J- vs. H-type assembly: pentamethine cyanine (Cy5) as a near-IR chiroptical reporter†

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The DNA-enabled dimerization of pentamethine cyanine (Cy5) dyes was studied by optical methods. The value of cyanine as a chiroptical reporter using a monomer-to-dimer switch is demonstrated. The specific shape of the CD signal and its high intensity are a result of J-type assembly.

The use of chiroptical labels is usually not considered as a first line method for biological investigations because of their relatively low sensitivity compared to other optical techniques. On the other hand, circular dichroism (CD) strongly depends on the three-dimensional arrangement of the interacting units and may, therefore, provide information not accessible by other methods. Chiroptical reporters are probes of structural information with special interest in cyanine derivatives as labels and probes, the pentamethine cyanine (Cy5) phosphoramidite (abbreviated herein as Cy) is commercially available. Oligonucleotide–cyanine conjugates reveal a high susceptibility of each monomer, revealing preferential H-type coupling.

We followed our general design of oligonucleotide conjugates with internally positioned artificial building blocks using the phosphoramidite approach (Scheme 1).9–11 Due to the widespread interest in cyanine derivatives as labels and probes, the pentamethine cyanine (Cy5) phosphoramidite (abbreviated herein as Cy) is commercially available. Oligonucleotides ON1, ON2 and unmodified references ON3, ON4 were obtained from custom suppliers (ESI†). The recently reported squaraine (Sq) modified oligonucleotide ON512 served as a reference oligomer and also as a counter strand in mixed hybrid experiments.

An overview of the absorbance and fluorescence data of cyanine modified oligonucleotides is presented in Table 1 and Fig. 1. Absorbance in the area up to 300 nm is mainly due to the nucleobases, whereas Cy and Sq absorption is manifested in the visible to near-IR area (500–700 nm). A key observation is the change in the shapes of the Cy and Sq spectra in the single strands (corresponding to the spectra of the monomer dyes) and those of the Cy–Cy and Cy–Sq assemblies in the hybrids. The development of a new, red-shifted absorption band of the ON1*ON2 hybrid compared to the monomer (668 vs. 648 nm) is characteristic of J-aggregation of the Cy dyes;13–15 a J-dimer in this case. In contrast, in the mixed assembly Cy–Sq (ON1*ON5) the major band is blue-shifted (591 nm) compared to the band of each monomer, revealing preferential H-type coupling.

Table 1 Spectroscopic properties of oligomers and hybrids (10 mM sodium phosphate buffer, pH = 7.4, 100 mM NaCl)

<table>
<thead>
<tr>
<th>Sample</th>
<th>λmax(Abs)/nm</th>
<th>fwhm*/cm−1</th>
<th>λmax(Em)/nm</th>
<th>ΦF</th>
</tr>
</thead>
<tbody>
<tr>
<td>ON1</td>
<td>255, 648</td>
<td>940</td>
<td>667</td>
<td>0.42b</td>
</tr>
<tr>
<td>ON2</td>
<td>260, 647</td>
<td>930</td>
<td>667</td>
<td>0.37b</td>
</tr>
<tr>
<td>ON5</td>
<td>259, 633</td>
<td>700</td>
<td>646</td>
<td>0.31b</td>
</tr>
<tr>
<td>ON1*ON2</td>
<td>257, 601, 668</td>
<td>—</td>
<td>668</td>
<td>0.039</td>
</tr>
<tr>
<td>ON1*ON5</td>
<td>258, 591, 631</td>
<td>—</td>
<td>648</td>
<td>0.041</td>
</tr>
</tbody>
</table>

† Electronic supplementary information (ESI) available: Synthetic and analytical details; additional spectra. See DOI: 10.1039/c3cc42103a

a fwhm – full width at half maximum. b λex = 600 nm. c λex = 625 nm.

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Fluorescence data show that either form of assembly results in a substantial decrease in the Cy signal (~90%, Table 1 and Fig. 1). The excitation spectrum (ESI† Fig. S4) has a maximum at 650 nm revealing that the remaining fluorescence signal of the ON1*ON2 hybrid originates from a monomer type chromophore. Variable temperature absorption spectra of single strand ON2 (ESI† Fig. S5) disclose no specific interactions of the cyanine with other parts of the same oligomer (negligible deviations in absorbance). A decrease in fluorescence is observed with rising temperature (70% reduction, 20 → 80 °C), which is in agreement with the thermochromic behaviour described for cyanines.16,17

The higher tendency of cyanines to form J-type assemblies15,17 compared to other organic molecules is well documented. H- and J-assembly of cyanines in crystals and extended aggregates on surfaces or in solution, including prolonged assemblies on DNA and other polymers as a template, has been reported.18–22 The precise composition of J-assemblies is often ill-defined and the number of units participating in coherent J-coupling is uncertain.15,23–25 The oligonucleotide-driven approach enables a molecularly defined constitution of the assembly. Spectroscopic changes distinctly show a transition between monomeric and dimeric forms of Cy molecules. The development of a J-band (∆λ = 20 nm) upon formation of a Cy-dimer is evident upon titration of ON1 to ON2 (Fig. 2). More accurately, assembly of Cy within the ON1*ON2 hybrid is accompanied by the development of two main bands consistent with Davydov exciton splitting21,22 of the monomer band (M, ∆λmax = 648 nm) into two levels, at lower (∆λmax = 668 nm, J-band) and higher (∆λmax = 601 nm, H-band) energies, which reveals an “oblique” orientation of the chromophores.3,26 This arrangement of the cyanines is crucial for the appearance of strong CD effects (see below).

