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Cross-species biosynthesis of maytansine in *Maytenus serrata*†‡

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Endophytic bacterial communities harboring different tissues of *Maytenus serrata* originating from Cameroon were investigated using targeted genome mining techniques coupled to bioanalytical approaches to elucidate the source of maytansine biosynthesis. It was revealed that the host plant, along with its cryptic endophytic microflora, produces the biosynthetically unique core structural moiety 3-amino-5-hydroxybenzoic acid (AHBA) that serves as the unique starter unit for maytansine biosynthesis. However, the biosynthetic step of halogenase-mediated incorporation of chlorine, which is missing in the host plant, is accomplished by the culturable stem endophytic bacterial community. Our results provide new insights into plant-endophyte communication for the biosynthesis of maytansine.

Endophytic microorganisms are ubiquitous in plants in the tissues and apoplastic compartments, both above- and below-ground, where they establish complex communities that can substantially contribute to the overall fitness of the host plants.^{1–5} Microorganisms rarely exist in “axenic forms” in nature and endophytes are no exception.⁶ In microbial communities within plants, the interplay between endophytes and plants, and the production of secondary metabolites results in a variety of functional traits of both agricultural and ecological importance.^{6–11} Thus far, it has been firmly established that suitable plant-endophyte and endophyte–endophyte interactions (fungi–fungi, fungi–bacteria, and/or bacteria–

bacteria) play a major role in the production of secondary metabolites.

In the last decades, studies on interspecies and multispecies crosstalk of endophytes have led to the elucidation of functional traits of endophytes concomitant to associated organisms.¹² Their coexistence within complex and mixed communities within their specific niches lead to their coevolution such that they can “withstand ecological forces” and function in ways that mostly cannot be accomplished by single organisms.¹³ For example, it was recently shown that a root-associated bacterial community of the tobacco plant, *Nicotiana attenuata*, functions as a consortium to provide protection to the host plant against sudden-wilt disease.¹⁴ Notably, this important ecological function could not be accomplished by single organisms of the root-associated bacterial community.¹⁴ Several recent studies have further lent evidence to the fact that certain so-called plant metabolites are actually biosynthesized by associated microbial community. For instance, we have shown that the original source organisms responsible for the biosynthesis of the important anticancer and cytotoxic compound maytansine (Fig. 1a) is the endophytic bacterial community harbored specifically within the roots of *Putterlickia verrucosa* and *Putterlickia retrospinoso* plants (Celastraceae).¹⁵ Chemical and molecular characterization of the root endophytic bacterial community revealed that maytansine is actually a biosynthetic product of *Putterlickia* root-associated endophytic bacterial community.

Given the fact that maytansine was discovered in the 1970s not only in *Putterlickia* species but also in other Celastraceae plants of the genus *Maytenus*,^{16,17} along with our aforementioned results on *Putterlickia* endophytes, we aimed to investigate the endophytes harbored in *Maytenus serrata* plants. The main difference, however, was that maytansine was found to be localized in the above-ground tissues of *Maytenus* plants compared to its accumulation in the roots of *Putterlickia* plants.^{16,17} Given these facts, in the present work, we answer the following open questions:

