



Cite this: *Dalton Trans.*, 2015, **44**, 19207

Interplay between singlet and triplet excited states in a conformationally locked donor–acceptor dyad†

Mikhail A. Filatov,^{*‡a,b} Fabian Etzold,^a Dominik Gehrig,^a Frédéric Laquai,^{a,c} Dmitri Busko,^a Katharina Landfester^a and Stanislav Balushev^{a,d,e}

The synthesis and photophysical characterization of a palladium(II) porphyrin – anthracene dyad bridged via short and conformationally rigid bicyclo[2.2.2]octadiene spacer were achieved. A spectroscopic investigation of the prepared molecule in solution has been undertaken to study electronic energy transfer in excited singlet and triplet states between the anthracene and porphyrin units. By using steady-state and time-resolved photoluminescence spectroscopy it was shown that excitation of the singlet excited state of the anthracene leads to energy transfer to the lower-lying singlet state of porphyrin. Alternatively, excitation of the porphyrin followed by intersystem crossing to the triplet state leads to very fast energy transfer to the triplet state of anthracene. The rate of this energy transfer has been determined by transient absorption spectroscopy. Comparative studies of the dynamics of triplet excited states of the dyad and reference palladium octaethylporphyrin (PdOEP) have been performed.

Received 27th September 2015,
Accepted 13th October 2015

DOI: 10.1039/c5dt03784k

www.rsc.org/dalton

Introduction

Chromophores producing long-lived triplet excited states under excitation with light have attracted much attention during the past decade due to great promise for application in diverse areas of research, particularly aimed at sunlight harvesting.¹ Chemical modification is a key tool for the control of excited-state properties, and consequently emission and energy transfer parameters (efficiency and rates) of the chromophore.² Particularly, introduction of intramolecular excited state donor–acceptor couplings prolongs the triplet lifetime by enabling a reversible intramolecular exciton transfer,

the so called “energy reservoir” effect.³ Such modifications allow for improving the efficiencies of corresponding photonic applications.⁴ Phosphorescent donor-bridge-acceptor (D-b-A) dyes are important materials for a number of optoelectronic,⁵ sensing,⁶ bioimaging⁷ and applications based on triplet-triplet annihilation photon upconversion (TTA-UC).⁸

In the TTA-UC process, D-b-A triplet sensitizers promise to provide a new generation of molecular photonic devices. Efficient implementation of the process requires two components in a mixture: a sensitizer (transition metal complex) and an emitter (aromatic hydrocarbon or BODIPY), which generate higher energy photons through a sequence of energy transfer processes.⁹ The development of a single molecular system combining the properties of sensitizer and emitter and capable of anti-Stokes emission is of fundamental importance and is desired for sensing applications. Attempts to incorporate 9,10-diphenylanthracene (DPA) into ruthenium(III) complex structures provided a delayed fluorescence (TTA-based upconversion emission) of covalently bound anthracene, however with low efficiency compared to the mixture.¹⁰ The key factors governing energy transfer properties in such dyads are to be elucidated in order to develop efficient single molecular upconversion systems.

A number of studies carried out with covalently/noncovalently linked porphyrin–porphyrin¹¹ and porphyrin–nonporphyrinic chromophore¹² assemblies have dealt with the effect of D-b-A distance on the rates of the triplet–triplet energy transfer (TTET) process, however, relatively little attention has

^aMax Planck Institute for Polymer Research, Ackermannweg 10, D-55128 Mainz, Germany. E-mail: filatovm@tcd.ie

^bInstitute of Polymers, Bulgarian Academy of Sciences, Acad. G. Bonchev Str., block 103-A, BG – 1113 Sofia, Bulgaria

^cPhysical Sciences and Engineering Division (PSE), Material Science and Engineering (MSE), Solar and Photovoltaics Engineering Research Center (SPERC), King Abdullah University of Science and Technology (KAUST), Thuwal 23955-6900, Kingdom of Saudi Arabia

^dOptics and Spectroscopy Department, Faculty of Physics, Sofia University “St. Kliment Ochridski”, 5 James Bourchier, 1164 Sofia, Bulgaria

^eFreiburg Institute for Advanced Studies (FRIAS), Albert-Ludwigs-Universität Freiburg, Albertstraße 19, D-79104 Freiburg, Germany

†Electronic supplementary information (ESI) available: Optical spectra, NMR and mass-spectra of newly synthesized compounds. See DOI: 10.1039/c5dt03784k

‡Current address: Trinity Biomedical Science Institute, Trinity College Dublin, 152-160 Pearse Street, Dublin 2, Ireland.



been paid to the effects of mutual orientation between donor and acceptor.¹³ Giribabu and co-workers demonstrated striking orientation dependence of the energy transfer in a porphyrin-based D-b-A system, in which the donor subunits are linked both at the axial and peripheral sites of the porphyrin macrocycle (acceptor). Photoluminescence studies revealed that energy transfer (ET) from the porphyrin donor to the acceptor subunits bound at the peripheral positions is efficient, while no ET takes place to the acceptors bound at the axial positions.¹⁴

Another fundamental question arising from these studies is whether the electronic excitation leading to a long-lived triplet state is accompanied by a change in the molecular conformation, especially for porphyrins, known to possess various macrocycle distortion modes.¹⁵ Little is known about photophysical behaviour of conformationally rigid D-b-A porphyrin systems, specifically regarding the triplet state dynamics and phosphorescence energies. Although porphyrin D-b-A systems have been extensively studied,¹⁶ no examples of such system, which can be regarded as “rigid” meaning that no conformational changes in the excited state can take place, are known. In this work we studied a D-b-A system, capable of TTET, in which conformational changes are excluded due to the specific arrangements of donor and acceptor moieties along a bridging bicyclic subunit.

A molecular design concept of the target system is illustrated in Fig. 1. The assembly of porphyrin D-b-A dyads has commonly been based on the functionalization of *meso*-positions of the macrocycle, since corresponding synthetic methods are well developed.¹⁷ However, *meso*-substitution is known to cause non-planar distortion of the porphyrin macrocycle, adopting saddle, waved, ruffled or domed conformation.¹⁸ Systematic analysis of photophysical properties of *meso*-substituted porphyrins reveals that: (1) chromophores attached through *meso*-positions are in partial conjugation to the porphyrin's aromatic system and (2) rotational motions

along C_{meso}-C_{aryl} bonds are possible.¹⁹ It results in lower quantum yields of emission and slows down excited state energy transfer processes. On the other hand, it was shown that β -substituted porphyrins with four bicyclo[2.2.2]octene-fused frameworks adopt a slightly waved, planar conformation.²⁰ In all reported crystal structures of these porphyrins, the porphyrin chromophore was almost flat and showed only slight out-of-plane distortion.²¹ Separation of aromatic systems by a rigid bridges based on polycyclic hydrocarbon scaffolds has been shown to provide well-defined distances and orientations of the photoactive fragments.²² Based on these findings, we designed a porphyrin D-d-A dyad, in which the macrocycle is surrounded by four anthracene moieties connected *via* bicyclo[2.2.2]octene spacer (Fig. 1B) and studied its optical properties and excited state dynamics.

Results and discussion

Synthesis

The synthesis of the target molecule is shown in Scheme 1. Starting quinizarine was converted to 1,4-anthraquinone **1** by the reduction with NaBH₄. Compound **1** was further subjected to Diels–Alder reaction with 1,3-cyclohexadiene in boiling chloroform which delivered product **2**. It should be noted, that in the case of toluene as a solvent, heating of the reaction mixture above 100 °C resulted in the formation of tetracene-5,12-dione as a by-product *via* thermal retro-Diels–Alder reaction.

Acenes are known to possess low solubility, requiring the introduction of alkyl, alkoxy or alkoxy carbonyl substituents. Introduction of such substituents affects excited states energies.²³ The latter is of particular importance with respect to photophysical properties of a designed system, since an overlap of donor and acceptor triplet manifolds is to be achieved. Thus, we treated compound **2** with a base (DBU), to form deprotonated hydroquinone, which was then reacted with MeI to provide 1,4-dimethoxyanthracene derivative **3** in good overall yield. Anthracene **3** was subjected to a reaction with PhSCl that resulted in the corresponding chlorosulfide, which was further oxidized to give chlorosulfone **4**. Subsequent HCl elimination under DBU treatment yielded the corresponding allylic sulfone **5**, which was used for Barton–Zard reaction with isocyanacetate leading to pyrrole **6**. Its reduction by LiAlH₄ delivered pyrrole derivative **7** which was introduced into acid-catalyzed tetramerization²⁴ without isolation to give corresponding porphyrin **8** in a good yield (25%).

