Different formation kinetics and photoisomerization behavior of self-assembled monolayers of thiols and dithiolanes bearing azobenzene moieties

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Self-assembled monolayers (SAMs) containing azobenzene moieties are very attractive for a wide range of applications, including molecular electronics and photonics, bio-interface engineering and sensing. However, very little is known about the aggregation and photo-switching behavior that azobenzene units undergo during the SAM formation process. Here, we demonstrate that the formation of thiol-based SAMs containing azobenzenes (denoted as AzoSH) on gold surfaces is characterised by a two-step adsorption kinetics, while a three-step assembly process has been identified for dithiolane-based SAMs containing azobenzenes (denoted AzoSS). The H-aggregation on the AzoSS SAMs was found to be remarkably dependent on the time of self-assembly, with less aggregation as a function of time.

While photoisomerization of the AzoSH was suppressed for all different assembly times, the reversible trans–cis photoisomerization of AzoSS SAMs formed over 24 hours was clearly observed upon alternating UV and Vis light irradiation. We contend that detailed information on formation kinetics and related optical properties is of crucial importance for elucidating the photo-switching capabilities of azobenzene-based SAMs.

Introduction

Photo-switchable self-assembled monolayers (SAMs) have been receiving considerable attention, motivated by their potential applications in molecular electronics and photonics, bio-interface engineering, catalysis and sensing. One of the most frequently studied classes of photo-switchable molecules is the azobenzene-containing molecules, since the azobenzene moiety is known to undergo cis–trans isomerism. Photoisomerization from the thermodynamically more stable trans isomer to the cis isomer and vice versa can be selectively induced by irradiation with either ultraviolet (UV) light or visible (Vis) light. The free volume required for the cis form is larger than for the trans form assuming either an inversion or a rotation mechanism of the azo bond. Thus, photoisomerization rates strongly depend on the local free volume available for the conformation change. In this regard, an azobenzene-terminated alkanethiol SAM hardly exhibits any photoisomerization owing to the existence of both H-aggregation and spatial constraints. Regarding the former, neighboring azobenzenes have a high tendency to form head-to-head H-aggregates in a SAM due to π–π stacking interactions, leading to a dense molecular packing, that suppresses azobenzene photoisomerization.

Efforts have been made to increase the spacing between the azobenzene moieties in order to promote the reversible trans–to-cis photoisomerization process in SAMs on gold. Photo-switching of azobenzene SAMs has been achieved by employing asymmetrical disulfides, asymmetrical thioethers, bulky alkyl groups into the benzene ring of the azobenzene unit, or by inclusion of a bulky carborane unit into the diazo functional group in order to space out the backbones. Other strategies relied on azobenzene derivatives with bulkier terminal headgroups than thiols, including a tripod adamantane-based thioacetate head-group, an asparagusic acid-based 1,2-dithiolane headgroup and an α-lipoic acid-based 1,2-dithiolane headgroup. Regarding the latter, no photoisomerization was observed when the molecules were absorbed from the trans isomer, whereas molecules adsorbed in cis configuration were photoreactive. The inhibition of structural arrangement associated with trans–cis isomerization when the molecules were absorbed from the trans isomer is most

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probably due to hydrogen bonds between the amide groups by which the azobenzene moieties were attached to the dithiolane headgroup moiety.\textsuperscript{12} Although SAMs of an azobenzene terminated dithiolane analogue, in which the amide group in the spacer was replaced by an ester moiety, were prepared by the same authors,\textsuperscript{21} no photoisomerization studies were conducted. Thus, the photoisomerization capabilities of \(\alpha\)-lipoic acid-based SAMs containing azobenzene moieties are not fully understood and remain to be further investigated. Furthermore, in this study and others,\textsuperscript{17–21} little attention has been paid to the aggregation and photoswitching behavior that azobenzene units undergo during the SAM formation process. Single monolayer formation times have been used for accessing the switching of SAMs containing azobenzene moieties and times of formation have been varied from 0.5 h to 24 h.\textsuperscript{17,18,20,21}

Stimulated by the lack of information in certain aspects of azobenzene-based SAMs as discussed above, in this study we investigate in detail monolayers formed from an \(\alpha\)-lipoic acid-based azobenzene (\textit{AzoSS}, Fig. 1), in which an ester linkage is incorporated into the chain between the \(\alpha\)-lipoic acid and the azobenzene, and a thiol-based azobenzene analogue (\textit{AzoSH}, Fig. 1), in which the \(\alpha\)-lipoic acid is replaced by an alkanethiol which is linked to the azobenzene using an ether bond. The goal of this work is to extend our knowledge on azobenzene-based SAMs, to answer two principal questions. First, how does H-aggregation and the optically induced switching depend on the time of SAM formation, and are these properties dependent on the headgroup? Second, are \(\alpha\)-lipoic acid-based azobenzene SAMs capable of undergoing reversible photoisomerization in the absence of hydrogen bonding on the SAM structure? The adsorption kinetics of the \textit{AzoSS} and \textit{AzoSH} SAMs are studied by means of contact angle, ellipsometry, surface plasmon resonance (SPR) spectroscopy and X-ray photoelectron spectroscopy (XPS). The relation between time of SAM formation and its effect on H-aggregation and photoswitching capabilities for both SAMs, \textit{AzoSH} and \textit{AzoSS} SAMs, is also ascertained by UV/Vis spectroscopy using the spectroscopic characteristics of the azo chromophores in the \textit{cis} and \textit{trans} conformations.