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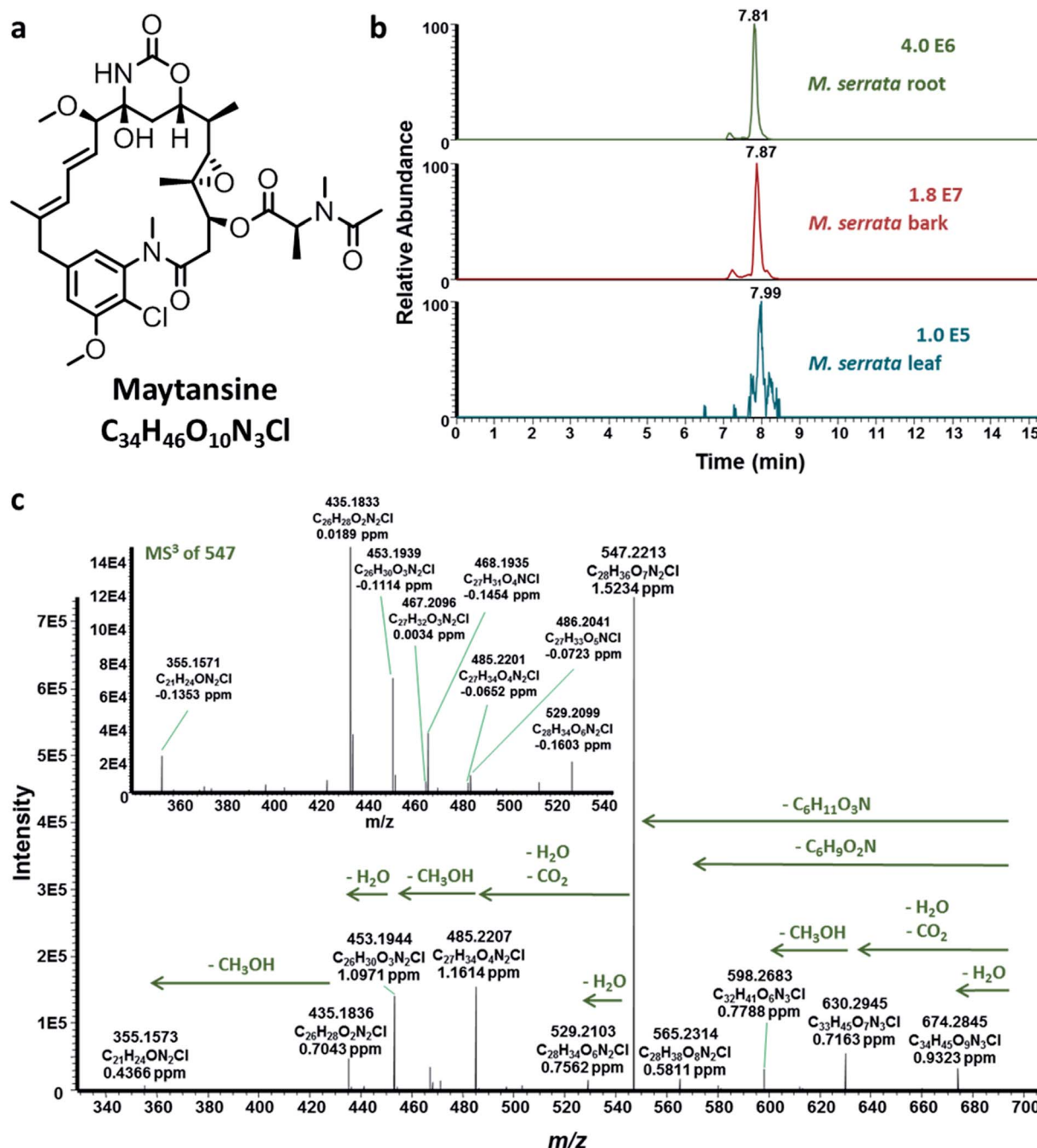


Fig. 1 (a) Maytansine. (b) Extracted ion chromatograms of maytansine (m/z 692.2944 \pm 2 ppm) of the root, bark and leaf extracts of *M. serrata* sampled from the Cameroonian rainforest. (c) Representative HRMS² and HRMS³ fragmentation of maytansine produced *in planta* by cross-species biosynthesis (host plant and endophytic bacterial community).

(A) Are endophytic communities responsible for maytansine biosynthesis in *Maytenus serrata* plants, similar to that of *Putterlickia* species?

(B) If so, where are these endophytes localized and what is their phylogenetic relationship to those harbored in *Putterlickia* species?

In order to shed light on the aforementioned questions, we collected *M. serrata* plants from two different locations encompassing very different ecological niches; the first location

was the rainforest at Bafoussam in Cameroon, whereas the second sampling was done at the Asakraka-Asikam district in Eastern Ghana. The above- and below-ground plant tissues were extracted and analyzed using HPLC-ESI-HRMSⁿ (see ESI†), which revealed that maytansine was present in the tissues of *M. serrata* sampled only from Cameroon (Fig. 1b and c and ESI, Fig. S1†), but not from Ghana. The striking results reinforced the notion that biosynthesis of maytansine in *M. serrata* plants is performed by the endophytes and not by the plants



