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Re and ^{99m}Tc complexes of BodP₃ – multi-modality imaging probes†

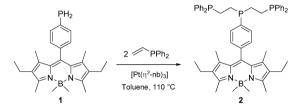
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A fluorescent tridentate phosphine, BodP₃ (2), forms rhenium complexes which effectively image cancer cells. Related technetium analogues are also readily prepared and have potential as dual SPECT/fluorescent biological probes.

^{99m}Tc-phosphine complexes are used in approved and emerging Single-Photon Emission Computed Tomography (SPECT) imaging agents due to the attractive nuclear properties of 99m Tc (γ = 140 keV, $t_{1/2}$ = 6 h) and to the facile formation of inert Tc-P coordinate bonds.¹ Resolving the cellular fate of such radiopharmaceuticals remains challenging due to the spatial limitations of SPECT, thus there is a drive to develop novel probes with fluorescent tags in order to facilitate high-resolution imaging by fluorescence microscopy.² Such a species has to be (i) kinetically inert, (ii) highly fluorescent upon metal ligation, and (iii) resistant to degradation by biological molecules. Here we report our work on developing phosphorus-based probes for this and related applications, including therapeutics. We recently described the synthesis of tridentate phosphine 2, BodP₃, (Scheme 1) from air-stable 1, with both retaining the attractive photophysical properties common to Bodipy.3 Bianchini reported the synthesis of the single isomer cis, fac-[ReCl(CO)2(triphos-Me)] by refluxing mer-[ReCl(CO)₃(PPh₃)₂] with 1,1,1-tris(diphenylphosphinomethyl)ethane.4 With this in mind, 2 was reacted with mer-[ReCl(CO)₃(PPh₃)₂] under similar conditions; however, a mixture of three stereoisomers was generated, 3a-c.

These were separated via chromatography and all three were then characterised by X-ray crystallography (Fig. 1 and ESI†).

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Scheme 1 The hydrophosphination reaction between primary phosphine 1 and vinyldiphenylphosphine, to produce the tridentate derivative 2, BodP₃.

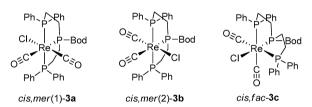


Fig. 1 The three isomers isolated from the reaction of the tridentate phosphine 2 and [ReCl(CO)₃(PPh₃)₂] in refluxing toluene.

Bis(diphenylphosphinoethyl)phenylphosphine (triphos-Ph) reacts with [ReCl(CO)₅] in refluxing toluene to form the related isomers reported here, but on prolonged reaction times (>24 h) at 170 °C, only the cis,mer(1) isomer was produced.⁵ Phosphine 2 was therefore reacted with mer-[ReCl(CO)₃(PPh₃)₂] in refluxing mesitylene and after 4 h, 31P{1H} NMR spectroscopy indeed showed only 3a. In comparison, the radioactive synthon fac- $[^{99m}Tc(CO)_3(OH_2)_3]^+$ is readily prepared in situ from an IsoLink[®] kit.6 Substitution of the aqua ligands under aqueous or mixed polar organic conditions readily occurs, requiring the preparation of rhenium analogues under more polar conditions, to better correlate with the 99mTc experiments. Thus triphos-Ph was reacted with [Re(CO)₅][OTf] in refluxing ethanol to give fac-[Re(CO)₃(triphos-Ph)][OTf], 4; a sample suitable for X-ray crystallographic analysis was obtained by pentane diffusion into a tetrahydrofuran solution (Fig. 2). To synthesise a fluorescent analogue, 2 was reacted with [Re(CO)5][OTf] to give exclusively fac-[Re(CO)₃(2)][OTf], 5, characterised by X-ray diffraction (Fig. 3).

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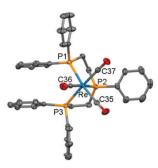


Fig. 2 X-ray crystallographic structure of 4 with 50% probability displacement ellipsoids. Hydrogen atoms and the anion are omitted for clarity. Selected bond distances [Å] and angles [°]: Re-P1 2.4680(12), Re-P2 2.4313(13), Re-P3 2.4666(12), Re-C35 1.957(5), Re-C36 1.971(6), Re-C37 1.954(6); P1-Re-P2 81.88(4), P1-Re-P3 95.12(4), P2-Re-P3 80.92(4), P1-Re-C35 169.49(16), P2-Re-C36 173.86(14), P3-Re-C37 174.70(16),

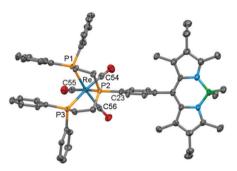


Fig. 3 X-ray structure of 5. Hydrogen atoms, the anion and solvent molecules are omitted for clarity. Selected bond distances [Å] and angles [°]: Re-P1 2.4544(13), Re-P2 2.4230(13), Re-P3 2.4831(13), Re-C54 1.926(6), Re-C55 1.975(6), Re-C56 1.944(6); P1-Re-P2 81.44(5), P1-Re-P3 91.64(4), P2-Re-P3 81.11(4), P1-Re-C56 167.69(18), P2-Re-C55 173.27(17), P3-Re-C54 174.80(17),