### Results and discussion

Two new azobenzene derivatives, \textit{AzoSS} and \textit{AzoSH} were synthesized as outlined in Fig. 1. Synthesis of \textit{AzoSS} was initiated through diazotisation of the 4-aminobenzoic acid tert-butyl ester, which was reacted with phenol under basic conditions to give the azobenzene 1. The azobenzene 1 was coupled with thioctic acid using \(N,N'\text{-dicyclohexylcarbodiimide (DCC)}\) in the presence of 4-dimethylaminopyridine (DMAP) to afford the \textit{AzoSS}. The synthesis of \textit{AzoSH} involved initial formation of the thioacetate 2 via thioacetylation of 6-bromo-1-hexene in the presence of CH\(_3\)COSH and the radical initiator, 2,2\textsuperscript{-}azobisisobutyronitrile (AIBN), followed by alkylation with the phenoxide moiety in 1, to form the azobenzene thioacetate 3. Subsequently, the thioacetate 3 was hydrolysed under acidic conditions to obtain the desired \textit{AzoSH}. These hydrolysis conditions did not hydrolyse the tert-butyl ester group.

Fig. 2 shows the UV absorption spectra of \textit{AzoSS} and \textit{AzoSH} in ethanol (0.0625 mM) before and after irradiation with UV (365 nm) and subsequent visible light irradiation (436 nm). Typical of azobenzene derivatives, the \textit{trans} form is more stable and is the dominant isomer before UV irradiation. The \textit{trans} form of \textit{AzoSS} exhibits a strong \(\pi–\pi^*\) absorption peak with a maximum wavelength (\(\bar{\lambda}_{\text{max}}\)) at 329 nm and a weak \(n–\pi^*\) band at around 445 nm. The corresponding bands for the \textit{trans} form of \textit{AzoSH} are found at 360 nm and 445 nm. Thus, the \(\bar{\lambda}_{\text{max}}\) of the \(\pi–\pi^*\) absorption band is influenced by the linking functionality (\textit{i.e.} ether or ester group) between the azobenzene and the

![Fig. 1 Synthesis of AzoSS and AzoSH: (i) NaN\(_2\), 1.1 M HCl, MeOH, 0 °C; (ii) Phenol, KOH\(_{\text{aq}}\), MeOH, rt, 4 h; (iii) CH\(_3\)COSH, AIBN, PhMe, reflux, 2 h; (iv) DCC, DMAP (cat), DCM, rt, N\(_{2}\) atm, 16 h; (v) K\(_2\)CO\(_3\), acetone, refluxed, 16 h; (vi) 0.1 M HCl, MeOH, reflux, N\(_{2}\) atm, 4 h.](image-url)
headgroup, with $\lambda_{\text{max}}$ of the AzoSH being bathochromically shifted by 25 nm with respect to that of AzoSS. For both compounds, AzoSS and AzoSH, upon UV irradiation for 1 min, a drastic reduction in the $\pi-\pi^*$ band is observed, whereas the $\pi-\pi^*$ becomes more intense. These results suggest that the azobenzene molecules are converted into cis isomers in both solutions. Subsequent visible light irradiation for 1 min gives rise to cis to trans isomerization, maximizing the $\pi-\pi^*$ absorption band. Although the cis to trans isomerization is fully reversible for the AzoSS molecule, the AzoSH molecule displays slightly less reversible behaviour. This photoequilibrium composition remained unchanged even after the irradiation was prolonged to 5 min.