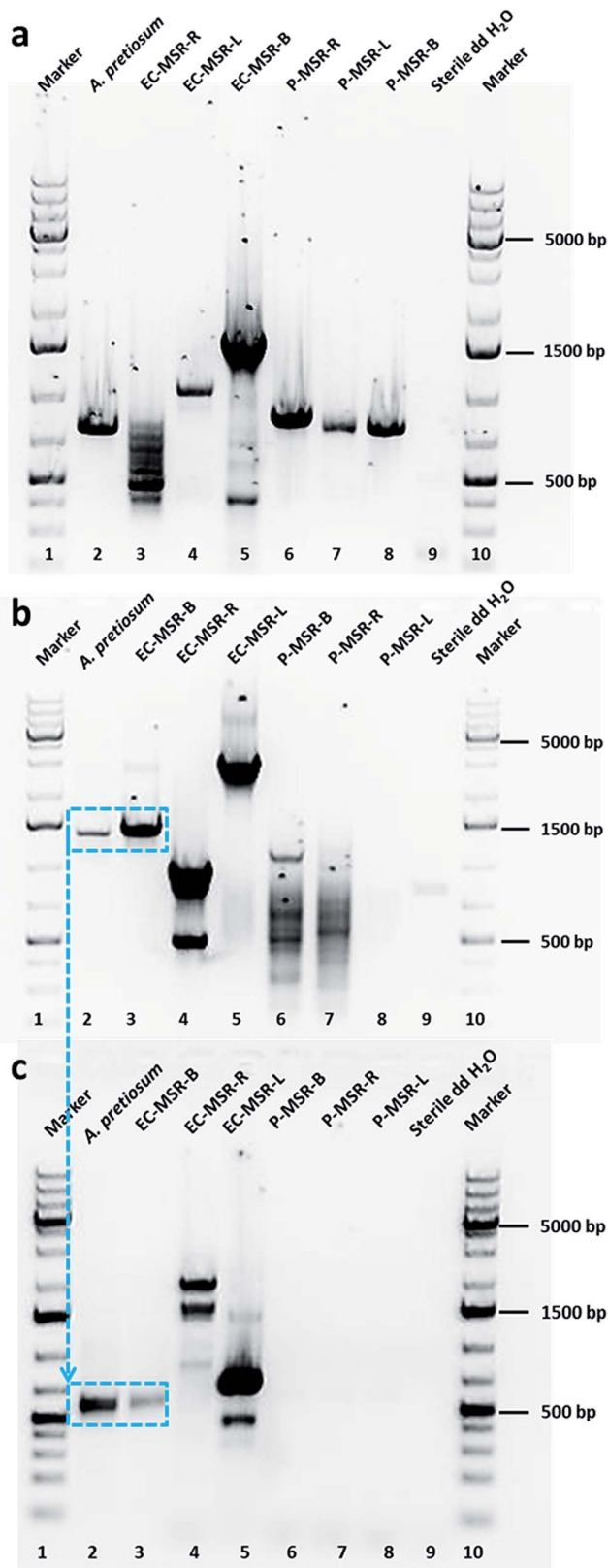


Fig. 2 (a) Stained agarose gels of purified PCR-amplified products of AHBA synthase genes (spanning around 755 bp) in root, leaf and bark tissues of *Maytenus serrata*. No band of AHBA synthase genes in endophytic bacterial communities of *Maytenus* plants. *Actinosynnema pretiosum* ssp. *auranticum* (DSM 44131) is the positive control and sterile double distilled water is the negative control. (b) Stained agarose

gels of purified PCR-amplified products of halogenase genes (spanning around 550 bp) in endophytic bacterial community of bark tissue of *Maytenus* plants. No band of halogenase genes in plant tissues and leaf and root endophytic communities of *Maytenus* plants. *A. pretiosum* (DSM 44131) is the positive control and sterile double distilled water is the negative control. (c) Stained agarose gels of purified PCR-amplified products of halogenase genes (spanning around 550 bp) in endophytic bacterial community of bark tissue of *Maytenus* plants. No band of halogenase genes in plant tissues and leaf and root endophytic communities of *Maytenus* plants. PCR purified product of *asm12* gene of *A. pretiosum* (DSM 44131) is the positive control and sterile double distilled water is the negative control.