The melting temperature (Tm) of the ON1*ON2 hybrid (67 °C, Fig. 2) is 4 °C lower than that of the unmodified duplex ON3*ON4 (71 °C). This is comparable to the effect of a nucleobase mismatch or an abasic site in a DNA duplex27 and in contrast to the effects of other modifications of the same motif, such as pyrene,28 PD19 or flourene.20 The latter modifications increase duplex stability due to stacking in a presumed face-to-face H-type fashion.

The most inspiring results came from circular dichroism (CD) experiments. The CD spectrum of the ON1*ON2 hybrid is shown in Fig. 3 (left). A very strong intensity and a robust, specific signal shape are manifested. Several Cotton effects are evident: 668 nm (Δε + 250), 633 (Δε − 50) and 601 nm (Δε − 80). CD effects are absent or negligible in the corresponding single strands, which is attributed to the flexibility of the single strands. Signal shape and intensity are functions of dipolar strength of the electronic transitions and the mutual arrangement of the interacting transition dipoles, i.e. they depend on both, energetic and geometric factors. Usually, an exciton coupled CD of H-assembled chromophores is of symmetrical shape with positive and negative couplings of comparable intensities; it can be reasonably described by considering the main electronic transition only.21,22 The complexity of the CD couplet in the ON1*ON2 hybrid is due to the J-type dimer formation which leads to a substantial increase in the oscillator strength of the low energy band and its narrow character. The simulation of the observed spectrum has the best fit when the monomer band (648 nm) gives an asymmetric exciton couplet (+668 nm, −633 nm, with amplitude A +300 and width W = 35 nm) and the other bands (601 and 556 nm) are (mono-directional) negative signals (see ESI† S9). The CD spectrum is asymmetric in shape with a maximum that corresponds to the ∆λmax of the J-band, and a crossing point at 645 nm, which is close to the ∆λmax of the monomer absorbance. Notably, the overall integral intensity (∫500–715nm) of the positive (+) signal is significantly enhanced compared to the negative (−) one, ∫500–715(+)/∫500–715(−) = 1.4. Non-equivalence in the intensities of exciton couplings of the main transition (∫625–715nm) is very pronounced and manifested in a 6-fold more intense positive couplet (∫645–715(+)/∫645–625(−) = 6.2; Fig. 3 and ESI† S9). Quantitative characterization of signal asymmetry is possible using g-factor values,2 which gives gCy/CyCy = 4.2 and gCy/ON1 = 2.6 (see ESI†).

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Fig. 1. Top: absorption (left) and fluorescence (right) spectra of single strands ON1 (red) and ON2 (black), and their hybrid ON1*ON2 (green); bottom: absorption and fluorescence spectra of ON1 (red), ON2 (black) and hybrid ON1*ON5 (green, 1.5 μM single strand conc., 10 mM phosphate, pH = 7.4, 100 mM NaCl).

Fig. 2. Left: development of exciton splitting (J- and H-bands) upon Cy–Cy assembly; right: thermal denaturation of ON1*ON2, cooling (−), heating (−−); (Tm = 67 °C; M: Cy monomer, D: Cy dimer); single strand conc. 1.5 μM, conditions: see Fig. 1.

Fig. 3. Left: CD spectra of the ON1*ON2 hybrid in the temperature range 10–80 °C; right: CD spectra of different assemblies (20 °C); single strand conc. 1.5 μM, conditions: see Fig. 1.
The Sq dye (Scheme 1) has similar optical properties to Cy (λ_{max} = 645 nm, ε_{Cy} = 250 000 M^{-1} cm^{-1}; λ_{maxSq} = 635 nm, ε_{Sq} = 258 000 M^{-1} cm^{-1}) and was used as a reference compound and a counterpart in coupling experiments with non-identical chromophores. Sq–Sq excitonic interactions in the corresponding DNA hybrids\(^{13}\) show a significantly more populated transition at higher energy (H-aggregation) and a symmetrical CD signal (Fig. 3, right). Non-degenerate interaction of Cy–Sq within ON1*ON5 also results in a higher fraction of H-type coupling (Fig. 3, right), and gives a CD of complex shape with an overall dependence of signal intensity at 668 nm.

The remarkably high sensitivity of the Cy chiroptical label is further demonstrated by the titration experiment shown in Fig. 4: a 7.4 × 10^{-8} M oligonucleotide concentration is easily detected in a standard 1 cm cell [see ES1]. In conclusion, using a derivative of the widely used chromophore Cy5, we have demonstrated the value of cyanine dyes as chiroptical reporters. Cy–Cy interaction leads to intense CD effects in a DNA duplex. The origin of the high signal intensity resides in the formation of a J-type dimeric assembly that exhibits an asymmetric exciton coupled CD signal with a significant enhancement of one of the coupllets. This type of aggregation is well known for cyanine dyes,\(^{16–20}\) however, to the best of our knowledge, was not applied so far in the design of chiroptical labels using a monomer-to-dimer switch.

Notes and references


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