SAM formation was evaluated using ellipsometry, contact angle and XPS (Fig. 3–5) after 24 h immersion time of a cleaned gold substrate in a 0.0625 mM ethanolic solution of AzoSS or AzoSH. The ellipsometric thicknesses of the fully formed SAMs are 1.8 nm (AzoSS) and 1.6 nm (AzoSH), and both are less than the theoretical molecular length of the molecules (both 2.5 nm). This discrepancy, between molecular length and SAM thickness, is expected and is in agreement with the literature, being ascribed to both the tilt angle and density of the SAM surfactants.22,23 The advancing ($\theta_{\text{adv}}$) and receding ($\theta_{\text{rec}}$) contact angles for AzoSS ($\approx 90^\circ$ and $\approx 75^\circ$, respectively) and AzoSH ($\approx 90^\circ$ and $\approx 80^\circ$, respectively) are in good agreement with the literature24,25 for tert-butyl ester SAMs, noting that the final hysteresis values ($\theta_{\text{adv}}$–$\theta_{\text{rec}}$) of AzoSS ($\sim 15^\circ$) are slightly larger than AzoSH ($10^\circ$) indicating a less dense packed SAM for AzoSS. XPS analysis confirms the presence of the elemental species N, C, O and S on the AzoSS SAMs (Fig. 5).

The single peak at 400.5 eV in the N (1s) spectrum is assignable to the nitrogen of the azo group.16 The C (1s) spectrum can be deconvoluted into three peaks, which are attributed to five different binding environments (Fig. 5). The main, predominant peak (284.7 eV) is attributed to C–C bonds26 within both the alkyl chains and phenyl rings. The first of the two smaller peaks (286.4 eV) is attributed to C (1s) of the three binding environments of C–S, C–N and C–O–C.26 The third and final peak (288.7 eV) is attributed to the C (1s) photoelectron of the carbonyl moiety, C═O.26 The O (1s) spectra are de-convoluted into two different peaks, corresponding to two different binding environments, arising from the ester moieties, C–O–C (533.9 eV) and the carbonyl oxygen C═O (532.3 eV).26 The S (2p) spectrum displays a doublet structure at 163.6 eV (S (2p$_{3/2}$)) and 162.4 eV (S (2p$_{1/2}$)), which is assignable to the thiolate-type sulfur bound to the gold surface as previously reported for SAMs using a 1,2-dithiolane headgroup.21 The AzoSH SAMs exhibit similar N (1s) and S (2p) spectra (Fig. 5). The C (1s) and O (1s) are also similar, but with lower intensities for the C (1s) peak at 288.7 eV and O (1s) at 532.3 eV due to the absence of the second carbonyl moiety on the AzoSH molecule.

The kinetics of formation of the AzoSS and AzoSH SAMs was investigated by ellipsometry and contact angle. SAMs were formed by immersion of freshly cleaned Au substrates in 0.0625 mM ethanolic solutions of either AzoSS or AzoSH for 0.5 h, 1.5 h, 3 h, 6 h, 9 h, 18 h, 21 h, 24 h and 48 h, followed by rinsing with ethanol. Ellipsometric thickness and contact angles ($\theta_{\text{adv}}$ and $\theta_{\text{rec}}$) were recorded for each time interval (Fig. 3 and 4). In the case of AzoSS monolayers, three distinct SAM formation stages can be discerned in both Fig. 3 and 4 when considering the ellipsometric thickness and the $\theta_{\text{rec}}$ angle, which is significantly different to a simple alkyl thiol and indeed the AzoSH growth kinetics (see later). These stages occur between 0 and 6 h (Stage I), 6 and 21 h (Stage II) and after 21 h (Stage III). Considering the very initial stage (Stage I), the thickness rises to $\approx 2.4$ nm within 0.5 h and subsequently drops to 1.2 nm over the first 6 h. Over the next 15 h, the AzoSS SAM undergoes a growth in thickness, as might be expected for SAM formation (Stage II). Finally, after 21 h the thickness plateaus at $\approx 1.8$ nm, indicating that the SAM is fully formed (Stage III).

In contrast, for AzoSH there is no Stage I as for AzoSS, and the SAM thickness rises to $\approx 0.80$ nm within 0.5 h and then to $\approx 1.2$ nm by 6 h and reaches a plateau of $\approx 1.6$ nm by 24 h,
following standard alkyl thiol growth kinetics. This two-stage adsorption behavior of AzoSH is also in agreement with adsorption kinetic studies conducted by Tamada et al.\textsuperscript{11} on azobenzene-containing alkanethiol SAMs on gold. As a control, and to rule out the initial increase in thickness of the AzoSS surface we considered that a bare Au surface is a relatively high energy surface and is prone to the physisorption of airborne contaminants.\textsuperscript{27} Thus, we carried experiments without the AzoSS in the solution and measured the ellipsometric thickness after several time intervals over 6 h, which showed the ellipsometric thickness of the contaminant layer to be $\approx 1.24$ nm, irrespective of immersion time. This thickness is lower than the values observed (2.4 nm) within the first 4 h of SAM formation with AzoSS, and hence Stage I in Fig. 3 cannot be attributed to the non-specific physisorption of airborne contaminants.