themselves,⁵ similar to what was demonstrated earlier for *Putterlickia* plants.¹⁵ Taking cues from our preliminary results, we examined the biosynthesis of maytansine in the Cameroonian *M. serrata* and the possible contribution of endophytic bacterial community harbored in different tissues (root, bark and leaf) of the plant. Firstly, we evaluated the different tissues of *Maytenus* plant and the endophytic bacterial community for the presence of genes biosynthesizing the important starter enzyme, AHBA synthase (3-amino-5-hydroxybenzoic acid synthase).¹⁵ This enzyme comprises of a unique mC7N structural moiety which forms the common precursor of different ansamycins.¹⁸ Thus, a pair of degenerate primers (primer pair P3; ESI, Table S1†) were designed amplifying the conserved amino acid regions of different genes namely *asnF*, *asm24*, *mitA*, *napF* and *rifK*.¹⁵ As a positive control, the genomic DNA of *Actinosynnema pretiosum* subsp. *auranticum* (DSM 44131) was amplified resulting in an approx. 755 bp fragment. Further, the root, leaf and bark tissues and the respective endophytic bacterial communities were subjected to the optimized PCR conditions using the same degenerate primer pair. All the plant tissues amplified the expected fragment of AHBA synthase gene (Fig. 2a). Interestingly, none of the endophytic bacterial communities produced the desired amplification product. The positive PCR amplified products were sequenced, aligned, identified, and further translated into protein sequences. The positive control showed 100% homology to 3-amino-5-hydroxybenzoic acid synthase (AHBA synthase; EMBL AAC13997.1 and UniProt Identifier Q44131). Similarly, the amplified products from the different plant tissues also showed sequence homology to 3-amino-5-hydroxybenzoic acid synthase (ESI, Table S2†). The sequences of the amplified PCR products have been submitted to the EMBL-Bank. We also evaluated the aerial and non-aerial tissues of the Ghanaian *M. serrata*, which does not contain maytansine, for the presence of AHBA synthase genes. Interestingly, AHBA synthase genes were found to be absent in this plant (data not shown). Thus far, we could conclude that only the *M. serrata* plant growing in the rainforest of Cameroon contains the genetic blueprint of the starter enzyme (AHBA synthase) for the biosynthesis of maytansine.

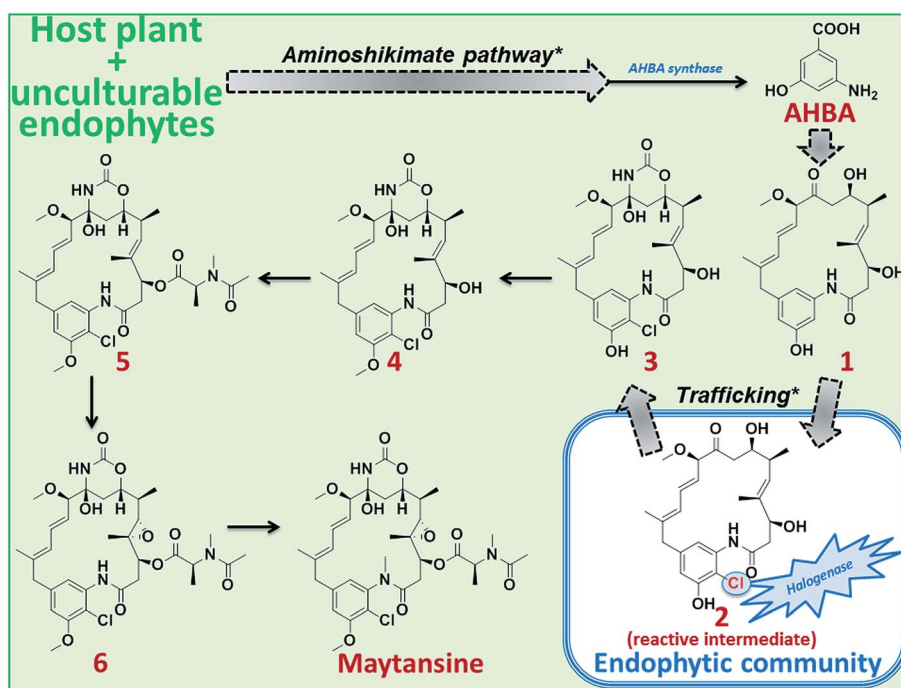
Taking ansamitocin biosynthetic pathway into consideration, we decided to further investigate the post-polyketide synthase modification of the ansamitocin pathway. One of the key steps in this pathway is the conversion of proansamitocin to

gels of purified PCR-amplified products of halogenase genes (*asm12*; spanning around 1300 bp) in endophytic bacterial community of bark tissue of *Maytenus* plants. No band of halogenase genes in plant tissues and leaf and root endophytic communities of *Maytenus* plants. *A. pretiosum* (DSM 44131) is the positive control and sterile double distilled water is the negative control. (c) Stained agarose gels of purified PCR-amplified products of halogenase genes (spanning around 550 bp) in endophytic bacterial community of bark tissue of *Maytenus* plants (1300 bp fragment used a template). No band of halogenase genes in plant tissues and leaf and root endophytic communities of *Maytenus* plants. PCR purified product of *asm12* gene of *A. pretiosum* (DSM 44131) is the positive control and sterile double distilled water is the negative control.