Finally, the free-base porphyrin **8** was subjected to palladium(II) insertion reaction leading to the target complex **9**. Careful control over the reaction temperature was critically important to prevent by-products formation. Palladium insertion into porphyrin macrocycles usually requires prolonged reflux in high boiling point solvents such as benzonitrile.²⁵ In the case of porphyrin **9**, heating over 150 °C was found to cause a retro-Diels–Alder reaction,²⁶ leading to the corresponding tetracene-fused derivatives, as was evidenced by mass-spectroscopy.

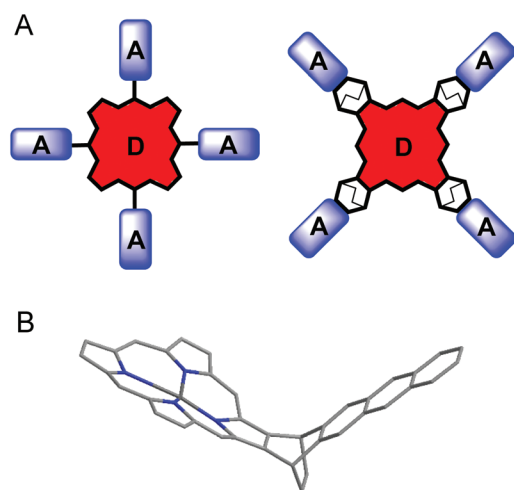
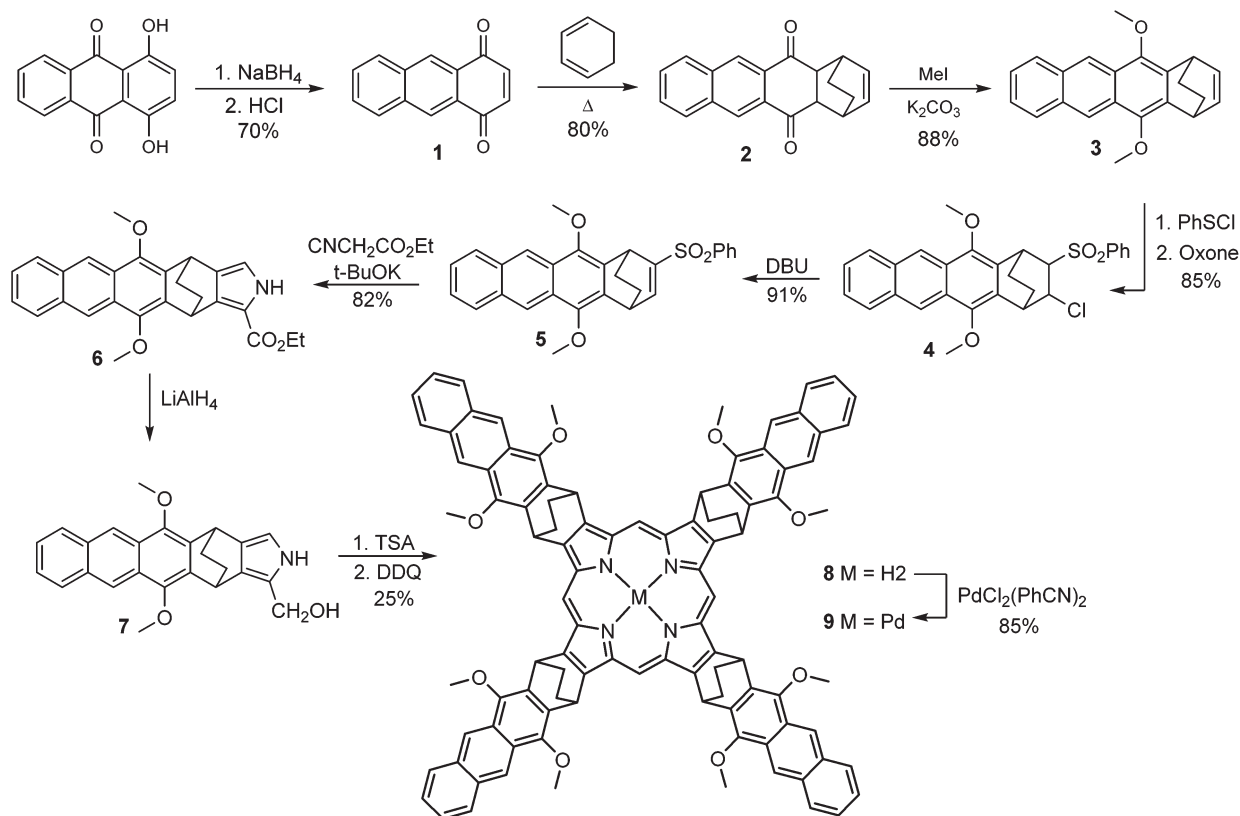


Fig. 1 (A) Possible structures of porphyrin D-b-A system, (B) porphyrin-anthracene dyad with bicyclo[2.2.2]octane spacer.





Scheme 1 Synthesis of target molecule.

In the course of isolation and purification of crude products **8** and **9** we noticed a formation of by-products upon their illumination with light under air. This probably results from porphyrin-sensitized singlet oxygen formation and its successive addition to the anthracene moiety, similarly to previously reported anthraporphyrins and anthracene-substituted porphyrins.²⁷ However, protecting the corresponding solutions from light during purification we succeeded to isolate products **8** and **9** in pure form as evidenced by solution NMR and mass-spectroscopy data (see ESI†).

It should be noted, that the obtained product **9** obviously consists of a mixture of stereoisomers, resulting from the fact that the attached anthracene fragment is not symmetrical with the respect to the porphyrin mean plane. However, taking into account that the geometry of the photoactive fragment (porphyrin-bridge-anthracene) is identical for each of the possible isomers, their photophysical properties are unlikely to show individual features.

Optical properties

Absorption and emission spectra of compound **3** (Fig. 2A) generally resembled those of a reference compound, namely 9,10-diphenylanthracene (DPA) in its maxima, band structure and lifetimes. On the other hand, a considerably lower fluorescence quantum yield was observed (0.51). In order to obtain its phosphorescence spectrum, we performed a photo-

luminescence measurement at 77 K in frozen methyl-cyclohexane solution. In this case a red-shifted emission with a considerably longer lifetime (~300 ns) than the fluorescence (~14.9 ns) was observed (see Fig. S18 and S19†). However, a comparison with literature data shows, that the observed emission does not correspond to the anthracene triplet state, but rather to the emission of excimers.²⁸

UV/Vis and photoluminescence spectra of **8** and **9** are presented in Fig. 2. Similarity of the absorption features, *i.e.* maxima and vibrational structure, to those of octaethylporphyrin (OEP) and its Pd(II) complex (see Fig. S20 and S21† for comparison) confirms the absence of conjugation between porphyrin and anthracene. The only difference observed is a slight bandwidth broadening due to increased molecular size. Absorption of anthracene subunits is seen in the 300–400 nm region.

Upon excitation at the wavelength corresponding to the porphyrin Q-band absorption, compound **8** exhibits fluorescence consisting of two bands with maxima at 620 and 687 nm, typical of free base porphyrins, and virtually identical regarding peak maxima and vibronic structure to that obtained with the same excitation wavelength for OEP free-base. Fluorescence quantum yields were obtained as 0.17 and 0.13 for **8** and OEP, respectively (compared to tetraphenylporphyrin free-base as a reference). When excitation was done at 336 nm, in the region of the anthracene chromophore absorption, emission spectra showed predominantly emission from the



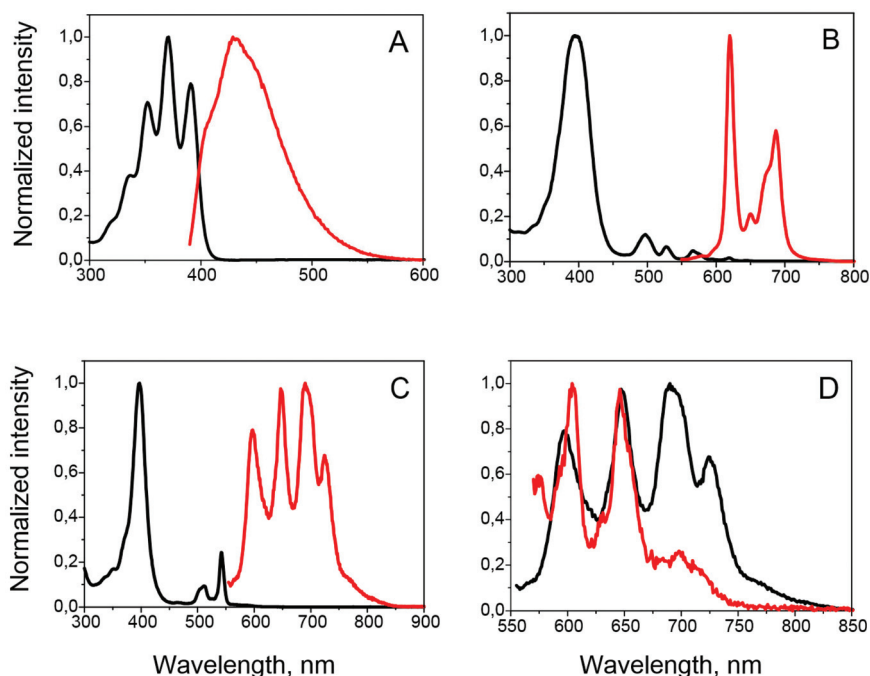


Fig. 2 Absorption (black) and emission (red) spectra of anthracene **3** (A), porphyrins **8** (B) and **9** (C). (D) Comparison of porphyrin **9** (3×10^{-5} M) emission under inert atmosphere (black) and under air (red).

porphyrin (Fig. S20[†]). An excitation spectrum registered at different emission wavelengths (Fig. S20[†]) closely resembles the absorption spectrum of **8**.

