri, P. A. Burke and R. Langer, *Nat. Rev. Drug Discovery*, 2014, **13**, 655–672.
- 4 S. Frokjaer and D. E. Otzen, *Nat. Rev. Drug Discovery*, 2005, **4**, 298–306.
- 5 J. L. Cleland, A. Daugherty and R. Mrsny, *Curr. Opin. Biotechnol.*, 2001, **12**, 212–219.
- 6 G. Orive, R. M. Hernández, A. R. Gascón, A. Domínguez-Gil and J. L. Pedraz, *Curr. Opin. Biotechnol.*, 2003, **14**, 659–664.
- 7 M. B. Brown, G. P. Martin, S. A. Jones and F. K. Akomeah, *Drug Delivery*, 2006, **13**, 175–187.
- 8 R. Duncan, *Nat. Rev. Cancer*, 2006, **6**, 688–701.
- 9 M. L. Houchin and E. M. Topp, *J. Pharm. Sci.*, 2008, **97**, 2395–2404.
- 10 O. Germershaus, T. Lühmann, J. C. Rybak, J. Ritzler and L. Meinel, *Int. Mater. Rev.*, 2015, **60**, 101–131.
- 11 M. Rajaonarivony, C. Vauthier, G. Couarraze, F. Puisieux and P. Couvreur, *J. Pharm. Sci.*, 1993, **82**, 912–917.
- 12 P. Calvo, C. Remunan-Lopez, J. L. Vila-Jato and M. J. Alonso, *J. Appl. Polym. Sci.*, 1997, **63**, 125–132.
- 13 A. Grenha, C. I. Grainger, L. A. Dailey, B. Seijo, G. P. Martin, C. Remuñán-López and B. Forbes, *Eur. J. Pharm. Sci.*, 2007, **31**, 73–84.
- 14 G. Coppi, V. Iannuccelli, E. Leo, M. T. Bernabei and R. Cameroni, *Drug Dev. Ind. Pharm.*, 2001, **27**, 393–400.
- 15 K. A. James, M. P. Fresneau, A. Marazuela, A. Fabra and M. J. Alonso, *J. Controlled Release*, 2001, **73**, 255–267.
- 16 M. Günbeyaz, A. Faraji, A. Özkul, N. Puralı and S. Şenel, *Eur. J. Pharm. Sci.*, 2010, **41**, 531–545.
- 17 A. Bozkir and O. M. Saka, *Drug Delivery*, 2004, **11**, 107–112.
- 18 S. Mansouri, Y. Cuie, F. Winnik, Q. Shi, P. Lavigne, M. Benderdour, E. Beaumont and J. C. Fernandes, *Biomaterials*, 2006, **27**, 2060–2065.
- 19 M. J. Santander-Ortega, T. Stauner, B. Loretz, J. L. Ortega-Vinuesa, D. Bastos-González, G. Wenz, U. F. Schaefer and C. M. Lehr, *J. Controlled Release*, 2010, **141**, 85–92.
- 20 H. Yamada, B. Loretz and C.-M. Lehr, *Biomacromolecules*, 2014, **15**, 1753–1761.
- 21 C. Thiele, D. Auerbach, G. Jung, L. Qiong, M. Schneider and G. Wenz, *Polym. Chem.*, 2011, **2**, 209–215.
- 22 P. L. Bragd, A. C. Besemer and H. Bekkum, *Carbohydr. Res.*, 2000, **328**, 355–363.
- 23 N. Blumenkrantz and G. Asboe-Hansen, *Anal. Biochem.*, 1973, **54**, 484–489.
- 24 J. C. Sheehan and G. P. Hess, *J. Am. Chem. Soc.*, 1955, **77**, 1067–1068.
- 25 T. Shiroza, K. Furihata, T. Endo, H. Seto and N. Otake, *Agric. Biol. Chem.*, 1982, **46**, 1425–1427.
- 26 E. F. Paschall, *US Pat.*, US2876217 A, 1959.
- 27 G. C. Carlyle and O. B. Wurzburg, *US Pat.*, US2813093 A, 1957.
- 28 Y. Kato, R. Matsuo and A. Isogai, *Carbohydr. Polym.*, 2003, **51**, 69–75.
- 29 P. Calvo, C. Remunan-Lopez, J. L. Vila-Jato and M. J. Alonso, *Pharm. Res.*, 1997, **14**, 1431–1436.
- 30 J. Jiang, G. Oberdörster and P. Biswas, *J. Nanopart. Res.*, 2008, **11**, 77–89.
- 31 D. J. Giard, S. A. Aaronson, G. J. Todaro, P. Arnstein, J. H. Kersey, H. Dosik and W. P. Parks, *J. Natl. Cancer Inst.*, 1973, **51**, 1417–1423.
- 32 M. Huang, E. Khor and L. Y. Lim, *Pharm. Res.*, 2004, **21**, 344–352.
- 33 C. M. Keck and R. H. Müller, *Eur. J. Pharm. Biopharm.*, 2013, **84**, 445–448.
- 34 J. J. Zakowski and D. E. Bruns, *Crit. Rev. Clin. Lab. Sci.*, 1985, **21**, 283–322.
- 35 P. Lenler-Petersen, A. Grove, A. Brock and R. Jernes, *Eur. Respir. J.*, 1994, **7**, 941–945.
- 36 S. Mahadevan, C. J. Dillard and A. L. Tappel, *Arch. Biochem. Biophys.*, 1969, **129**, 525–533.
- 37 B. Winchester, *Glycobiology*, 2005, **15**, 1R–15R.
- 38 H. Herd, N. Daum, A. T. Jones, H. Huwer, H. Ghandehari and C. M. Lehr, *ACS Nano*, 2012, **7**, 1961–1973.
- 39 B. Forbes, A. Shah, G. P. Martin and A. B. Lansley, *Int. J. Pharm.*, 2003, **257**, 161–167.
- 40 C. Ehrhardt, C. Kneuer, J. Fiegel, J. Hanes, U. F. Schaefer, K. J. Kim and C. M. Lehr, *Cell Tissue Res.*, 2002, **308**, 391–400.