The Re-P and Re-C bond lengths and angles are typical for such complexes. 4,5 The photophysical properties of 2, 3a-c and 5 were then measured (Table 1). The absorption spectra of 2 and the complexes showed a strong S_0 – S_1 (π – π *) transition with a maximum of 512 or 513 nm, assigned to the Bodipy core.⁷

Compound 2 has a typically high molar absorption coefficient of 90 000 M⁻¹ cm⁻¹ which is lowered for the associated metal complexes (60 000–64 000 M⁻¹ cm⁻¹). A lower-intensity, broader absorption band between 370 and 380 nm ($\varepsilon = 2500-4200 \text{ M}^{-1} \text{ cm}^{-1}$) is

Table 1 Photophysical data for phosphine 2 and its Re complexes

	λ _{abs} ^a / nm	$\varepsilon^a/M^{-1} \text{ cm}^{-1}$	λ _{em} ^a / nm	$\Phi_{ ext{F}}{}^{a,b}$	λ _{em} ^c / nm	$\Phi_{ ext{F}}^{\ \ b,c}$	λ _{em} ^d / nm	$\Phi_{ ext{F}}^{\;\;b,d}$
2	513	90 000	527	0.34	525	0.40	523	0.39
3a	513	64000	527	0.28	529	0.26	526	0.27
3b	513	63 000	527	0.26	528	0.26	525	0.27
3 c	512	e	526	0.18	527	0.24	524	0.25
5	513	60 000	528	0.24	531	0.18	527	0.20

^a In degassed tetrahydrofuran at room temperature. ^b Measured with respect to 4,4-difluoro-8-phenyl-1,3,5,7-tetramethyl-2,6-diethyl-4-bora-3a,4a-diaza-s-indacene; dyes were excited at 485 nm. ^c In degassed dichloromethane at room temperature. ^d In degassed methanol at room temperature. ^e Insufficient quantity of sample isolated for measurement.

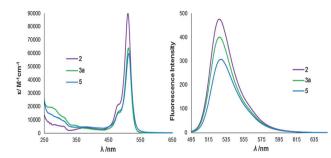


Fig. 4 Absorption and emission spectra of 2 and its metal complexes cis,mer-[ReCl(CO)₂(2)][OTf] 3a and fac-[Re(CO)₃(2)][OTf] 5 in THF. For 3b and 3c see ESI t

attributed to the S_0 – S_2 (π – π *) transition of the Bodipy core;⁷ the absorption profiles are displayed in Fig. 4. Phosphine 2 and its complexes all exhibit emission at room temperature in tetrahydrofuran, dichloromethane and methanol, on excitation at 485 nm. The emission maximum (λ_{em}) is seen at 527 nm in tetrahydrofuran for 2, and is shifted to lower wavelengths in dichloromethane and methanol. Upon complexation, no or very little change is observed in the emission maxima. For all the complexes, when the solvent is changed from dichloromethane to methanol, the emission maxima are slightly blue-shifted. The Stokes shift for the complexes are small (14-15 nm in THF), suggesting negligible structural change on excitation. The fluorescence quantum yield $(\Phi_{\rm F})$ for 2 is 0.34 in tetrahydrofuran, which is comparable to the parent primary phosphine 1 ($\Phi_{\rm F} = 0.33$).³ This is important, as it shows that the two additional phosphorus groups in this tridentate derivative do not impact negatively on the fluorescence, and indicates reductive-PeT is not occurring.

On coordination, the fluorescence quantum yields are slightly lowered for all the complexes (Table 1). One explanation for this may be the heavy atom effect, causing spin-orbit coupling and giving rise to intersystem crossing to the triplet state.8

The $\Phi_{\rm F}$ values are high in comparison to tridentate quinolinederived nitrogen-based rhenium complexes, which have $\Phi_{\rm F}$ of 0.003-0.015, and therefore these phosphorus-based probes may be even more sensitive in vitro cell imaging agents.²

The corresponding technetium complexes were first investigated using triphos-Ph as a mimic for 2, to establish the general labelling conditions. Reacting fac-[99m Tc(CO)₃(OH₂)₃] $^+$ with 1 \times 10⁻⁵ M triphos-Ph in a pH 7.2 sodium phosphate buffer-ethanol solution at 85 °C for 1 h resulted in quantitative radiochemical conversion to 6. A single HPLC peak for 6 closely matched that of fac-[Re(CO)₃(triphos-Ph)]⁺ 4 (Scheme 2 and Fig. 5). Under the same

$$\begin{array}{c} Ph_2P \\ PPh_2 \\ \hline PPh_3 \\ \hline P$$

Scheme 2 Synthesis of fac-[99mTc(CO)3(triphos-Ph)]+ 6 and fac- $[^{99m}Tc(CO)_3(2)]^+$ 7.