Emission spectra of **9** solution (5×10^{-6} M), prepared in glove-box conditions (1 ppm O_2), differ to those of the parent PdOEP (Fig. S21[†]) and consist of four bands at 597, 648, 689 and 724 nm (Fig. 2c). Upon selective excitation in the region of anthracene absorption, similar emission spectra were observed. Excitation spectra measured at all emission maxima are similar to the absorption spectrum of **9** (Fig. S21[†]). When the sample was left open to air, the emission beyond 670 nm was almost completely quenched (Fig. 2D), while the peaks at 597 and 648 nm were still observed, although with rather low intensity. Lifetimes of the emission at 597 and 648 nm were measured to be 602 and 648 μ s, respectively (Fig. S23[†]).

Excited state dynamics in PdOEP and porphyrin **9**

In order to further clarify the excited state dynamics, porphyrin **9** was studied by ps- μ s broadband transient absorption spectroscopy (TA); measurements on the pristine PdOEP were also performed and compared to those of **9**.

The measurements conducted on pristine PdOEP were done in toluene solution at a concentration of 10^{-4} M. After excitation into the Soret band at 400 nm, porphyrin singlet states are generated on a time scale faster than the TA instrument response. The respective sub-ps transient absorption spectrum is shown in Fig. 3. The prominent peaks between 500–550 nm are assigned to the bleaching of the ground state, as they perfectly well match the Q-band ground state absorp-

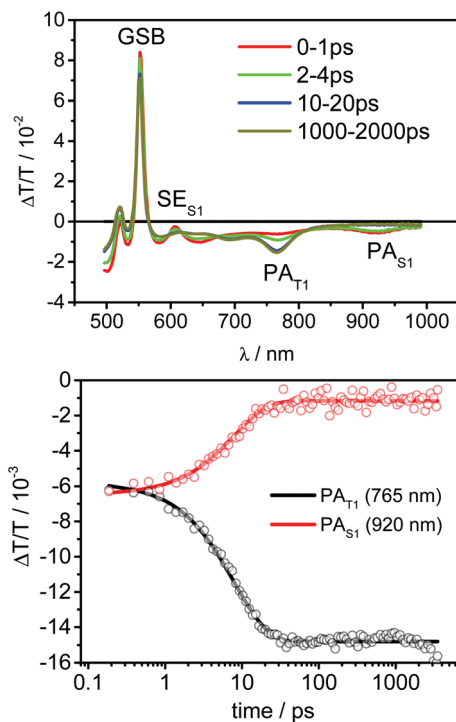


Fig. 3 Transient absorption spectra of PdOEP after excitation at 400 nm and kinetics of the singlet and triplet population.

tion spectrum. The positive feature at 610 nm results from stimulated emission (SE) of the porphyrin S_1 states (PS_1) and can be used as a measure for the contribution of singlet



excited states to the total excited state population. Between 850–1000 nm the photoinduced absorption (PA_{S_1}) of the singlets shows another prominent peak, which consequently can be used to monitor the dynamics of the excited singlet state population. After a few ps both the SE and the PA_{S_1} of the singlets have vanished and another broad PA has appeared with a distinct peak between 740–790 nm. As this feature stayed constant throughout the time window of the TA experiment (4 ns), we assigned it to long-lived porphyrin T_1 states (PT1). Thus the initially generated porphyrin singlet excitons undergo intersystem crossing to form porphyrin triplet states. Global fitting of the dataset with a single exponential function described the decay of the singlet excited states and concomitant rise of the triplet state population very well, allowing us to extract a rate constant for the intersystem crossing, as shown in the bottom of Fig. 3. The kinetics were extracted at the indicated spectral regions after the initial photogeneration of the singlet excitons (faster than the response time of the setup). The solid lines represent a monoexponential fit with a rate constant of $k_{ISC} = 1/(7.97) \text{ ps}^{-1}$ obtained from a global fit of the whole data set. The correlated decrease of porphyrin singlet PA_{S_1} and increase of porphyrin triplet PA_{T_1} allows to assign this rate constant to the ISC process. The obtained individual component spectra for the two states are shown in the ESI.†

With the knowledge of the singlet and triplet state transient absorption spectra, we interpreted the transient absorption data of compound **9** obtained in solution at a concentration of 10^{-4} M. Fig. 4 shows the respective transient absorption spectra.

At early times, precisely 0–4 ps, porphyrin singlets can be identified by their characteristic SE peak around 600 nm and

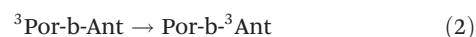
the photoinduced absorption feature at 850–950 nm. Both features disappear on the same time scale as observed for the PdOEP indicating triplet state formation on the porphyrin. However, the triplet signatures quickly decayed in contrast to the pristine PdOEP, and a very weak residual signal remained. The fast decay of the triplet signatures indicates efficient triplet energy transfer to the anthracene, which we modeled by a sequential model including two processes that follow single exponential dynamics, of which one describes the ISC and the other the triplet energy transfer from the porphyrin core to the adjacent anthracene.

Fig. 4 displays the kinetics in the region of SE of the porphyrin singlets (600 nm) and the PA_{T_1} peak region of the porphyrin triplets (765 nm) together with the fit to the experimental data. Due to the superposition of the individual photoinduced absorption signatures, an initial signal rise can be observed, corresponding to the ISC process from porphyrin singlets to porphyrin triplets. After the ISC is completed the triplet population stays constant for more than 2 ns in the pristine PdOEP, while in compound **9** triplet energy transfer to the anthracene takes place and rapidly reduces the porphyrin triplet population. The rise of the signals on the sub-10 ps time scale corresponds to the population of the porphyrin triplet state, which is subsequently transferred to the anthracene, causing the signal to decay up to 100 ps. The remaining weak PA then stayed constant in signal height over the measured time frame. Due to its long-lived character and different shape compared to the triplet-induced absorption of the porphyrin, we assigned this feature to triplet states on anthracene. The global fit yielded an ISC rate of $k_{ISC} = 1/8.97 \text{ ps}^{-1}$ and a triplet energy transfer rate of $k_{trans} = 1/14.1 \text{ ps}^{-1}$. The spectra obtained for the porphyrin singlet and triplet state from the global fit are in good agreement with those obtained from the pristine porphyrin and thus further validate our data analysis (Fig. S26 and S27†).

Excited state energy transfer processes in porphyrin **9**

The energetic overlap of the porphyrin and anthracene triplet manifolds in molecule **9** was confirmed by measuring the emission of **3** in a mixture with PdOEP (20:1) in toluene. Upon excitation at 542 nm, blue fluorescence (457 nm) was observed due to the TTA-UC process (Fig. 5). The efficiency of the process was found to be 7.5%, comparable with those of the PdOEP-DPA (12.5%).

Thus, intersystem crossing giving the triplet state localized on the porphyrin subunit (1) is followed by excited state triplet transfer to the anthracene, as was shown by TA data (2):



On the other hand, steady-state emission spectra and corresponding excitation spectra of **8** and **9** clearly indicate intramolecular excited state singlet energy transfer (3) from anthracene to the porphyrin core, since excitation in the region of anthracene absorption leads to the emission corres-

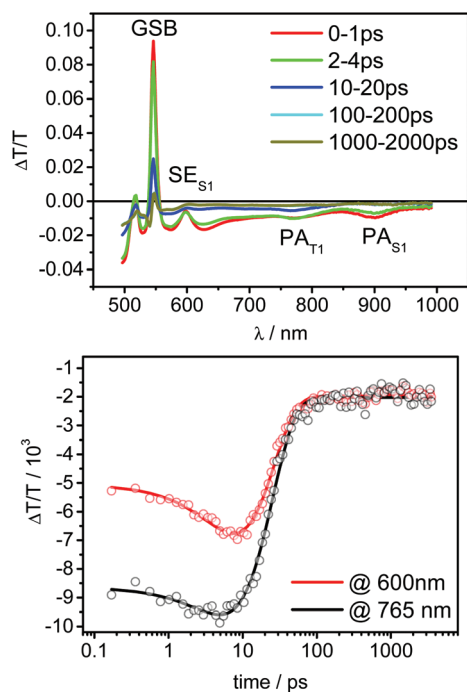


Fig. 4 Transient absorption spectra of compound **9** and kinetics observed at the indicated wavelength regions.