The $\theta_{\text{rec}}$ angles of the AzoSS reveal similar three stage behavior to the ellipsometric data. The $\theta_{\text{rec}}$ reaches $\approx 65^\circ$ within 0.5 h, and then decreases to $\approx 35^\circ$ over 1.5 h (Stage I), then increases to $\approx 70^\circ$ over the following 4.5 h (Stage II), and remains constant for the next 12 h (Stage III), suggesting the presence of a more sparsely packed monolayer at 24 h, as indicated by a large hysteresis of $\approx 15^\circ (\theta_{\text{adv}}-\theta_{\text{rec}})$ relative to AzoSH. In contrast the AzoSH $\theta_{\text{rec}}$ angle increases consistently over the SAM formation time, starting at $\approx 75^\circ$ and reaching a plateau after 21 h at $\approx 80^\circ$, giving a final smaller hysteresis of $10^\circ (\theta_{\text{adv}}-\theta_{\text{rec}})$, indicating a more ordered and packed monolayer structure than the AzoSS.

Label-free sensing technologies such as SPR and quartz crystal microbalance (QCM) allow for real-time monitoring of adsorption processes and kinetics.\textsuperscript{28–30} Thus, in addition to the ellipsometry and contact angle measurements, we have carried out SPR analysis in order to monitor the adsorption kinetics of the AzoSS and AzoSH onto the gold surfaces (Fig. 6). The SPR baseline for the clean gold chips was established using ethanol, following which the AzoSS or AzoSH in ethanol (0.0625 mM) was introduced into the SPR flow cell at the rate of 10 $\mu$L min$^{-1}$ (Fig. 6). Data were collected for 21 h, followed by washing with ethanol. SPR reveals clear differences in the kinetics of monolayer formation for AzoSS and AzoSH SAMs. The AzoSS SAM formation proceeds through three stages. In Stage I (0–2.3 h), an initial adsorption of molecules on the surface that peaks at 1950 response units is followed by a 15% decrease in the SPR signal after 30 minutes. This decrease suggests a significant level of desorption of molecules from the gold surface. After Stage I, the formation kinetics proceeds into two more stages – Stage II and Stage III. Stage II occurs between 2.3 h and 6.6 hours and Stage III from 6.6 hours to the time the rinsing is initiated. Both stages are characterised by an initial increase
in the amount of AzoSS on the surface, evidenced by the increase in the SPR response units, that plateaus for Stage II and Stage III after 1.5 h and 3 h, respectively. When ethanol replaces the AzoSS solution, there is a significant drop in the SPR signal to 1980 response units, indicating the removal of a significant amount of non-chemisorbed AzoSS molecules from the surface.

The AzoSH SAM formation proceeds through a two-stage process, involving a rapid initial adsorption of AzoSH on the surface followed by a much slower adsorption that plateaus at ~3330 response units. After contact with the AzoSH thiol solution is finished, the signal decreases slightly with only a small amount of AzoSH being removed from the surface.

These distinct stages between the kinetics of SAM formation of the AzoSS and AzoSH, as shown in the ellipsometric (Fig. 3), contact angle (Fig. 4) and SPR (Fig. 6) analysis, suggest that the interplay between the dithiolane and the azobenzene moiety interacting with the Au substrate plays an important role in defining the SAM adsorption process for AzoSS, and a different interplay to the thiol and azobenzene in AzoSH, which follows classical alkyl thiol adsorption kinetics.27

Thus, it is clear that the adsorption kinetics of the AzoSS and AzoSH SAM formation are significantly different, which may be accounted for by the competing interactions of the azobenzene moiety and either the thiol or the dithiolane for the gold surface. If we consider that the RS–Au bond forms more rapidly from a thiol than it does from a disulfide or dithiolane as previously suggested by Whitesides and co-workers,31 the initial physisorption of the azobenzenes in a lying-down configuration on the gold surface may persist for longer with the AzoSS than the AzoSH. This behaviour allows a multilayer of AzoSS molecules to form by virtue of van der Waals and π–π interactions between surface physisorbed AzoSS and those approaching the interface. Indeed, the high thickness of AzoSS SAMs at lower formation times (Stage I) supports such a hypothesis when coupled to the rapid decline in thickness, such that at low formation times for AzoSS (Stage I) a transient multilayer adsorption process occurs with the azobenzenes lying-down on the gold surface before the S–S bond can cleave and form the Au–S bond. This explanation is consistent with the literature32–34 on the growth of SAMs in which the SAM formation phases (initial physisorbed, lying-down phase, followed by a chemisorbed, standing-up phase) are determined, among several other parameters, by the structure of the SAM molecule, i.e. headgroup, backbone and endgroup.

