We discovered that positively charged terbium complexes bearing 1,4,7,10-tetraazacyclodecane functionalized with amide ligands are highly sensitive to dynamic luminescence quenching by NAD(P)H. We exploited this phenomenon to establish a general time-resolved luminescence-based assay platform for sensitive detection of NAD(P)H-dependent enzyme activities.

Reduced nicotinamide adenine dinucleotide (phosphate), or NAD(P)H, is an important cofactor of oxidoreductases, which transfer electrons from NAD(P)H to their substrates, affording NAD(P)⁺ and the products. These enzymes function in many metabolic pathways, and are considered potential therapeutic targets.¹,² Assay of their activities is also helpful for clinical diagnosis.³ However, current assays of NAD(P)H-dependent enzyme activities are mostly based on absorption measurement, and this strategy has many drawbacks. For example, monitoring the absorption of NAD(P)H itself at 340 nm, which is a common approach, gives poor sensitivity in small volumes, such as 384- or 1536-well microplates,⁴ and absorbance in the UV region is influenced by many extraneous factors. Colouring of tetrazolium targets.¹,² Assay of their activities is also helpful for clinical diagnosis.³ However, current assays of NAD(P)H-dependent enzyme activities are mostly based on absorption measurement, and this strategy has many drawbacks. For example, monitoring the absorption of NAD(P)H itself at 340 nm, which is a common approach, gives poor sensitivity in small volumes, such as 384- or 1536-well microplates,⁴ and absorbance in the UV region is influenced by many extraneous factors. Colouring of tetrazolium salts upon NAD(P)H-dependent reduction is also used in cell proliferation and toxicity assays, but chemical instability of these reagents⁴ limits their applicability. Fluorescence measurement of NAD(P)H at 460 nm is another option, but is rarely used in practice, partly due to poor sensitivity at low NAD(P)H concentrations and an inner filter effect at high NAD(P)H concentrations.⁴ Although a few chemodosimeters for NAD(P)H have been reported,⁵ they are not suitable for monitoring enzyme activity in real time. Hence, we thought that a novel strategy to detect NAD(P)H that overcomes these disadvantages would have many potential applications, especially for high-throughput screening (HTS) in drug discovery and for clinical diagnostic tests.

For this purpose, we focused on luminescent lanthanide complexes. Lanthanide (Tb³⁺ or Eu³⁺) complexes have long luminescence lifetimes of millisecond order,⁶ which enables sensitive detection by measuring only the long-lived component of luminescence (i.e., time-resolved measurement). By exploiting this capability, functional luminescent probes for various analytes including ions⁷ and enzymes⁸ have been developed,⁹ but no luminescent probes for NAD(P)H have yet been reported. Interestingly, Parker et al. recently showed that some lanthanide complexes are susceptible to dynamic quenching by a few biological reductants such as ascorbate and urate, probably via exciplex formation.¹⁰,¹¹ Because they were interested in the use of luminescent complexes in cells, they regarded this quenching as undesirable, with one exception,¹² and they developed complexes that were tolerant of reductants.¹⁰ However, we thought that we could make use of this kind of intermolecular quenching to develop a novel and general strategy to detect NAD(P)H-dependent enzyme activity. In principle, this should be feasible, because NAD(P)H is one of the most potent biological reductants (Table S1, ESI†), which would efficiently quench luminescence, whereas NAD(P)⁺, an oxidized form of NAD(P)H, would not. Herein, we show that Tb³⁺ complexes are indeed dynamically quenched by NAD(P)H and the magnitude of the quenching can be modulated by adjusting the overall charge of the complex (Fig. 1a). Based on this finding, we have developed a novel general luminescence intensity- and lifetime-based assay for NAD(P)H-dependent enzyme activities.