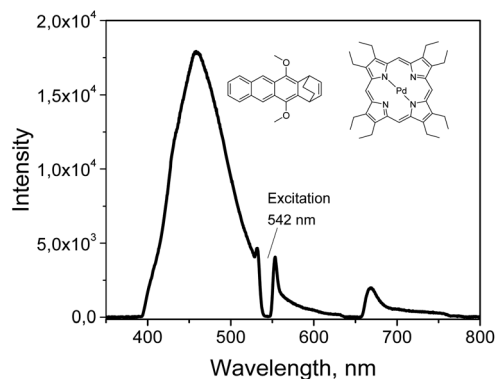


Fig. 5 Upconversion spectrum of PdOEP (10^{-5} M) and 5,12-dimethoxy-1,4-dihydro-1,4-ethanotetracene (5×10^{-3} M) in air-free toluene.

ponding to the porphyrin chromophore. This is consistent with the excited-state behaviour of the previously reported porphyrin-anthracene dyads.²⁹ Although direct excitation of both anthracene and porphyrin chromophores can't be excluded due to the overlap of the absorption, emission of the PdOEP-anthracene **3** mixture (1 : 4 ratio), coming predominantly from anthracene (Fig. S23[†]) upon excitation in the same region, confirms energy transfer in dyad **9**.



Comparison of the emission spectra of palladium porphyrin **9** with the reported fluorescence and phosphorescence spectra of anthracene **3** and other anthracenes (9,10-dimethoxy or 9,10-diphenylanthracene) shows that there is no contribution from the anthracene subunits to the observed emission comprised by a series of peaks at 597, 648, 689 and 724 nm. Analytical data, suggesting reasonable purity, in conjunction with the good agreement of excitation and absorption spectra allow to rule out that some of these peaks are resulting from impurities. Alternatively, the fact that the emission peaks at 597 and 648 nm were observed in oxygen-saturated conditions, when the phosphorescence is completely quenched, allows to assign this emission as a fluorescence from the singlet state of the porphyrin,³⁰ which is normally very weak for Pd and Pt porphyrins due to the high intersystem crossing efficiency.³¹

We conclude that the observed spectrum of **9** represents a combination of the emission from singlet and triplet excited states. Intensive singlet emission in degassed solution of **9** may result either from triplet-triplet annihilation or incomplete intersystem crossing.³² The latter can be however excluded in the case of molecule **9** based on the transient absorption studies results. We assigned the emission at 597 nm as a delayed fluorescence, while the emission at 648 nm as a superposition of the delayed fluorescence and phosphorescence. Indeed, similarly long-lived singlet emission was observed for PdOEP, although with much lower intensity compared to its phosphorescence.³³

To prove the formation of porphyrin singlet states we performed long delay transient absorption measurement, *i.e.* at

ns- μ s timescale. TA spectra obtained after 1 ns are in agreement with the proposed process. After 1 μ s the emergence of a clear positive signal between 550–600 nm (Fig. S28[†]) is seen. Due to its spectral position this signal can only be assigned to the porphyrin's singlet state. The lifetime of this signal is in the range of hundreds of microseconds (Fig. S29[†]), which is in line with the observation of long-lived photoluminescence from compound **9** in the same spectral region and supports our assignment. However, it remains unclear why the singlet states do not undergo rapid intersystem crossing to the triplet state as observed upon direct photoexcitation of the porphyrin's singlet state.

Intermolecular energy transfer processes, such as TTET, are concentration dependent. Indeed, registering the emission of **9** at different concentrations shows changes in the delayed fluorescence intensity, which almost completely disappears upon strong dilution. At 10^{-6} M concentration emission spectrum of **9** becomes nearly identical to those of PdOEP (Fig. 6), due to a loss of intermolecular TTA process efficiency.

Influence of the concentration on the emission spectra, *i.e.* intense delayed fluorescence at high concentration, evidences that intermolecular energy transfer between the molecules takes place in solution. Taking into account previous findings on triplet excited state relaxation channels,³³ this confirms that triplet-triplet annihilation between two anthracene localized triplet states takes place, which results in the formation of a singlet excited state of anthracene:



Such process is also consistent with the reported photo-physical behavior of 9,10-diphenylanthracene appended ruthenium(III) complex structures, which undergo TTA process in the excited state.¹⁰

As shown in the steady-state emission experiments, singlet energy transfer from the anthracene to the porphyrin is very

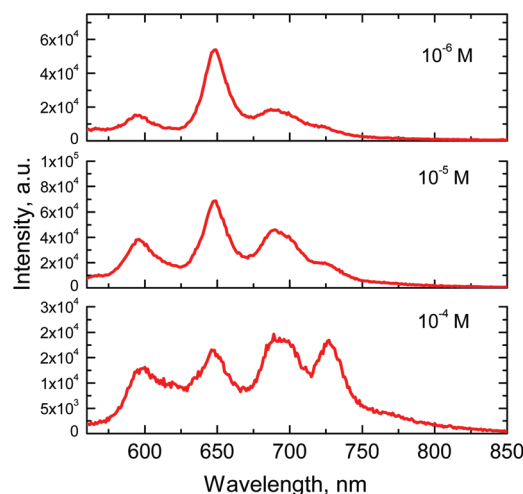


Fig. 6 Emission spectra of porphyrin **9** at different concentrations in toluene. Excitation wavelength – 542 nm.



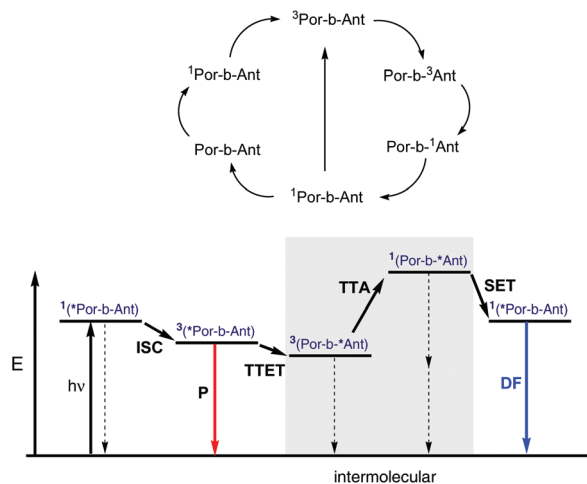


Fig. 7 Transitions among excited states of **9** and qualitative Jablonski diagram. $h\nu$ – photon excitation, ISC – intersystem crossing, P – phosphorescence, TTET – triplet–triplet energy transfer, TTA – triplet–triplet annihilation, SET – singlet energy transfer, DF – delayed fluorescence.

efficient. Thus TTA process (4) should be followed by the formation of porphyrin S_1 states *via* intermolecular energy transfer process (2). Although the intersystem crossing is very fast, emission of the porphyrin singlet is observed due to recombination to the ground state. ns– μ s TA experiments on compound **9** further corroborate porphyrin singlet excited state formation.

The observed processes are summarized in Fig. 7.

An additional point is noteworthy: in the spectra of compound **9** no upconverted luminescence coming from the anthracene subunits was observed. This is in contrast with previous results reported for Ru(III) cyclometallated complexes, surrounded by diphenylanthracenes.¹⁰ A possible explanation could be that it is the short distance between donor and acceptor subunits in **9**, which promotes fast energy back transfer in the singlet excited state formed in the course of TTA.