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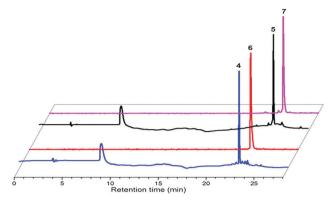


Fig. 5 Overlay of HPLC chromatograms from the crude reactions between triphos-Ph or **2** and $[Re(CO)_5]^+$ (UV, 220 nm) or $fac_1^{99m}Tc(CO)_5(OH_2)_5]^+$ (Nal, γ -detector) to yield fac-[Re(CO)₃(triphos-Ph)]⁺ (4) at 22.9 min, fac- $[^{99m}Tc(CO)_3(triphos-Ph)]^+$ (6) at 23.0 min, $fac-[Re(CO)_3(2)]^+$ (5) at 24.4 min and $fac-[^{99m}Tc(CO)_3(2)]^+$ (7) at 24.5 min.

conditions, the Bodipy phosphine, 2, produced fac-[99mTc(CO)₃(2)]⁺ 7 in similar conversion, giving a single major HPLC peak that correlated with the rhenium analogue 5 (Fig. 5). The stabilities of 6 and 7 were examined by radio-HPLC using competitive amino acid challenge assays (1 mM histidine or cysteine) to simulate an in vivo environment ([PO₄]³⁻ 10 mM, pH 7.2, 37 °C). Analysis by HPLC indicated that both complexes remained >97% stable up to 18 h; no *trans*-chelation of the *fac*-[^{99m}Tc(CO)₃]⁺ core in **6** or **7** with either amino acid was observed during the study. The radiolabelling vields, purity and stability of 6 and 7 indicate that the tridentate phosphine ligand system is comparable for fac-[99mTc(CO)₃]⁺ to other potent tridentate chelates (e.g. histidine, bis(2-pyridylmethyl)amine [DPA]). Therefore, to the best of our knowledge, complex 7 represents the first example of a phosphine-based, multi-functional imaging tool, combining (i) a tridentate phosphine for kinetic stability, (ii) a fluorophore for in vitro imaging, and (iii) a radioactive metal for *in vivo* imaging *via* γ-detection by SPECT.

Radiolabelling of the complexes constitutes the first, crucial step towards their use in nuclear medicine for in vivo tissue imaging, but SPECT does not provide information at the subcellular level due to resolution limitations (1-2 mm). In contrast, optical imaging methods allow for the direct visualisation of the uptake and localisation of complexes within cells which often contributes to the understanding of the mechanism of action of such probes in cellular environments, due to the sub-micron resolution level. 10 Thus, in a preliminary screening, the rhenium complexes 3a and 5 were imaged in prostate carcinoma (PC-3) cells (cultured as described in the ESI†), by epi-fluorescence microscopy using single-photon excitation between 460 and 500 nm and an emission filtered at 510 nm (Fig. 6). Remarkably, exchanging a chloride for a carbonyl ligand renders the cellular behaviour of 3a and 5 very different; whereas [ReCl(CO)₂(2)]⁺ allows for high-resolution imaging and enables visualisation of organelles without any apparent cytotoxicity, fac-[Re(CO)₃(2)]⁺, 5, on the other hand causes some morphological changes. Further investigations would elucidate the sub-cellular localisation of the rhenium complexes into specific organelles. MTT assays were carried out

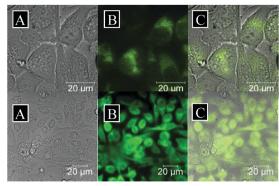


Fig. 6 Epi-fluorescence imaging of PC-3 living cells with compounds 3a and 5. Top: cis,mer-[ReCl(CO)₂(2)], 3a, 100 μM, 1% DMSO, 15 minutes. Bottom: fac-[Re(CO)₃(**2**)]⁺, **5**, 50 μ M, 2% ethanol, 15 minutes. (A) Brightfield image, (B) green channel $\lambda_{\rm ex}$ = 460–500 nm, long pass filtered at 510 nm, (C) overlay of A and B.

in PC-3 cells, which indicated an MI₅₀ (the concentration required to reduce mitochondrial metabolism to 50%) of 45 μ M \pm 5 μ M for complex 5, confirming its cytotoxic effect (ESI†). Furthermore, MTT assays indicated that complex 3a was innocuous up to 250 μM after 48 h incubation (ESI†). Moreover, both 3a and 5 possess negligible cytotoxicity at the concentrations required for detection via SPECT, confirming that they are highly appropriate for use as imaging probes, 11 and our work is now focused in this direction.

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