A striking difference between the ellipsometry and SPR results is that in the former characterisation, the initial adsorption of AzoSS molecules on the gold surface is much more
pronounced and the desorption occurs more slowly. Ellipsometry results demonstrate that the desorption occurs for a period of 5.5 h while SPR desorption takes place for 30 min. The differences obtained by ellipsometry and SPR are believed to be related to the flow-induced shear stress applied during the SPR analysis which is not present during the preparation of the SAM surfaces used for ellipsometry measurements. These findings suggest that flow-induced shear stress may prevent to a certain extent lateral stacking of the AzoSS molecules on the surface. Despite the use of different SAM preparation conditions

![Fig. 5](image_url) N (1s), C (1s), O (1s) and S (2p) XPS spectra of AzoSS and AzoSH SAMs, together with the corresponding fits.
for both analyses, both ellipsometry and SPR clearly illustrate the differences in kinetics of AzoSS and AzoSH SAM formation.

The UV/Vis spectra of both AzoSS and AzoSH SAMs (Fig. 7) were investigated as a function of time. Post SAM formation samples were washed and dried, then immediately analysed using UV/Vis absorption spectroscopy. The UV/Vis spectra of AzoSS SAMs show the three formation stages (I–III), in accordance with those seen by ellipsometry and contact angle. At low formation times (0.5–6 h, Stage I), $\lambda_{\text{max}}$ (Table 1) is hypsochromically shifted with respect to $\lambda_{\text{max}} = 329$ nm in an ethanolic solution, which can be interpreted in terms of strong intermolecular interactions in the film leading to formation of H-aggregates. In Stage II (6–12 h of SAM formation), $\lambda_{\text{max}}$ is still hypsochromically shifted with respect to the solution value (329 nm), but the hypsochromic shifts are generally not as large as in Stage I, suggesting that the degree of aggregation is reduced with increasing formation time. In Stage III (24 h of SAM formation), the spectrum revealed an intense $\pi-\pi^*$ absorption band at $\approx 329$ nm, which is in agreement with the solution value, therefore indicating that azobenzene is not aggregated.

AzoSH UV/Vis spectra are characterized by two absorption bands (Fig. 7), with the absorption at lower wavelengths...
found to follow a three-stage assembly process, with ellipsometry, contact angle, SPR and UV/Vis spectroscopy data suggesting that the \textit{AzoSS} moieties initially adsorb on the surface as multilayer aggregates. The results highlight the critical role of time of SAM formation in dictating H-aggregation and photoisomerization on \textit{AzoSS} SAMs. \textit{AzoSS} SAMs are aggregated at low formation times and the degree of aggregation decreases with increasing formation time, until non-aggregated SAMs are formed after 24 h. Azobenzene, within \textit{AzoSH} SAMs are generally H-aggregated, irrespective of formation time. While SAMs of \textit{AzoSH} do not exhibit photoisomerization, clear and highly reversible photo-induced effects are observed for \textit{AzoSS} SAMs.

### Experimental

**Chemicals and materials**

Commercially available chemicals and solvents were purchased from Aldrich Chemicals and Fisher Chemicals and were used as received. Thin-layer chromatography (TLC) was carried out on aluminium plates coated with silica gel 60 F254 (Merck 5554). The TLC plates were either air-dried and analysed under a short wave UV lamp (254 nm) or developed in either permanganate or sodium hydroxide solution and heated-dried. Column chromatographic separations were performed using silica gel 120 (ICN Chrom 32–63, 60 Å).

**Synthesis of AzoSH and AzoSS**

\[(E)-\text{tert-Butyl} + 4(4\text{-hydroxyphenyl})\text{diazenyl}\text{benzoate} \] (1). A solution of NaNO\textsubscript{2} (1.22 g, 17.68 mmol) in H\textsubscript{2}O (3.4 mL) was added dropwise to a solution of 4-aminobenzoic acid (3.40 g, 17.62 mmol) in 1 M HCl (10 mL) at 0 °C. The subsequent diazotised solution was added dropwise to a chilled solution of phenol (1.66 g, 17.66 mmol) and KOH (1.97 g, 35.18 mmol) in MeOH (21 mL). The resultant reaction mixture was further stirred for 4 h. The reddish/orange precipitate was collected via suction filtration to yield a reddish/orange solid (4.22 g, 85%). \textsuperscript{1}H NMR (300 MHz, CDCl\textsubscript{3}, Me\textsubscript{4}Si, 25 °C) \(\delta\) ppm 8.11 (d, 2H, \(J = 8.45\) Hz), 7.90 (d, 2H, \(J = 8.75\) Hz), 7.88 (d, 2H, \(J = 8.45\) Hz), 6.97 (d, 2H, \(J = 8.75\) Hz, ArH), 1.64 (s, 9H); \textsuperscript{13}C NMR (75 MHz, CDCl\textsubscript{3}, Me\textsubscript{4}Si, 25 °C) \(\delta\) ppm 164.2, 157.1, 151.8, 144.0, 128.9, 124.0, 120.7, 114.5, 80.4, 26.7; m/z (ESMS): 321 [(M + Na\textsuperscript{+}), 100%]; m/z (HRMS): found 321.1217. Calc. mass for \(C_{17}H_{14}NaO_3\)Na: 321.1215.