It was of particular interest to investigate whether molecule **9** can act as a triplet sensitizer for TTA-UC process in spite of the observed interplay between the singlet and triplet excited states. Another important question was if the intramolecular TTET from porphyrin to anthracene subunit, forming anthracene localized triplet state, can facilitate TTA-UC in a two-component system containing “free” anthracene **3** in the solution (emitter). Fig. 8 shows upconversion spectra for the samples containing **9** (3×10^{-5} M) as a sensitizer and 5, 10 or 20 equivalents of **3** as an emitter. The spectra were registered at different concentrations of “free” anthracene **3** while keeping the sensitizer concentration constant. Efficiency of the process reaches 3.8% at excitation intensities >1.5 W cm^{-2} . However, in contrast to previously studied upconversion systems, which require excessive amounts of emitter with respect to amount of sensitizer used (usually 1:20–50), this couple shows strong emission even in 1:5 ratio (see Fig. S24[†] for comparison with PdOEP). Moreover, even in 1:1 ratio weak upconversion signal was still detected (Fig. S25[†]). This suggests that dimethoxyanthracene antennas are enhancing the rate of TTET or

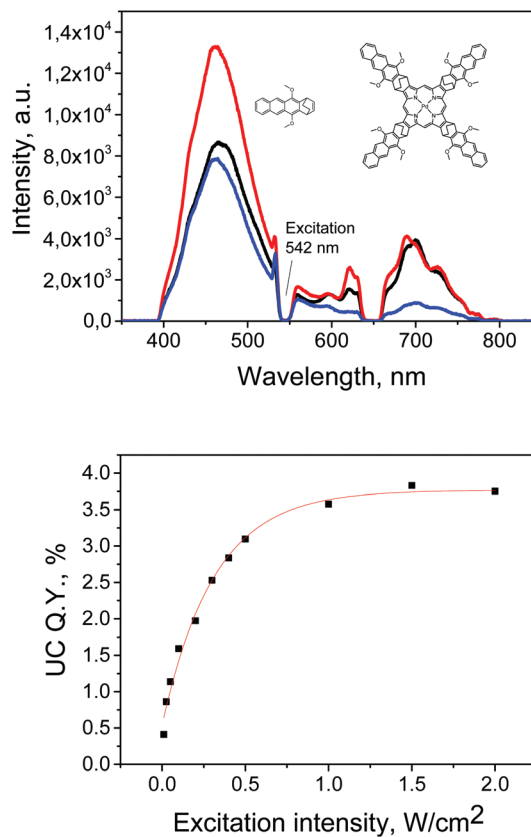


Fig. 8 Upconversion luminescence of the mixture of **9** and **3** and quantum yield dependence on the excitation intensity (for the mixture of 6×10^{-4} M **3** and 3×10^{-5} M **9** respectively). Solvent: toluene. Samples were excited at $\lambda = 542$ nm (100 mW cm^{-2}).

triplet–triplet annihilation as well, or both rates. However, this is beyond the scope of the present manuscript and will be reported in a separate study.

Excited state behavior of **9** is intriguing from the viewpoint of chromophores design for photonic applications. Delayed fluorescence has been demonstrated as a tool for a real-time temperature probe with ratiometric response for a broad temperature interval.³⁴ The main limitation of this approach is the relatively weak intensity of the delayed fluorescence of the metallated macrocycles, compared to phosphorescence signal. To increase the intensity of delayed fluorescence, encapsulation into polymer films (*e.g.* polystyrene), was applied.³³ Although steric effects of the triplet–triplet annihilation process have not been adequately studied so far, experimental observations suggest that the TTA efficiency is critically dependent on the mutual orientation of two annihilating triplet excited state molecules. In particular, simulations showed that the magnetic field effect on TTA is very sensitive to mutual chromophore alignment, and the direction of the effect is consistent with a local ordering of excited state molecules.³⁵ The fact that molecule **9** possesses strong delayed fluorescence in solution, may imply involvement of the factor of mutual orientation between annihilating excited states in the inter-



molecular TTA process. For further investigation of these steric factors, more dyad molecules with distinct geometries are to be prepared and investigated.

Conclusions

In conclusion, we have synthesized and characterized a conformationally rigid porphyrin-anthracene dyad that exhibits strong delayed fluorescence and phosphorescence in solution. Using broadband transient absorption spectroscopy, we have elucidated the formation of anthracene-localized triplet states *via* intramolecular energy transfer from the porphyrin core with an inverse rate constant of 14 ps and its further transformation into the porphyrin singlet state at μs – timescale. Excited state dynamics along with other spectroscopy data suggest that the porphyrin delayed fluorescence arises as a result of intermolecular triplet–triplet annihilation process. Observed properties of the prepared molecule demonstrate an example of singlet and triplet excitation management in donor–acceptor dyad and are of interest with respect to photon energy conversion systems.

Experimental

Equipments and methods

Anthracene-1,4-dione was prepared according to published procedure.³⁶ NMR spectra were recorded with the solvent proton or carbon signal as an internal standard. Emission quantum yields of the porphyrin compounds were measured relative to the fluorescence of free-base tetraphenylporphyrin ($\phi_f = 0.11$)³⁷ in deoxygenated toluene. For quantum yields measurements, sample concentrations were chosen to obtain an absorbance of 0.03–0.07 at the excitation wavelength, at least three measurements were performed for each sample. Emission of the diluted (1×10^{-6} M) rubrene solution in toluene under 542 nm excitation was used as a reference for UC quantum yield calculations. UC measurements were performed using previously described set-up.³⁸ All the measurements were performed using deoxygenated samples prepared in glovebox (1 ppm oxygen).

Transient absorption measurements were performed with a home-built pump–probe setup. For measurements in the time range of 1–3 ns with a resolution of ~ 100 fs the output of a commercial titanium:sapphire amplifier (3.5 mJ, 1 kHz, 100 fs) was split with one portion used to generate a 400 nm excitation pulse (second harmonic generation in a BBO crystal cut at $\theta = 29.2^\circ$) and another used to generate a white light probe using a home-built two-stage broadband (480–850 nm) non-collinear optical parametric amplifier for white light generation and amplification in the visible spectral range. The variable delay of up to ~ 3 ns between pump and probe was introduced by a broadband retroreflector mounted on a mechanical delay stage. Only reflective optics were used to guide the probe beam to the sample to minimize chirp. The excitation

pulse was chopped at 500 Hz, while the white light pulses were dispersed onto a linear photodiode array which was read out at 1 kHz. Adjacent diode readings corresponding to the transmission of the sample after an excitation pulse and without an excitation pulse were used to calculate $\Delta T/T$.

Preparation and characterization of the compounds

1,4,4a,12a-Tetrahydro-1,4-ethanotetracene-5,12-dione (2). A mixture of anthracene-1,4-dione **1** (12 g, 58 mmol), 1,3-cyclohexadiene (9.28 g, 118 mmol) and chloroform (20 mL) was placed into a high-pressure glass tube and heated at 80 °C for 48 h. After cooling to room temperature the reaction mixture was evaporated in vacuum. Crude product was recrystallized from ethanol. **2**: 13.4 g (80%), yellow crystals, m.p. 195–196 °C. ¹H NMR (250 MHz, CD₂Cl₂) δ 8.54 (s, 2H), 8.15–7.96 (m, 2H), 7.75–7.57 (m, 2H), 6.22–6.02 (m, 2H), 3.42–3.21 (m, 4H), 1.92–1.71 (m, 2H), 1.45–1.31 (m, 2H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 198.3, 135.6, 134.3, 132.2, 130.3, 129.7, 128.9, 51.4, 36.6, 25.4. Anal. Calcd for C₂₀H₁₆O₂: C, 83.31; H, 5.59. Found: C, 83.18; H, 5.68.

5,12-Dimethoxy-1,4-dihydro-1,4-ethanotetracene (3). A mixture of **2** (12 g, 42 mmol), methyl iodide (17.9 g, 126 mmol), anhydrous K₂CO₃ (11.6 g, 84 mmol), and acetone (150 mL) was refluxed for 30 h. The resulting mixture was filtered, concentrated by evaporation, and the residue was recrystallized from ethanol. **3**: 11.7 g (88%), pale yellow crystals, m.p. 174–175 °C. ¹H NMR (250 MHz, CD₂Cl₂) δ 8.61 (s, 2H), 8.04 (dd, $J = 6.4, 3.2$ Hz, 2H), 7.46 (dd, $J = 6.5, 3.2$ Hz, 2H), 6.61 (dt, $J = 7.8, 3.9$ Hz, 2H), 4.44 (d, $J = 1.3$ Hz, 2H), 4.01 (s, 6H), 1.64 (dt, $J = 29.2, 14.0$ Hz, 4H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 145.3, 135.6, 132.7, 131.8, 128.8, 127.1, 125.7, 121.2, 63.1, 34.1, 26.0. UV/Vis (toluene) λ_{max} (log ϵ): 391 (3.67), 371 (3.77), 352 (3.62), 337 (3.35). Anal. Calcd for C₂₂H₂₀O₂: C, 83.51; H, 6.37. Found: C, 83.59; H, 6.61.