19-chloroproansamitocin that introduces chlorine into the structure. Chlorination is an important FADH₂-dependent halogenation catalyzed by different halogenase enzymes. In the ansamitocin biosynthetic pathway of positive control *A. pretiosum*, this step is achieved by the gene *asm12*, which is a FADH₂-dependent halogenase.¹⁹ We, therefore, aimed to evaluate the plant tissues and the endophytic communities for the presence of halogenase genes in order to elucidate which organism (host plant or endophytic bacterial community) is responsible for incorporation of chlorine towards completing the biosynthesis of maytansine. We employed two strategies to answer this question. In our first approach, we evaluated for the presence of halogenase gene *asm12*, which catalyzes the first step post-polyketide synthase modification responsible for the chlorination step in ansamitocin biosynthesis. For the detection of *asm12* gene, the genomic DNA of plant tissues and endophytic community were subjected to PCR amplification using primer pairs P1 (ESI, Table S1†). The P1 primers were designed based on the sequence of *asm12* gene from *A. pretiosum* (DSM 44131). None of the plant tissues (leaf, stem, or root) amplified the desired product (Fig. 2b). Furthermore, the leaf and root endophytic bacterial community also did not amplify the desired product (Fig. 2b). Interestingly, only the bark endophytic bacterial community (EC-MSR-B) could produce the desired fragment (Fig. 2b). As a positive control, the genomic DNA of *A. pretiosum* was amplified (spanning around 1300 bp). The positive fragments were sequenced, aligned, identified, and further translated into protein sequences. The positive control showed 99% homology to *asm12* halogenase gene (EMBL AAM54090.1) and 100%

homology to UniProt Identifier Q8KUG0. Although the bark endophytic community amplified the desired fragment, the homology with phylogenetically-related reference halogenase sequence (*i.e.*, *asm12* of *A. pretiosum*) was relatively low. In order to avoid any false positives, in our second approach we further analyzed the presence of any FADH₂-dependent halogenase irrespective of *asm12* gene in particular. Therefore, we performed a second PCR analysis with degenerate primers deduced from highly conserved regions of six different FADH₂-dependent halogenases (ansamitocin being only one of them) having different enzyme specificities to produce different halogenated metabolites.²⁰ We re-evaluated the plant tissue and endophytic bacterial communities for the presence of halogenase genes by subjecting them to PCR amplification with primer pair P2 (ESI, Table S1†). For the positive control, we used the PCR purified 1300 bp *asm12* gene of *A. pretiosum* as template for primer walking with P2. Simultaneously, the 1300 bp positive fragment of bark endophytic bacterial community was also subjected to second PCR using primer pair P2. As expected, we observed an approximately 550 bp amplified fragment for positive control and only bark endophytic bacterial community (Fig. 2c). None of the plant tissues or other endophytic bacterial communities amplified this fragment. The positive fragments were evaluated as detailed above. The positive control naturally showed 99% homology EMBL AF453501.1 and 97.2% homology to UniProt Identifier Q8KUG0. Interestingly, this time the bark endophytic bacterial community showed 100% homology to halogenase (EMBL CP001630.1) and 100% homology to UniProt Identifier C6W874 (tryptophan halogenase).



Scheme 1 Proposed plant-endophyte cross-species biosynthetic pathway of maytansine in *M. serrata*. Plausible steps of the aminoshikimate pathway and of trafficking of maytansine precursors between host plant and its endophytic bacterial community leading to the cross-species biosynthesis of maytansine are marked with asterisks. Dotted arrows represent proposed hypothetical steps.