\textit{S-(6-Bromohexyl) ethanethioate} (2). A solution of 6-bromo-1-hexene (2.00 g, 12.27 mmol), thioacetic acid (1.87 g, 24.61 mmol) and AIBN (catalytic amount) in PhMe (20 mL) was refluxed for 2 h. The resultant reaction mixture was allowed to cool to room temperature and saturated NaHCO\textsubscript{3} aqueous solution was added and the organic layer was extracted with DCM (3 × 20 mL). The combined organic layers were dried (MgSO\textsubscript{4}), filtered and concentrated \textit{in vacuo}. The crude product was purified by column chromatography (eluent: hexane) to yield a colourless oil (2.00 g, 68%). \textsuperscript{1}H NMR (300 MHz, CDCl\textsubscript{3}, Me\textsubscript{4}Si, 25 °C) \(\delta\) ppm 3.34 (t, 2H, \(J = 6.14\) Hz), 2.26 (s, 3H), 1.83–1.75 (m, 2H), 1.58–1.48 (m, 2H), 1.44–1.29 (m, 4H); \textsuperscript{13}C NMR (75 MHz, CDCl\textsubscript{3}, Me\textsubscript{4}Si, 25 °C) \(\delta\) ppm 195.9, 133.7, 132.6, 130.6, 29.3, 28.2, 28.9, 27.9, 27.6; m/z

### Conclusions

We have compared the formation kinetics and photoisomerization properties of azobenzene SAMs comprising either a thiol or a dithiolane headgroup. The azobenzene dithiolane SAMs were
(ESMS): 261 ([M + Na]+, 100%), 263 ([M + Na]+, 100%); m/z (HRMS): found 260.9923. Calc. mass for C$_{24}$H$_{28}$N$_2$O$_4$SNa: 260.9925.

(E)-t-Butyl 4-(4-{(1,2-dithiolan-3-yl)pentanoyl}oxy)phenyl)diazeylnbenzoate (AzOSS). To a stirred solution of thiotic acid (2.00 g, 9.60 mmol) and DMAP (0.06 g, 0.48 mM) in anhydrous DCM, at room temperature, 1 (0.72 g, 2.40 mmol) was added. The reaction mixture was cooled to 0 °C in an ice bath and DCC (1.00 g, 4.80 mM) added and stirred for 10 min. The ice bath was then removed and the reaction mixture brought to room temperature and stirred overnight. The crude reaction mixture was washed with HCl (1 M) (1 × 100 mL) followed by 5% NaHCO$_3$ (1 × 100 mL) and finally with water (1 × 100 mL). The organic layer was then dried (MgSO$_4$) filtered and concentrated in vacuo. The crude product was purified by column chromatography (1% MeOH in DCM) to yield an orange solid (0.70 g, 60%). Elemental analysis found: C, 61.88%; H, 7.50%; N, 6.77%. Calc. for C$_{23}$H$_{30}$N$_2$O$_3$S: C, 61.7%; H, 6.2%; N, 5.8%. 1H NMR (300 MHz, CDCl$_3$, Me$_4$Si, 25 °C): $\delta$H ppm 8.41 (d, $J = 8.60$ Hz, 2H), 8.27 (d, $J = 8.60$ Hz, 2H), 8.19 (d, $J = 8.60$ Hz, 2H), 7.54 (d, $J = 8.60$ Hz, 2H), 3.89 (m, 1H), 3.54 (m, 2H), 2.90 (t, 2H), 2.05 (m, 4H), 1.90 (s, 9H); NMR (75 MHz, CDCl$_3$, Me$_4$Si, 25 °C): $\delta$C ppm 132.3, 126.2, 124.4, 124.2, 58.1, 42.1, 40.4, 36.5, 36.0, 26.4; m/z (ESMS): 509.2 ([M + Na]+, 100%).