2-Chloro-5,12-dimethoxy-3-(phenylsulfonyl)-1,2,3,4-tetrahydro-1,4-ethanotetracene (4). Synthesis was performed following a previously published general procedure for the preparation of α -chlorosulfones.³⁹ Thiophenol (3.52 g, 32 mmol) was added dropwise to a suspension of *N*-chlorosuccinimide (4.27 g, 32 mmol) in CH₂Cl₂ (50 mL). The mixture was stirred for 1 h, then the precipitate of succinimide formed was filtered off. The orange filtrate was added dropwise to a stirred solution of **3** (10 g, 32 mmol) in CH₂Cl₂ (100 mL) at 0 °C. The mixture was stirred at room temperature for 2 h, and then evaporated in vacuum. The residue was dissolved in THF–methanol (1 : 1) mixture (300 mL), and a suspension of Oxone (39.3 g, 64 mmol) in water (100 mL) was added under vigorous stirring. The mixture was stirred at room temperature for 3 days, diluted with water (200 mL), and extracted with CH₂Cl₂. The combined organic layers were dried with Na₂SO₄ and evaporated to dryness. Solid residue was recrystallized from ethanol. **4**: 13.41 g (85%), yellow semisolid froth. ¹H NMR (250 MHz, CD₂Cl₂) δ 8.69 (d, $J = 5.0$ Hz, 2H), 8.08 (dt, $J = 6.0, 3.5$ Hz, 2H), 7.93–7.80 (m, 2H), 7.61–7.41 (m, 5H), 4.36–4.29 (m, 1H), 4.23 (dd, $J = 4.8, 2.9$ Hz, 1H), 4.10 (s, 3H), 3.99 (s, 3H), 3.82 (dd, $J = 5.3, 2.9$ Hz, 1H), 3.64 (dd, $J = 5.6, 1.8$ Hz, 1H), 2.43 (m, 1H),



2.13–2.00 (m, 1H), 1.79–1.59 (m, 1H), 1.56–1.43 (m, 1H). ^{13}C NMR (63 MHz, CD_2Cl_2) δ 147.8, 147.3, 138.3, 134.5, 132.1, 131.9, 129.6, 129.6, 128.9, 128.9, 127.5, 127.4, 127.2, 126.3, 126.3, 126.0, 122.0, 121.8, 72.8, 63.6, 63.4, 57.0, 37.0, 29.3, 27.3, 18.7. Anal. Calcd for $\text{C}_{28}\text{H}_{25}\text{ClO}_4\text{S}$: C, 68.21; H, 5.11. Found: C, 68.63; H, 5.55.

5,12-Dimethoxy-2-(phenylsulfonyl)-1,4-dihydro-1,4-ethano-tetracene (5). Synthesis was performed following a modified literature procedure.⁴⁰ To a solution of chlorosulfone **4** (12 g, 24 mmol) in CH_2Cl_2 (100 mL) was added a solution of 1,8-diazabicycloundec-7-ene (3.83 g, 25 mmol) CH_2Cl_2 (10 mL), dropwise over a period of 10 min at 0 °C. The mixture was stirred for 1 h at room temperature, washed with water, dried with Na_2SO_4 , and evaporated in vacuum. Solid residue was recrystallized from ethanol. **5**: 9.97 g (91%), yellow semisolid froth. ^1H NMR (250 MHz, CD_2Cl_2) δ 8.58 (d, $J = 9.7$ Hz, 2H), 8.14–7.98 (m, 2H), 7.96–7.85 (m, 2H), 7.64–7.42 (m, 6H), 4.82–4.62 (m, 2H), 4.01 (s, 3H), 3.83 (s, 3H), 1.92–1.24 (m, 4H). ^{13}C NMR (63 MHz, CD_2Cl_2) δ 147.8, 146.4, 146.1, 144.9, 140.3, 134.0, 132.0, 131.9, 130.6, 129.9, 129.9, 129.3, 129.2, 128.8, 128.5, 128.5, 127.1, 127.0, 126.1, 121.7, 121.5, 63.5, 63.2, 35.7, 34.7, 26.6, 25.9. Anal. Calcd for $\text{C}_{28}\text{H}_{24}\text{O}_4\text{S}$: C, 73.66; H, 5.30. Found: C, 73.26; H, 5.68.

Ethyl 5,12-dimethoxy-4,13-dihydro-2H-4,13-ethano-anthra [2,3-f]isoindole-1-carboxylate (6). Barton–Zard synthesis of pyrrole **6** was performed following a previously published general procedure.⁴⁰ A solution of ethyl isocyanoacetate (2.03 g, 18 mmol) in THF (10 mL) was added dropwise over a period of 10 min to a stirred suspension of potassium *tert*-butoxide (2.13 g, 19 mmol) at 0 °C. The resulting mixture was stirred for 30 min, and then a solution of sulfone **5** (8 g, 17.5 mmol) in THF (20 mL) was added dropwise over a period of 10 min. The reaction mixture was stirred at room temperature for 4 h, then evaporated in vacuum, dissolved in CH_2Cl_2 (50 mL), washed with water, dried with Na_2SO_4 , and evaporated in vacuum. Solid residue was recrystallized from ethanol. **6**: 6.14 g (82%), white powder, m.p. 204–205 °C. ^1H NMR (250 MHz, CD_2Cl_2) δ 8.76 (s, 1H), 8.62 (d, $J = 5.4$ Hz, 2H), 8.03 (dt, $J = 9.4, 3.7$ Hz, 2H), 7.52–7.40 (m, 2H), 6.82–6.75 (m, 1H), 5.33–5.29 (m, 1H), 4.84 (d, $J = 2.0$ Hz, 1H), 4.44–4.28 (m, 2H), 4.05 (s, 3H), 4.02 (s, 3H), 1.90–1.76 (m, 4H), 1.44 (t, $J = 7.1$ Hz, 3H). ^{13}C NMR (63 MHz, CD_2Cl_2) δ 207.0, 161.9, 146.1, 145.6, 134.7, 133.0, 132.3, 131.7, 131.8, 130.0, 128.8, 128.8, 127.2, 127.1, 125.9, 125.8, 121.4, 121.3, 115.7, 114.7, 63.2, 63.0, 60.7, 31.8, 31.4, 31.1, 28.0, 27.3, 14.9. Anal. Calcd for $\text{C}_{27}\text{H}_{25}\text{NO}_4$: C, 75.86; H, 5.89. Found: C, 76.12; H, 6.14.

Porphyrim 8, free base. Pyrrole **6** (1 g, 2.34 mmol) was dissolved in 50 ml of dry THF under Ar atmosphere, and cooled to 0 °C. Then LiAlH_4 (0.89 g, 23 mmol) was added in portions and the stirring was continued 2 h. Water (5 mL) was added to the reaction mixture and the precipitate was filtered off using celite. The filtrate was diluted with water and extracted with CH_2Cl_2 . The organic layer was washed with water and brine, dried over Na_2SO_4 , and the solvent was removed in vacuum. The residue was dried in vacuum, and then dissolved in 300 ml of dry CHCl_3 . After the addition of *p*-toluenesulfonic

acid (0.01 g, 58 μmol), the reaction mixture was stirred for 18 h and DDQ (0.53 g, 2.34 mmol) was added. All the following manipulations were performed either in the dark or in vessels covered by aluminum foil. After the additional stirring for 18 h, the solution was washed with saturated NaHCO_3 solution, water and brine, dried over Na_2SO_4 , and the solvent was removed in vacuum. The residue was purified by silica gel column chromatography (CH_2Cl_2). **8**: 214 mg (25%), brown powder, m.p. > 300 °C. ^1H NMR (500 MHz, CD_2Cl_2) δ 10.89–10.79 (m, 4H), 8.85–8.74 (m, 8H), 8.17–8.04 (m, 8H), 7.56–7.44 (m, 8H), 6.86 (d, $J = 8.8$ Hz, 8H), 4.65–4.33 (m, 24H), 2.76–2.44 (m, 16H). ^{13}C NMR (75 MHz, CD_2Cl_2 – trifluoroacetic acid) δ 148.9, 147.7, 147.6, 138.0, 132.4, 132.3, 131.0, 130.7, 129.0, 129.0, 128.9, 127.6, 127.3, 126.4, 126.3, 126.0, 122.1, 64.1, 34.6, 30.7, 28.1. UV/Vis (toluene) λ_{max} (log ϵ): 394 (5.3), 497 (4.37), 528 (4.1), 566 (3.96), 619 (3.45). HRMS (ESI-TOF): m/z found 1463.5874, calcd for $[\text{M}^+]$ $\text{C}_{100}\text{H}_{78}\text{N}_4\text{O}_8$ 1463.5853.