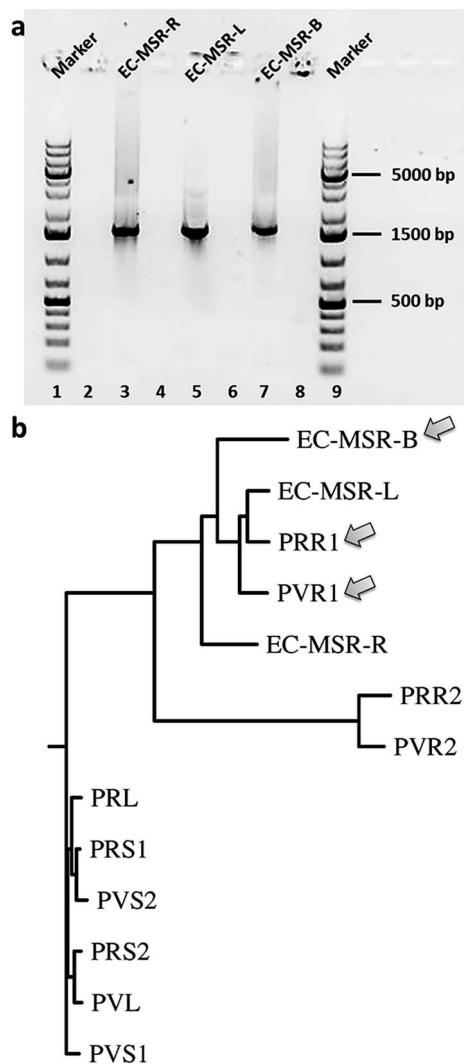


Fig. 3 (a) Stained agarose gels of purified PCR products of 16S rRNA analyses of endophytic bacterial communities of *Maytenus serrata* (spanning around 1500 bp). (b) The phenogram-like rooted phylogenetic tree diagram of endophytic bacterial communities of the present study (EC-MSR-B, EC-MSR-L, EC-MSR-R), compared to ones reported earlier from *Putterlickia* plants,¹⁵ using Drawgram in PHYLIP 3.66 based on their 16S rRNA regions.

As expected, none of the isolated endophytes and endophytic bacterial communities could produce maytansine when fermented under laboratory conditions (outside the host plant). Furthermore, HRMS/MS allowed us to complement the genetic data; we could detect the major precursors (1–6) of maytansine in the plant tissues, particularly in the bark (ESI, Fig. S2 and Table S3†). Thus far, using the combination of molecular and analytical data, we propose a plant-endophyte cross-species biosynthetic pathway for production of maytansine that occurs only in some *M. serrata* plants (such as the one sampled from Cameroonian rainforest) (Scheme 1). The host plant along with its unculturable or cryptic endophytes produces AHBA, the unique starter moiety of maytansine biosynthesis, by means of an aminoshikimate pathway^{21,22} that is absent in the cultivable bark endophytic bacterial community. AHBA is further converted to precursor 1,

which is taken up by the bark endophytic bacterial community to introduce chlorine (precursor 3 via reactive intermediate 2) using the halogenase enzyme that is absent in the host plant and its cryptic endophytes (ESI, Fig. S2 and Table S3†). Further biosynthetic steps through intermediates 4–6 to the production of maytansine is achieved by the host plant (Scheme 1).

Finally, we aimed to elucidate the phylogenetic relationship between the different endophytic bacterial communities isolated from the leaves, stems, and roots of the maytansine-containing *M. serrata* plant and those harbored in *Putterlickia* species.¹⁵ The 16S rRNA amplification of the endophytic bacterial communities (Fig. 3a) followed by phylogenetic evaluation using T-Coffee (ESI, Fig. S3†) was done according to Kusari *et al.* (2014).¹⁵ The phenogram (Fig. 3b) revealed a clear clustering of leaf, bark and root endophytic bacterial communities of *M. serrata* to that of primary root endophytic bacterial communities of *Putterlickia* plants. Interestingly, the bark endophytic bacterial community containing the halogenase gene (EC-MSR-B) clustered very close to the root endophytic bacterial communities of *Putterlickia* plants that were found to produce maytansine (PRR1 and PVR1).¹⁵ This shows plausible species-level relatedness between endophytic bacterial communities that contain the genes involved in the biosynthetic pathway of maytansine. Interestingly, the endophytic bacterial communities of *M. serrata* under our study also showed evolutionary relationships to bacteria harboring diverse hosts (niches) ranging from termites to earthworms (ESI, Fig. S4†).