We first hypothesized that the efficiency of the expected quenching by NAD(P)H would depend on the structure of the...
Communication

Dynamic quenching

Electrostatic attraction

NAD(P)H

Net charge +3

Net charge -2 (-4)

Fig. 1 (a) Schematic representation of the interaction of the Tb$^{3+}$ complex and NAD(P)H. (b) Chemical structure of complexes.

chelating moiety, in particular its charge. To test this idea, we designed and synthesized three Tb$^{3+}$ complexes with different net charges (Fig. 1b). Negatively charged NAD(P)H is expected to show greater electrostatic attraction towards a more positively charged complex, as previously reported for ATP.13 As a scaffold complex, we selected A3Tb, previously reported by Parker et al.,11b,12 and prepared a series of side-chain derivatives with altered net charge, because 1,4,7,10-tetraazacyclododecane and its derivatives form kinetically and thermodynamically stable complexes with lanthanide ions.14 Three of the ring nitrogen atoms were substituted with amide and/or carboxylate groups that coordinate to Tb$^{3+}$. Azaxanthone was used as an antenna moiety because it sensitizes Tb$^{3+}$ with high efficiency.15 The changes in luminescence intensity and lifetime of A1Tb, the complex with the highest positive charge, upon titration of NADH are illustrated in Fig. 2. Notably, the values of both parameters were greatly decreased in the presence of as little as 1 μM NADH, and the addition of 100 μM NADH almost totally quenched the luminescence. The quenching was concentration-dependent, and the Stern–Volmer constants ($K_{sv}$) of luminescence intensity and lifetime were almost identical at low NADH concentrations (Fig. S1, ESI†), indicating that the quenching process primarily occurs in the excited state of the Tb$^{3+}$.16 The data demonstrate that Tb$^{3+}$ complexes with appropriate structures are indeed dynamically quenched by NADH with high efficiency. On the other hand, complexes with less positive charge and neutral charge, A2Tb and A3Tb, respectively, showed only moderate decreases in luminescence intensity and lifetime upon titration of NADH up to 100 μM (Fig. S2 and S3, ESI†).

We further investigated the photophysical properties of these Tb$^{3+}$ complexes and the bimolecular quenching constants ($k_q$) upon titration of NADH. No significant differences were observed in the shapes of the absorption spectra and emission spectra, which may imply that the coordination structures of the three complexes are very similar (Fig. S4 and Table S2, ESI†). As summarized in Table 1, the more positively charged complex was more sensitive to NADH. The trend was more clearly seen in quenching of A1Tb by NADPH, which possesses greater negative charge than NADH, and is therefore expected to have stronger electrostatic interaction than NADH with positively charged compounds [compare Tables S3 and S4, ESI†]. On the other hand, 100 μM NAD$^+$ had little effect on luminescence intensity and no effect on lifetime (Table S5, ESI†). We also measured the effect of several adenine-containing biological compounds, and showed that they do not cause dynamic quenching of the complex (Table S5, ESI†). All these data suggest that electrostatic attraction between Tb$^{3+}$ and phosphate groups, as well as reductive electron transfer or charge-transfer exciplex formation between Tb$^{3+}$ and the reduced nicotinamide moiety at the excited state, is important for the dynamic quenching by NAD(P)H.17 However, considering the energy diagram of Tb$^{3+}$ complexes (Fig. S5, ESI†), which starts with antenna excitation, followed by intersystem crossing and intramolecular energy transfer, and results in the excited state of Tb$^{3+}$, it is possible that NAD(P)H also acts on the antenna moiety. Therefore, we further examined whether the excited state of the antenna is involved in the quenching process by direct excitation of the Tb$^{3+}$ ion at 488 nm (Tables S3 and S4, ESI†). The obtained quenching constants were the same for direct excitation (488 nm) and antenna excitation (340 nm), supporting the idea that NAD(P)H only acts in the excited state of Tb$^{3+}$. In accordance with this view, the fluorescence of the antenna moiety, 2-Me azaxanthone, was hardly quenched in the presence of 10 μM NADH (Fig. S6, ESI†). Importantly, the $k_q$ value of A1Tb for NAD(P)H (over $1 \times 10^8$ M$^{-1}$ s$^{-1}$) was much larger than those for GSH and ascorbate, so A1Tb is tolerant of these reductive reagents, which are often used in enzyme assay (Table S1, ESI†).