Porphyrim 9. All the following manipulations were performed either in the dark or in vessels covered by aluminum foil. A mixture of porphyrim **8** (0.1 g, 0.068 mmol), bis(benzonitrile)palladium(II) chloride (0.052 g, 0.136 mmol), and benzonitrile (5 mL) was heated at 120 °C for 8 h under Ar atmosphere. Then the reaction mixture was evaporated to dryness in vacuum, dissolved in CH_2Cl_2 (10 ml) and filtered through a layer of silica (eluent CH_2Cl_2). Filtrate was evaporation in vacuum. **9**: 91 mg (85%), purple powder, m.p. > 300 °C. ^1H NMR (250 MHz, d^8 -toluene) δ 11.04–10.74 (m, 4H), 8.86 (d, $J = 10.9$ Hz, 8H), 8.01–7.73 (m, 8H), 7.30–7.15 (m, 8H), 6.69 (dd, $J = 20.9, 13.2$ Hz, 8H), 4.37–3.95 (m, 24H), 2.68–2.15 (m, 16H). ^{13}C NMR was not registered because of insufficient solubility. UV/Vis (toluene) λ_{max} (log ϵ): 397 (5.3), 512 (4.26), 542 (4.68). HRMS (ESI-TOF): m/z found 1567.4713, calcd for $[\text{M}^+]$ $\text{C}_{100}\text{H}_{76}\text{N}_4\text{O}_8\text{Pd}$ 1567.4731.

Acknowledgements

M. Filatov acknowledges the European Commission for Marie Curie Individual Fellowship (CONSORT, Grant No. 655142). S. Balushev acknowledges the financial support from the Bulgarian National Science Fund (DFNI E 02/11 – *SunStore*-project “Molecular solar thermal systems, enhanced by annihilation upconversion”) and from the FCFP FRIAS COFUND Fellowship Programme (FP7-MCA-609305). F. Laquai thanks the Max Planck Society for funding the Max Planck Research Group. F. Etzold acknowledges financial support in the framework of the SPP1355 “Elementary Processes in Organic Photovoltaics”. D.W. Gehrig thanks the Fonds der Chemischen Industrie (FCI) for a Kekulé scholarship.

Notes and references

- 1 D. Gust, T. A. Moore and A. L. Moore, *Acc. Chem. Res.*, 2001, **34**, 40; H. Yersin, A. F. Rausch, R. Czerwieniec, T. Hofbeck and T. Fischer, *Coord. Chem. Rev.*, 2011, **255**, 2622; P. Duan,



- N. Yanai, H. Nagatomi and N. Kimizuka, *J. Am. Chem. Soc.*, 2015, **137**, 1887; F. N. Castellano, *Acc. Chem. Res.*, 2015, **48**, 828; M. Filatov, S. Ritz, I. Ilieva, V. Mailänder, K. Landfester and S. Balushev, SPIE Newsroom. DOI: 10.1117/2.1201403.005378. Published Online: 7 April 2014; S. Balushev, T. Miteva, V. Yakutkin, G. Nelles, A. Yasuda and G. Wegner, *Phys. Rev. Lett.*, 2006, **97**, 143903.
- J. Zhao, W. Wu, J. Suna and S. Guo, *Chem. Soc. Rev.*, 2013, **42**, 5323; S. Guo, W. Wu, H. Guo and J. Zhao, *J. Org. Chem.*, 2012, **77**, 3933; X. Cui, J. Zhao, Z. Lou, S. Li, H. Wu and K. Han, *J. Org. Chem.*, 2015, **80**, 568; D. Huang, J. Sun, L. Ma, C. Zhang and J. Zhao, *Photochem. Photobiol. Sci.*, 2013, **12**, 872; P. D. Frischmann, K. Mahata and F. Würthner, *Chem. Soc. Rev.*, 2013, **42**, 1847; S. Y. Takizawa, K. Shimada, Y. Sato and S. Murata, *Inorg. Chem.*, 2014, **53**, 2983; T. Takahashi, K. Shizu, T. Yasuda, K. Togashi and C. Adachi, *Sci. Technol. Adv. Mater.*, 2014, **15**, 034202.
 - A. Harriman, M. Hissler, A. Khatyr and R. Ziessel, *Chem. Commun.*, 1999, 735; N. D. McClenaghan, F. Barigelletti, B. Maubert and S. Campagna, *Chem. Commun.*, 2002, 602; D. S. Tyson, J. Bialecki and F. N. Castellano, *Chem. Commun.*, 2000, 2355; A. F. Morales, G. Accorsi, N. Armaroli, F. Barigelletti, S. J. A. Pope and M. D. Ward, *Inorg. Chem.*, 2002, **41**, 6711; B. Maubert, N. D. McClenaghan, M. T. Indelli and S. Campagna, *J. Phys. Chem. A*, 2003, **107**, 447; D. S. Tyson and F. N. Castellano, *J. Phys. Chem. A*, 1999, **103**, 10955; S. G. Camera and H. E. Toma, *J. Photochem. Photobiol., A*, 2002, **151**, 57.
 - P. Duan, N. Yanai and N. Kimizuka, *Chem. Commun.*, 2014, **50**, 13111; J. Peng, X. Jiang, X. Guo, D. Zhao and Y. Ma, *Chem. Commun.*, 2014, **50**, 7828.
 - A.-M. Stadler, F. Puntoriero, F. Nastasi, S. Campagna and J.-M. Lehn, *Chem. – Eur. J.*, 2010, **16**, 5645; F. Laquai, Y.-S. Park, J.-J. Kim and T. Basche, *Macromol. Rapid Commun.*, 2009, **30**, 1203; B. Geiß and C. Lambert, *Chem. Commun.*, 2009, 1670; M. O. Senge, M. Fazekas, M. Pinteá, M. Zawadzka and W. J. Blau, *Eur. J. Org. Chem.*, 2011, 5797.
 - Z. Liu, W. He and Z. Guo, *Chem. Soc. Rev.*, 2013, **42**, 1568; Q. Zhao, F. Li and C. Huang, *Chem. Soc. Rev.*, 2010, **39**, 3007; O. S. Finikova, P. Chen, Z. Ou, K. M. Kadish and S. A. Vinogradov, *J. Photochem. Photobiol., A*, 2008, **198**, 75.
 - Q. Zhao, C. Huang and F. Li, *Chem. Soc. Rev.*, 2011, **40**, 2508; C. Wohnhaas, V. Mailänder, M. Droge, M. A. Filatov, D. Busko, Y. Avlasevich, S. Balushev, T. Miteva, K. Landfester and A. Turshatov, *Macromol. Biosci.*, 2013, **13**, 1422.
 - J. Zhou, Q. Liu, W. Feng, Y. Sun and F. Li, *Chem. Rev.*, 2015, **115**, 395.
 - T. F. Schulze and T. W. Schmidt, *Energy Environ. Sci.*, 2015, **8**, 103; V. Gray, D. Dzebo, M. Abrahamsson, B. Albinsson and K. Moth-Poulsen, *Phys. Chem. Chem. Phys.*, 2014, **16**, 10345; T. Ogawa, N. Yanai, A. Monguzzi and N. Kimizuka, *Sci. Rep.*, 2015, **5**, 10882.
 - D. V. Kozlov and F. N. Castellano, *Chem. Commun.*, 2004, 2860; G. Bergamini, P. Ceroni, P. Fabbri and S. Cicchi, *Chem. Commun.*, 2011, **47**, 12780.
 - A. Ambroise, J. Li, L. Yu and J. S. Lindsey, *Org. Lett.*, 2000, **2**, 2563; E. K. L. Yeow, P. J. Sentic, N. M. Cabral, J. N. H. Reek, M. J. Crossley and K. P. Ghiggino, *Phys. Chem. Chem. Phys.*, 2000, **2**, 4281; A. Osuka, J.-Y. Shin, R. Yoneshima, H. Shiratori, T. Ohno, K. Nozaki, Y. Nishimura, I. Yamazaki, S. Taniguchi, T. Shimizu and T. Okada, *J. Porphyrins Phthalocyanines*, 1999, **3**, 729; P. Brodard, S. Matzinger, E. Vauthey, O. Mongin, C. Papamicael and A. Gossauer, *J. Phys. Chem. A*, 1999, **103**, 5858; L. Flamigni, F. Barigelletti, N. Armaroli, B. Ventura, J.-P. Collin, J.-P. Sauvage and J. A. G. Williams, *Inorg. Chem.*, 1999, **38**, 661; K. Kils, J. Kajanus, J. Mårtensson and B. Albinsson, *J. Phys. Chem. B*, 1999, **103**, 7329.
 - S. Kawabata, I. Yamazaki and Y. Nishimura, *Bull. Chem. Soc. Jpn.*, 1997, **70**, 1125; R. Hermant, P. A. Liddell, S. Lin, R. G. Alden, H. K. Kang, A. L. Moore, T. A. Moore and D. Gust, *J. Am. Chem. Soc.*, 1993, **115**, 2080; A. Osuka, H. Yamada, K. Maruyama, N. Mataga, T. Asahi, M. T. Ohkouchi, T. Okada, I. Yamazaki and Y. Nishimura, *J. Am. Chem. Soc.*, 1993, **115**, 9439; J. S. Lindsey, P. A. Brown and D. A. Siesel, *Tetrahedron*, 1989, **45**, 4845; F. Effenberger, H. Schlosser, P. Bäuerle, S. Maier, H. Port and H. C. Wolf, *Angew. Chem., Int. Ed. Engl.*, 1988, **27**, 281; J. D. Megiatto Jr., D. I. Schuster, S. Abwandner, G. de Miguel and D. M. Guldi, *J. Am. Chem. Soc.*, 2010, **132**, 3847.
 - I. Andreu, I. Morera, F. Palumbo, G. Sastre, F. Boscá and M. Miranda, *Chem. Sci.*, 2015, **6**, 4035.
 - L. Giribabu, A. Ashok Kumar, V. Neeraja and B. G. Maiya, *Angew. Chem., Int. Ed.*, 2001, **40**, 3733.
 - M. O. Senge, *Chem. Commun.*, 2006, 243.
 - M. Sirish and B. G. Maiya, *J. Porphyrins Phthalocyanines*, 1998, **2**, 237; M. Sirish and B. G. Maiya, *J. Photochem. Photobiol., A*, 1995, **85**, 127; J. Seixas de Melo, A. J. F. N. Sobral, A. M. Rocha Gonsalves and H. D. Burrows, *J. Photochem. Photobiol., A*, 2005, **172**, 151.
 - M. O. Senge, *Chem. Commun.*, 2011, **47**, 1943; P. D. Harvey, M. A. Filatov and R. Guilard, *J. Porphyrins Phthalocyanines*, 2011, **15**, 1150; P. D. Harvey, A. Langlois, M. Filatov, D. Fortin, K. Ohkubo, S. Fukuzumi and R. Guilard, *J. Porphyrins Phthalocyanines*, 2012, **16**, 685.
 - R. E. Haddad, S. G. Azeau, J. Pecaut, J.-C. Marchon, C. J. Medforth and J. A. Shelnutt, *J. Am. Chem. Soc.*, 2003, **125**, 1253; C. Muzzi, C. J. Medforth, K. M. Smith, S.-L. Jia and J. A. Shelnutt, *Chem. Commun.*, 2000, 131; J. A. Shelnutt, in *The Porphyrin Handbook*, ed. K. M. Kadish, K. M. Smith and R. Guilard, Academic Press, Burlington, MA, 2000, vol. 7, ch. 50, p. 167.
 - L. Noss, P. A. Liddell, A. L. Moore, T. A. Moore and D. Gust, *J. Phys. Chem. B*, 1997, **101**, 458; A. Rosa, G. Ricciardi and E. J. Baerends, *J. Phys. Chem. A*, 2006, **110**, 5180; A. Y. Lebedev, M. A. Filatov, A. V. Cheprakov and S. A. Vinogradov, *J. Phys. Chem. A*, 2008, **112**, 7723.
 - S. Ito, H. Uno, T. Murashima and N. Ono, *Chem. Commun.*, 1999, 2275.
 - H. Uno, H. Watanabe, Y. Yamashita and N. Ono, *Org. Biomol. Chem.*, 2005, **3**, 448.