Taken together, our study reveals a unique aspect of plant-endophyte communication where only the bark endophytic bacterial community of a selected *M. serrata* plant had coevolved a means to produce maytansine jointly with the host plant, including trafficking of the precursors from and to the host plant. In other *M. serrata* plants where this endophytic community is absent, there is no production of maytansine, as exemplified not only by the Ghanaian *M. serrata* plant but also other *Maytenus* plants of different origin presently under our investigation (data not shown). Our results provide a scientific handle to delve into further details of plant-endophyte communication not only on the transcriptome level but also the metabolome level to pinpoint regulation of host-endophyte communication. Our study further opens avenues to determine qualitative and quantitative pattern of localization and trafficking of endophytes with regard to shared biosynthesis of maytansine. Further studies along these lines are currently underway.

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Notes and references

- 1 T. Hosni, C. Moretti, G. Devescovi, Z. R. Suarez-Moreno, M. B. Fatmi, C. Guarnaccia, *et al.*, *ISME J.*, 2011, **5**, 1857.
- 2 D. G. Meldau, S. Meldau, L. H. Hoang, S. Underberg, H. Wünsche and I. T. Baldwin, *Plant Cell*, 2013, **25**, 2731.
- 3 S. Kusari, S. Singh and C. Jayabaskaran, *Trends Biotechnol.*, 2014, **32**, 304.
- 4 P. Kusari, S. Kusari, M. Lamshöft, S. Sezgin, M. Spiteller and O. Kayser, *Appl. Microbiol. Biotechnol.*, 2014, **98**, 7173.
- 5 D. J. Newman and G. M. Cragg, *Front. Chem.*, 2015, **3**, 34.
- 6 S. Kusari, C. Hertweck and M. Spiteller, *Chem. Biol.*, 2012, **19**, 792.
- 7 V. Schroeckh, K. Scherlach, H. W. Nützmann, E. Shelest, W. Schmidt-Heck, J. Schuemann, *et al.*, *Proc. Natl. Acad. Sci. U. S. A.*, 2009, **106**, 14558.
- 8 B. Reinhold-Hurek and T. Hurek, *Curr. Opin. Plant Biol.*, 2011, **14**, 435.
- 9 A. Debbab, A. H. Aly and P. Proksch, *Fungal Divers.*, 2013, **61**, 1.
- 10 A. L. Navarro-Meléndez and M. Heil, *J. Chem. Ecol.*, 2014, **40**, 816.
- 11 L. Tyvaert, S. C. França, J. Debode and M. Höfte, *J. Appl. Microbiol.*, 2014, **116**, 1563.
- 12 S. Kusari, S. P. Pandey and M. Spiteller, *Phytochemistry*, 2013, **91**, 81.
- 13 J. de Vrieze, *Science*, 2015, **349**, 680.
- 14 R. Santhanam, V. T. Luu, A. Weinhold, J. Goldberg, Y. Oh and I. T. Baldwin, *Proc. Natl. Acad. Sci. U. S. A.*, 2015, **112**, e5013.
- 15 S. Kusari, M. Lamshöft, P. Kusari, S. Gottfried, S. Zühlke, K. Louven, *et al.*, *J. Nat. Prod.*, 2014, **77**, 2577.
- 16 S. M. Kupchan, Y. Komoda, W. A. Court, G. J. Thomas, R. M. Smith, A. Karim, *et al.*, *J. Am. Chem. Soc.*, 1972, **94**, 1354.
- 17 S. M. Kupchan, Y. Komoda, A. R. Branfman, A. T. Sneden, W. A. Court, G. J. Thomas, *et al.*, *J. Org. Chem.*, 1977, **42**, 2349.
- 18 Q. Kang, Y. Shen and L. Bai, *Nat. Prod. Rep.*, 2012, **29**, 243.
- 19 Y. Wu, Q. Kang, Y. Shen, W. Su and L. Bai, *Mol. Biosyst.*, 2011, **7**, 2459.
- 20 A. Hornung, M. Bertazzo, A. Dziarnowski, K. Schneider, K. Welzel, S. E. Wohlert, *et al.*, *ChemBioChem*, 2007, **8**, 757.
- 21 H. G. Floss, *Nat. Prod. Rep.*, 1997, **14**, 433.
- 22 H. Maeda and N. Dudareva, *Annu. Rev. Plant Biol.*, 2012, **63**, 73.