(E)-t-Butyl 4-{(4-{(6-acetylthio)hexyl}oxy)phenyl)diazeylnbenzoate (3). A slurry of 1 (1.00 g, 3.34 mmol), 2 (1.20 g, 5.02 mmol), and K$_2$CO$_3$ (0.92 g, 6.67 mmol) in acetone (100 mL) was refluxed for 4 h and finally with water (1.00 mL) and DMAP (0.06 g, 0.48 mM) in anhydrous DCM, at room temperature, 1 (0.72 g, 2.40 mmol) was added. The reaction mixture was cooled to 0 °C and then HCl (1 M) (1 × 100 mL) followed by 5% NaHCO$_3$ (1 × 100 mL) and then HPLC grade EtOH thoroughly for 100 mL. The organic layer was then dried and the reaction mixture brought to room temperature and K$_2$CO$_3$ was added. The ice bath was then removed and the reaction mixture brought to room temperature and stirred overnight. The crude reaction mixture was washed with HCl (1 M) (1 × 100 mL) followed by 5% NaHCO$_3$ (1 × 100 mL) and finally with water (1 × 100 mL). The resultant reaction mixture was cooled to 0 °C in an ice bath and DCC (1.00 g, 4.80 mM) added and stirred for 10 min. The ice bath was then removed and the reaction mixture brought to room temperature and stirred overnight. The crude reaction mixture was washed with HCl (1 M) (1 × 100 mL) followed by 5% NaHCO$_3$ (1 × 100 mL) and finally with water (1 × 100 mL). The resultant reaction mixture was cooled to 0 °C and then HCl (1 M) (1 × 100 mL) followed by 5% NaHCO$_3$ (1 × 100 mL) and finally with water (1 × 100 mL). The organic layer was then dried (MgSO$_4$) filtered and concentrated in vacuo. The crude product was purified by column chromatography (1% MeOH in DCM) to yield a orange solid (0.70 g, 60%). Elemental analysis found: C, 61.88%; H, 7.50%; N, 6.77%. Calc. for C$_{23}$H$_{30}$N$_2$O$_3$S: C, 61.7%; H, 6.2%; N, 5.8%. 1H NMR (300 MHz, CDCl$_3$, Me$_4$Si, 25 °C): $\delta$H ppm 8.41 (d, $J = 8.60$ Hz, 2H), 8.27 (d, $J = 8.60$ Hz, 2H), 8.19 (d, $J = 8.60$ Hz, 2H), 7.54 (d, $J = 8.60$ Hz, 2H), 3.89 (m, 1H), 3.54 (m, 2H), 2.90 (t, 2H), 2.05 (m, 4H), 1.90 (s, 9H); NMR (75 MHz, CDCl$_3$, Me$_4$Si, 25 °C): $\delta$C ppm 132.3, 126.2, 124.4, 124.2, 58.1, 42.1, 40.4, 36.5, 36.0, 26.4; m/z (ESMS): 509.2 ([M + Na]+, 100%).

(E)-t-Butyl 4-{(4-{(6-mercaptohexyl)oxy)phenyl)diazeylnbenzoate (AzoSS). A solution of 1.55 (s, 9H), 1.55–1.48 (m, 2H), 1.44–1.42 (m, 2H), 1.28 (t, 2H, $J = 7.49$ Hz); $^{13}$C NMR (75 MHz, CDCl$_3$, Me$_4$Si, 25 °C): $\delta$C ppm 165.3, 162.2, 155.1, 146.9, 133.1, 130.4, 125.1, 122.2, 114.8, 81.3, 68.2, 33.9, 29.1, 28.2, 28.1, 25.5, 24.5; m/z (ESMS): 415 ([M + H]+, 100%); m/z (HRMS): found 415.2048. Calc. mass for C$_{24}$H$_{28}$N$_2$O$_4$SNa 415.2055.

Compound characterisation

Nuclear magnetic resonance spectroscopy. 1H Nuclear Magnetic Resonance (NMR) spectra were recorded on a Bruker AC 300 (300.13 MHz) spectrometer. $^{13}$C NMR spectra were recorded on a Bruker AV 300 (75.5 MHz) using the pendent pulse sequences. In both techniques all chemical shifts are calibrated to the SiMe$_3$ peak and quoted in ppm upfield of the reference. Analysis was performed in either deuterated chloroform (CDCl$_3$). The coupling constants of 1H NMR are expressed in Hertz (Hz) with multiplicities abbreviated as follows: s = singlet, d = doublet, t = triplet and m = multiplet.

Mass spectroscopy. Electrospray Mass Spectroscopy (ESMS) and High Resolution Mass Spectroscopy (HRMS) were performed on a Micromass Time of Flight (TOF) using methanol as the running solvent.

Elemental analysis. Elemental analysis was performed on a Carlo Erba EA 1110 (C, H, N) instrument. Reported results are an average of two runs for each compound.

SAM characterisation

Preparation of SAMs. Polycrystalline gold substrates were purchased from George Albert PVD, Germany and consisted either of a 50 nm gold layer (used for ellipsometry, contact angle and XPS analysis) or a 20 nm transparent gold layer (used for UV spectroscopy), deposited onto a glass covered with a thin adhesion layer of titanium. The Au substrates were cleaned by immersion in piranha solution (70% H$_2$SO$_4$; 30% H$_2$O$_2$) at room temperature for 10 min, rinsing with Ultra High Pure (UHP) H$_2$O and then HPLC grade EtOH thoroughly for 1 min. (Caution: piranha solution reacts violently with all organic compounds and should be handled with care.) Immediately after cleaning, the substrates were immersed in freshly prepared ethanolic solutions of AzoSS (0.0625 mM) and AzoSH (0.0625 mM) in the trans form.