- 22 G. D. Scholes, K. P. Ghiggino, A. M. Oliver and M. N. Paddon-Row, *J. Am. Chem. Soc.*, 1993, **115**, 4345; S. Plunkett, K. J. Flanagan, B. Twamley and M. O. Senge, *Organometallics*, 2015, **34**, 1408.
- 23 J. Sanning, P. R. Ewen, L. Stegemann, J. Schmidt, C. G. Daniliuc, T. Koch, N. L. Doltsinis, D. Wegner and C. A. Strassert, *Angew. Chem., Int. Ed.*, 2015, **54**, 786.
- 24 N. Ono, H. Kawamura, M. Bougauchi and K. Maruyama, *Tetrahedron*, 1990, **46**, 7483.
- 25 O. S. Finikova, S. Y. Chernov, A. V. Cheprakov, M. A. Filatov, S. A. Vinogradov and I. P. Beletskaya, *Dokl. Chem.*, 2003, **391**, 222; M. A. Filatov, A. Y. Lebedev, S. A. Vinogradov and A. V. Cheprakov, *J. Org. Chem.*, 2008, **73**, 4175; A. V. Cheprakov and M. A. Filatov, *J. Porphyrins Phthalocyanines*, 2009, **13**, 291.
- 26 S. Ito, N. Ochi, T. Murashima, N. Ono and H. Uno, *Chem. Commun.*, 2000, 893; T. Okujima, T. Kikkawa, H. Nakano, H. Kubota, N. Fukugami, N. Ono, H. Yamada and H. Uno, *Chem. – Eur. J.*, 2012, **18**, 12854; Y. Tomimori, T. Okujima, T. Yano, S. Mori, N. Ono, H. Yamada and H. Uno, *Tetrahedron*, 2011, **67**, 3187; H. Uno, H. Uoyama, C. Chenxin, H. Tahara, Y. Shimizu, H. Hagiwara, Y. Hanasaki, H. Yamada and T. Okujima, *Heterocycles*, 2010, **80**; M. A. Filatov, E. Heinrich, K. Landfester and S. Balushev, *Org. Biomol. Chem.*, 2015, **13**, 6977.
- 27 M. A. Filatov, S. Balushev, I. Z. Ilieva, V. Enkelmann, T. Miteva, K. Landfester, S. E. Aleshchenkov and A. V. Cheprakov, *J. Org. Chem.*, 2012, **77**, 11119; M. A. Filatov, E. Heinrich, D. Busko, I. Z. Ilieva, K. Landfester and S. Balushev, *Phys. Chem. Chem. Phys.*, 2015, **17**, 6501.
- 28 T. Seko, K. Ogura, Y. Kawakami, H. Sugino, H. Toyotama and J. Tanaka, *Chem. Phys. Lett.*, 1998, **291**, 438.
- 29 M. Ezo, T. Minami, Y. Ogawa, S. Yagi, H. Nakazumi, T. Matsuyama, K. Wada and H. Horinaka, *Photochem. Photobiol. Sci.*, 2005, **4**, 641–646; E. J. Shin, *Bull. Korean Chem. Soc.*, 2006, **27**, 751; L. Giribabu and B. G. Maiya, *Res. Chem. Intermed.*, 1999, **25**, 769; L.-P. Long, M.-X. You, H. Wang, Y.-X. Wang and R.-H. Yang, *Sci. China Ser. B: Chem.*, 2009, **52**, 793.
- 30 Y. Y. Cheng, T. Khoury, R. G. Clady, M. J. Tayebjee, N. J. Ekins-Daukes, M. J. Crossley and T. W. Schmidt, *Phys. Chem. Chem. Phys.*, 2010, **12**, 66.
- 31 D. Madge, M. W. Windsor, D. Holten and M. Gouterman, *Chem. Phys. Lett.*, 1974, **29**, 183.
- 32 J. B. Birks, *Photophysics of Organic Molecules*, Wiley, New York, 1970, p. 372.
- 33 A. Turshatov and S. Balushev, *Triplet–Triplet Annihilation-Assisted Upconversion: All-Optical Tools for Probing the Physical Parameter of Soft Matter Handbook of Coherent-Domain Optical Methods*, Springer Science + Business Media, New York, 2013, ISBN-13: 978-1461451754.
- 34 T. Miteva, G. Nelles, A. Yasuda, S. Balouchev and J. Lupton, *U.S. Pat.*, 7517144B2, Apr. 14, 2009; G. A. Baker, S. N. Baker and T. M. McCleskey, *Chem. Commun.*, 2003, 2932; D. N. Messias, M. V. D. Vermelho, A. S. Gouveia-Neto and J. S. Aitchison, *Rev. Sci. Instrum.*, 2002, **73**, 476.
- 35 G. B. Piland, J. J. Burdett, D. Kurunthu and C. J. Bardeen, *J. Phys. Chem. C*, 2013, **117**, 1224.
- 36 D. H. Hua, M. Tamura, X. Huang, H. A. Stephany, B. A. Helfrich, E. M. Perchellet, B. J. Sperfslage, J.-P. Perchellet, S. Jiang, D. E. Kyle and P. K. Chiang, *J. Org. Chem.*, 2002, **67**, 2907.
- 37 P. G. Seybold and M. Gouterman, *J. Mol. Spectrosc.*, 1969, **31**, 1.
- 38 A. Turshatov, D. Busko, Y. Avlasevich, T. Miteva, K. Landfester and S. Balushev, *ChemPhysChem*, 2012, **13**, 3112.
- 39 P. B. Hopkins and P. L. Fuchs, *J. Org. Chem.*, 1978, **43**, 1208.
- 40 O. S. Finikova, A. V. Cheprakov, P. J. Carroll and S. A. Vinogradov, *J. Org. Chem.*, 2003, **68**, 7517.