Next, we tested whether this quenching can be used to monitor enzyme reactions, because A1Tb responds to 1–100 μM NAD(P)H, which is the optimal range for applications such as HTS. The selected model enzyme, lactate dehydrogenase (LDH, EC 1.1.1.27), is a NADH-dependent enzyme that catalyzes the reduction of pyruvate to l-lactate while converting NADH into

<table>
<thead>
<tr>
<th>Probe</th>
<th>Charge</th>
<th>$\tau$ (ms)</th>
<th>$K_{sv}$ (M$^{-1}$)</th>
<th>$k_q$ (M$^{-1}$ s$^{-1}$)</th>
</tr>
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<tbody>
<tr>
<td>A1Tb</td>
<td>+3</td>
<td>1.52</td>
<td>$1.9 \times 10^1$</td>
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</tr>
<tr>
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<td>$4.0 \times 10^1$</td>
<td>$2.1 \times 10^6$</td>
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<tr>
<td>A3Tb</td>
<td>0</td>
<td>1.87</td>
<td>$1.7 \times 10^1$</td>
<td>$9.1 \times 10^5$</td>
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</tbody>
</table>

* $K_{sv}$ and $k_q$ were calculated from Stern–Volmer plots of luminescence lifetimes upon titration of NADH. Excitation wavelength was 340 nm.
NAD\(^+\) (Fig. 3a); it plays a key role in the anaerobic pathway of glucose metabolism.\(^7\) Because cancer cells tend to use this pathway instead of the aerobic pathway (Warburg effect), a specific subtype of this enzyme, LDH-A, is considered as an attractive target for anticancer drugs.\(^5\) The results of LDH assay with A1Tb in a cuvette are shown in Fig. 3b. Addition of LDH was accompanied by a monotonic increase in both luminescence intensity and lifetime of A1Tb, while no change was observed in the absence of the enzyme. Unaltered LDH activity in the presence of A1Tb was confirmed by measuring NADH observed in the absence of the enzyme. Unaltered LDH activity was accompanied by a monotonic increase in both luminescence (left) and NADH absorption (right) measurements on microplates after incubation for 25 minutes. Error bars represent S.D. (n = 3). Asterisks indicate p < 0.05 vs. 0 mM \(\text{mL}^{-1}\).

![Diagram of LDH assay using lanthanide luminescence.](Image)

**Fig. 3** (a) Schematic illustration of LDH assay using lanthanide luminescence. (b) Time course of luminescence intensity (left) or lifetime (right) of 4 \(\mu\text{M}\) A1Tb in the presence (black diamond) and absence (gray circle) of LDH (50 \(\mu\text{mL}^{-1}\)). Initial concentrations of pyruvate and NADH were 125 \(\mu\text{mL}^{-1}\) and 100 \(\mu\text{mL}^{-1}\), respectively. (c) Comparison of time-resolved luminescence (left) and NADH absorption (right) measurements on microplates after incubation for 25 minutes. Error bars represent S.D. (n = 3). Asterisks indicate p < 0.05 vs. 0 mM \(\text{mL}^{-1}\).

In conclusion, we have developed a novel assay platform for NAD(P)H-dependent enzymes using luminescent Tb\(^{3+}\) complexes. Though many luminescent probes for biological molecules have been reported,\(^7\)\(^9\)\(^12\) most of them are specific to limited analytes and do not have practical utility for general assays. In contrast, probes for coenzymes\(^13\) including NAD(P)H are expected to have high generality. We believe that this is the first report to describe the interaction between NAD(P)H and luminescent lanthanide complexes.\(^17\) We showed that NAD(P)H dynamically quenches the luminescence of Tb\(^{3+}\) complexes, and this phenomenon can be used to monitor the activities of NAD(P)H-dependent enzymes. Although it is not suitable for imaging cellular NADH, our probe can monitor enzyme reactions in real time with good sensitivity. In this communication, we have focused on optimizing the net charge of the complexes, and the influence of other factors still needs to be studied to elucidate the quenching mechanism in more detail. We are currently synthesizing Tb\(^{3+}\) complexes with other antenna or chelator moieties to optimize sensitivity, and we aim to apply them for inhibitor screening and clinical diagnostic assays of NAD(P)H-dependent enzymes.

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**Notes and references**


16 Replacing Tb$^{3+}$ with Eu$^{3+}$ greatly reduced the extent of quenching, as has been reported for other reductants.10,11

17 There is a report on the ground state reduction of the Dy$^{3+}$ complex by NADH on the film (T. Basova, I. Jushina, A. G. Gürek, V. Ahsen and A. K. Ray, *J. R. Soc., Interface*, 2008, 5, 801), but we do not believe this is directly relevant to our probes, because our probes showed no change in absorption spectra.