Post-immersion in the SAM forming solution, the substrates were rinsed with HPLC grade EtOH and dried under a stream of N$_2$.

Ellipsometry. The ellipsoider used is a Jobin Yvon UVISEL with a broadband xenon light source. The light source's angle of incidence and wavelength range, throughout the experiment, was 70° and 280–820 nm, respectively. Calibration of the ellipsoider and alignment of the polariser and detector was performed through employment of an aluminium reference sample, with a thermally grown aluminium oxide (Al$_2$O$_3$) layer. The ellipsometric parameters, $\Delta$ and $\psi$, were recorded for both the clean bare substrates and for the substrates on which SAMs were formed. All measurements were made under conditions of ambient temperature, pressure and humidity. DeltaPsi software was used to determine the film thickness. Raw data were modelled using a Cauchy transparent model over the whole data range (280–820 nm). Reliability of the model was tested/calibrated using SAMs.
formed from tetra-, hexa- and octa-decanethiols. Measurements reported are an average of at least two independently prepared samples with an average of six readings per sample, taken in different places ensuring not to take measurements on visibly defective sites.

Contact angle. Measurements were preformed with UHQ H2O on a homemade manually operated goniometer. A 25 μL gas tight syringe (Hamilton) was used to add and remove droplets to the surface, droplets were typically of ~15 μL. The drop was recorded using a HitachiTM Charged Coupled Device (CCD) camera, connected to a personal computer by USB in order to capture a video of the advancing and receding angles. Angles were analysed at the three phase boundary using the commercially available video analysis software PTÅ (First Ten Angstroms) version 1.96. Contact angles were determined using the dynamic sessile drop method, using UHQ H2O. The frame rate of video capture was 4 frames per second and 50 frames were collected. Measurements reported are an average of two independently prepared samples with an average of five readings per sample, taken in different places. All measurements were taken under ambient conditions of temperature, pressure and humidity. The errors reported are the standard errors of the mean.

X-ray Photoelectron Spectroscopy (XPS). X-ray Photoelectron Spectroscopy (XPS) spectra were obtained on the VG ESCalab 250 instrument based at the Leeds EPSRC Nanoscience and Nanotechnology Research Equipment Facility (LENNF) at the University of Leeds, UK. XPS experiments were carried out using a monochromatic Al Kα X-ray source (1486.7 eV) and a take off angle of 90°. High-resolution scans of N (1s), S (2p), O (1s), C (1s) and Au (4f) were recorded using a pass energy of 150 eV at a step size of 0.05 eV. Fitting of XPS peaks was performed using the Advantage V2.2 processing software. Sensitivity factors used in this study were: N (1s), 1.73; S (2p), 2.08; C (1s), 1.00; O (1s) 2.8; Au (4f7/2), 9.58; Au (4f5/2), 7.54.

UV/Vis spectroscopy. The UV/Vis absorption spectra of the solution and SAMs were obtained using a double beam Cary 5000 UV/Vis spectrometer. Solution spectra were obtained by measuring the absorption of AzoSS and AzoSH in ethanol (0.0625 mM) in a quartz cell with a path length of 1 cm. Pure ethanol was used as a reference solution. SAM spectra were obtained with a clean gold substrate in the reference beam. Photoirradiation of the samples was carried out with a 200 W Mercury-Xenon Arc Lamp using filters for UV light centred at λ = 365 nm and for visible light centred at λ = 436 nm. SAMs were analysed immediately after formation and subjected to a cycle of irradiation of 3 min exposures to UV or Vis light. Between exposure to light and spectroscopic readings the samples were kept in the dark.

Surface Plasmon Resonance (SPR) spectroscopy. The SPR measurements were performed on a Reichert SR7000DC Dual Channel Spectrometer (Buffalo, NY, USA) at 15 °C. A two-channel flow cell with two independent parallel flow channels was used to carry out the SAM kinetic experiments. A gold-coated SPR chip was placed on the top side of the prism using index-matching oil. After a baseline signal was established by allowing degassed HPLC ethanol to flow at a rate of 10 μL min⁻¹ through the sensor, freshly prepared ethanolic solutions of the AzoSS (0.0625 mM) or AzoSH (0.0625 mM) in the trans form were allowed to flow over the surface for 21 h at a flow rate of 10 μL min⁻¹. In order to remove any unbound AzoSS or AzoSH, the sensor chips were washed with degassed ethanol at a flow rate of 50 μL min⁻¹.

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References


